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FUNGAL INFECTIONS IN JOS: A 9-YEAR STUDY

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The prevalent mycoses and their aetiologic agents were studied in 1,287 patients in Jos and environs. 917 (71.3%) of the study population were infected. A male preponderance was observed and the age groups 11-20 and 21 -30 were predominantly infected. The most frequently isolated aetiologic agent was Candida spp and the commonest site of infection was the trunk. Unusual dissemination of lesions caused by Trichosporon beigelii infections was observed. The seasonal occurrence of mycoses and the effect on prevailing aetiologic fungi was studied.

INTRODUCTION

Fungal infections have, in the recent past, received greater recognition from various parts of the world, especially with the of several fungal upsurge pathogens. This upsurge has been attributed to factors such immunodeficiency, the use invasive devices or procedures, indiscriminate administration of broad-spectrum antimicrobial agents, which make the individual susceptible to even the otherwise non-pathogenic fungi (1-4). Some persons are at increased risk of susceptibility to infection because of their geographical location, level of health care delivery ignorance (5-6).

In Nigeria, the most commonly encountered mycoses include superficial and cutaneous infections, but very ew reports are available from some parts of the country on the epidemiology of these infections. Focus in the

Northern Nigeria on mycoses has been on occupational groups, particularly on the Jos Plateau where mining is a major occupation (7).

The present report highlights the types and some aspects of epidemiology of mycoses in Jos, Plateau State, in the past 9 years.

MATERIALS AND METHODS

The sample population was made up of 1,287 patients, 690 males and 597 females, referred to the Mycology Laboratory of the Medical Microbiology Department of the University of Jos, from the Jos University Teaching Hospital, clinics. private the **primary** healthcare units within Jos and adjoining local settlements, between January 1991 to December 1999.

Personal data on sex, age, occupation, domicile, recreational activities and salient features of the lesions were recorded. Samples of

skin scrapings, hair, nail clippings, vaginal, wound and ear swabs, diarrhoeic stool and biopsy specimens were analyzed.

A portion of each specimen was examined under the light microscope, in 20% potassium hydroxide mount. Cultures of the specimens were also made on slopes of Sabouraud's Dextrose supplemented chloramphenicol (0.05mg/ml) and cycloheximide (0.5 mg/ml). Cultures were incubated at room temperature (25-30°C) for one to six weeks, examined regularly for fungal growth. The yeast isolates were further subjected to a battery of biochemical and physiological tests to identify them to specific levels. Isolates identification was achieved by comparison with standard descriptions in books and manuals (8-10).

RESULTS

Of the 1,287 patients, 917 (71.3%) had fungal infections (Table 1). A mean yearly increase in incidence rate of 7.04% was recorded. Of the positive cases, 502 (54.7%) were males while 415 (45.3%) were females. There was a male preponderance of infection, Of 690 generally. the male 502 (72.9%)subjects, infected while 415 (69.3%) of the 597 females were infected. The predominantly infected age groups were the 21 - 30 (24.0%) and 11 - 20 (20.0%) (Fig.1). The overall number of infected adults was higher than children.

The fungal lesions were found in various parts of the body, most commonly the trunk and multiple sites (Fig.2). Only four samples of tissue biopsy were analyzed and all yielded positive fungus cultures.

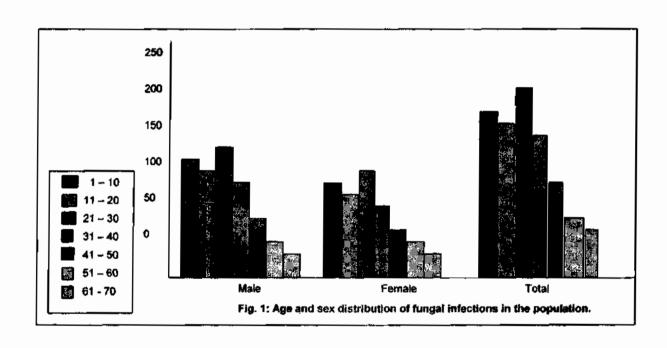
The most frequently encountered aetiologic agents over the nine year period are presented in Fig.3. Yeasts and yeast-like fungi predominated followed bv dermatophytes with 444 (48.7%) isolates. 392 (42.7%)respectively. Candida spp (69.0%) and Trichophyton mentagrophytes (41.9%) out numbered the yeast dermatophyte isolates. respectively, in occurrence. Of the isolates. C. Candida albicans predominated.

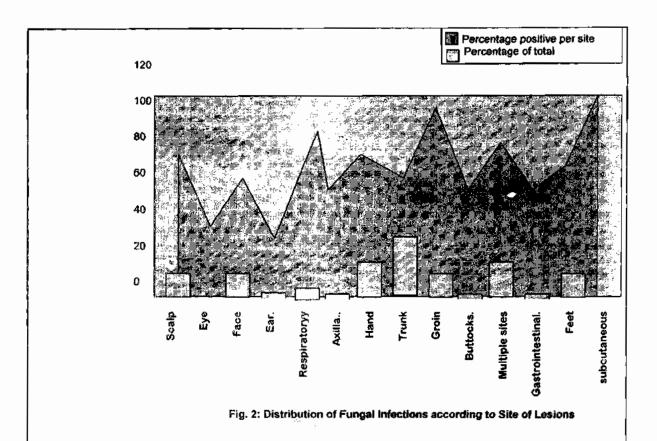
Aspergillus fumigatus formed majority of the hyaline hyphomycetes and more than 70% was isolated from respiratory infections. Wangiella dermatitidis was the predominant dematiaceous fungus with 11 (31.4%) of the 35 isolates.

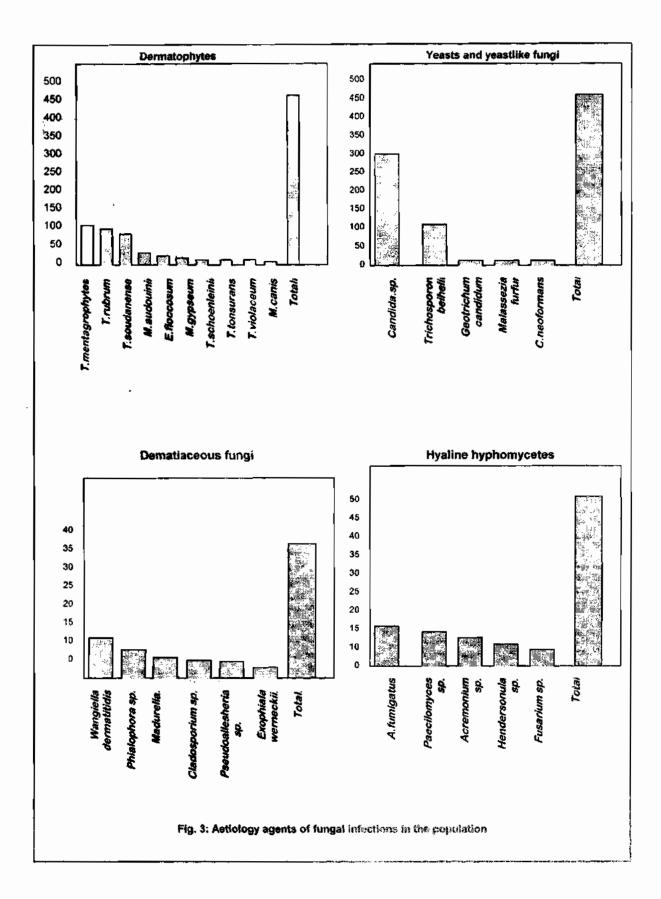
The mean monthly rate of infection in the population is shown in Fig. 4, the highest being in August (14.2%) and the lowest in December (4%). A striking observation here was the high occurrence of *Trichosporon beigelii* in the population, the lesions of which were frequently generalized.

Table 1: Incidence of Mycoses in the population

Year	No. sampled	No.	infected	Total	%
		Male	Female		
1991	232	51	39	90	38.8
1992	98	23	27	50	51.0
1993	83	30	23	53	63.8
1994	83	32	16	47	56.6
1995	112	57	34	91	81.2
1996	123	51	51	102	82.9
1997	197	83	90	173	87.3
1998	179	90	66	157	87.1
1999	180	85	69	154	85.5
Total	1287	502	415	917	71.3









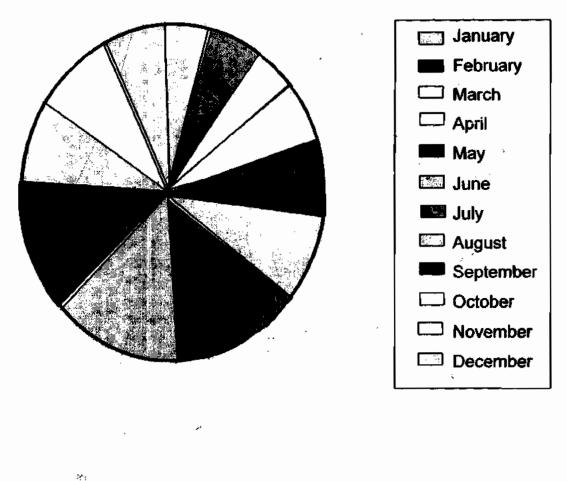


Fig. 4: Monthly Incidence of Mycoses in the Population

DISCUSSION

This study has revealed the increasing prominence of fungal infections as have documented from other parts of the world (1-4). Over the years a significant increase was observed in the rate of occurrence of these infections. Some reports (2,3) have revealed that more stressors and immunocompromise factors of have contributed a lot to this trend. Although the present study did not investigate such factors, they may not be far from affecting present population. the males were infected than females, similar to observations elsewhere within the country (12). Reports from parts of the world, however suggest an equal male - female ratio. This might not be unrelated to the life style of the citizens of Jos, and other parts of Nigeria where males are more exposed to outdoor sources of fungi such as recreational areas and engagement in mining activities.

The predominantly infected age groups, i.e. 21-30 and 11-20 years, include the most active in the populations both at work and play, hence their closer contact with sources of fungal pathogens. Also some these youths are completely ignorant of measures for prevention and control of fungal infections, and the living conditions in boarding schools and hostels where majority

resides. favour the spread mycotic infections. Positive lesions were obtained from various parts of he body, especially the trunk. This was most commonly ringworm corporis), infections (tinea agreement with the findings of Alsogar et al (10) in Saudi Arabia, and Egere and Gugnani, in Eastern Nigeria. Little is done laboratory on diagnosis of systemic mycoses because of limited resources. but the few tissue biopsies analyzed in this study yielded positive results. This is an important need for consideration by the authorities concerned.

Yeasts were predominant as causative agents of mycoses in this study. Candida species were the most frequently isolated, contrary to reports in Eastern Nigeria and some tropical countries. Trichosporon beigelii, as a single species was isolated in considerably high numbers. Α number investigators have highlighted the increasing importance of this organism. both in immunosuppressed and immuno-competent individuals. The generalized lesions of this organism in a significant number of apparently immunocompetent subjects in this study seems to buttress the reports of other workers elsewhere (1,13).

The predominant dermatophyte species isolated was *Trichophyton mentagrophytes*, in contrast to the reports from the

southern parts of the country, and other places (10,12). Also the aetiologic agents in this study are at variance with those obtained in a number of reports. There could climatic influence on actiology of mycoses, as suggested by some workers. The month of August recorded the highest number of infections, contrary to other reports (10) where the hot and dusty weather was suggested to promote spread of mycoses. In Jos the rains are relatively heavy in August and the atmosphere dust free. almost Probably because of the warmth and moisture obtained in this period. the development of fungal pathogens is favoured. The lowest infection rate was recorded in December in this study. This is one of the coldest months in the year here, exposure to outdoor like activities farming swimming is minimal. This report seems to be the first to consider effects of seasonal variation on mycoses and their actiologies in Northern Nigeria.

ACKNOWLEDGEMENT

We thank the Almighty God for the success of this work. Our sincere appreciation also goes to Mr. Yohanna Mato and Mrs Satya Noma, both laboratory assistants in the Mycology laboratory. Mrs. P Anele and Mr. Solomon Danladi for secretarial assistance.

REFERENCES

- 1. Ellis DH. Opportunisic Fungi Pose Growing Threat. Mycology Forum. 1995; **2(1)**: 2-7.
- Jarvis WA. Epidemiology of Nosocmial Fungal Infections, with Emphasis on Candida species. Clin. Infect. Dis. 1995; 20: 1526-1530.
- Fridkin SK, Jarvis WR.
 Epidemiology of Nosocomial
 Fungal Infections
 Clin. Microbiol. Rev. 1996;
 9(4): 499-511.
- Elewski BE, Greer DL. Hendersonula toruloidea and Scytallidium hyalinum. Review and update. Arch-Dematol. 1991; 127(7): 1041-1044.
- 5. McCouy CW, Boucias DG. Selection of Beauvaria bassiana pathotypesas potential microbial control agents of soil-inhabiting citrus weavils. Mems. Inst. Oswalso Cruz. 1989; 84: 75-80.
- 6. Ajello L. Ecology and epidemiology of hydrophillic infectious fungi and of medical parafungi mycologial importance: A new category of pathogens. In: Collier L, Balows A, Sussman M (eds). Topley & Wilson's Microbiology and Microbial infections. 9th edition. Arnold, London, 1998: 67-73.

- 7. Ayanbimpe GM, Bello CSS, Gugnani HC. The aetiological agents of superficial cutaneous in Jos, Plateau State of Nigeria. Mykoses. 1995; 38: 235-237.
- 8. Larone DH. Medically important fungi: a Guide to Identification 3rd edition. ASM Press, Washington, DC. 1995: 274.
- Lennette EH, Balows A, Hausler WJ Jr., Shadomy HJ. Manual of Clinical Microbiology. 4th edition. ASM, Washington, DC, 1985: 500-594.

- Al-Sogair SM, Moawad MK, Al-Humaidan YM. Fungal infections as a cause of disease in the Eastern province of Saudi Arabia: Prevailing fungi and pattern of infection. Mycoses. 1991;
 34: 333-337.
- 11. Frey D, Oldfield RJ, Bridger RC. A Colour Atlas of Pathogenic Fungi. Wolfe Medical Publication Limited, Holland. 1979: 168.
- 12. Egere JU, Gugnani HC. Etiology of Dermatophytosis in Eastern Nigeria: A7 Year Study. Mykosen .1982; 25(4): 178-181.
- 13. Kulfas LL. Don't overlook T. beigelii in disseminated fungaemia. Current Anti-infective. Therapy. 1994; 14 (1):7.

INCIDENCE OF DERMATOPHYTE INFECTIONS AMONGST SOME OCCUPATIONAL AND SELECT GROUPS IN JOS.

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Fifty-nine Agro farm workers, 265 inmates from Jos main prison, 60 hair weavers and 40 car washers were examined in Jos for dermatophyte infections. Dermatophyte isolates included *Trichophyton* and *Microsporum species*. The highest infection rate of 75% was recorded among the farm workers with toeweb infections due to *T. mentagrophytes* and *M. canis*. 25% of the other infections were from the groin region due to *T. tonsurans* and *T. mentagrophytes*. Fifty-nine percent of infection by *T. mentagrophytes* in prison inmates was restricted to the groin region. Other species of fungi isolated from the inmates were *Trichosporon cutaneum* and *Aspergillus niger*. Among the car washers tinea manum, ringworm of the hand, and interdigital infections due to *T.mentagrophytes* were observed. There was no visible infection among either the female hair weavers or the female prison inmates.

Keywords: Dermatophytosis, T. mentagrophytes, T. tonsurans, M. canis, Tinea pedis, Tinea cruris,

INTRODUCTION

Cutaneous fungal infections caused by dermatophytes and commonly known as ringworm infection have been known to be occupationally acquired. reported by Philpot (1) and Ogbonna et al (2). Such infections have been found to be common among anthropophilic or human species. The characteristic among such species is that they are associated with community living and the spread of such infections require close human contact. This might occur as a result communal use of certain facilities such as baths (3), combs, brushes and articles of clothing Incidence of ringworm infections through such sources could then be found among workers who share working boots and overalls such as coal miners, factory workers, camp and prison

inmates. It has even been reported in families by English (5) where overcrowding and concentrated use of common facilities result in infections. Tinea pedis (ringworm of the toeweb) could be spread through desquamated scales adhering to moist surfaces such as bathrooms or changing room floors (6).

Zoonosis is found in workers whose occupation lead to contact with infected animals or through contaminated working equipment. Such workers could include herdsmen (2). Hairdressers and barbers also run the risk of infection through infected hair and scalp of their customers. The combined effect of host factors. moisture. warmth, duration exposure to pathogen and other environmental stresses have been found by Gentles and Holmes (7), to

play important role in the initiation and establishment inflection within a community.

This study was undertaken to report on the first survey of incidence of dermatophyte infection and environment effects among selected occupational groups in the Jos Plateau of Nigeria.

MATERIALS AND METHODS Study population

The workers. were agricultural farm workers from Zalaki farm about 25 km from Jos metropolis, car washers and hair from within Jos. weavers addition, inmates of Jos main prison were also examined. The farm workers were selected due to their proximity and contact with the animals as possible source of infection. The hair weavers were examined owing to their possible handling of hairs of their infected customers, while the car washers could be exposed to particles of soil adhering to dirty cars, which could contain spores of dermatophytes. The prison inmates were selected because they are exposed to over-crowding and poor environmental sanitation conditions. All sampling carried out between September and November 1991.

The climate of Jos Plateau is peculiar to itself in Nigeria. There is Savannah woodland vegetation and temperate climate quite different from the hot humid parts of other parts of Northern Nigeria. The geographical location confers on it Mediterranean climate conditions with dry cold weather and very low humidity.

Specimen collection and processing

Fifty-nine animal farm workers whose ages were between 25-45 years were sampled. Their bodies were thoroughly examined for the presence of cutaneous dermatophyte infections. Lesions remotely resembling fungal infections were cleaned with 70% alcohol and scrapings were then collected from affected body areas with the aid of sterile blade into sterile paper packets, which were then transported to the laboratory for further fungal analysis. scrapings were cultured on Sabouraud Dextrose Agar supplemented with 0.05 mg/ml chloramphenicol and 0.5 mg/ml cycloheximide in order to suppress growth of bacterial contaminants. Cows on the farm were examined as possible source of human infection among the farm workers. Tufts of hair were collected from infected in paper wrappers cultured for the presence keratinophilic fungi using the "hair baiting" technique as described by Mckenzie. This involved the use of sterile petri dishes, which were half-filled with soil that have been sterilized by autoclaving at 1210C

for 30 minutes for three successive times. The infected hairs from the cows were then sprinkled on the sterilized soil after the soil has been sufficiently moistened with sterile distilled water. The petri dishes were incubated at room temperature in the dark by placing them in closed cupboard.

In Jos main prison, 265 male inmates whose ages range from 18-70 years were thoroughly examined. Scrapings were collected from the affected parts of the body after cleaning the area with 70% alcohol. The scrapings were plated out on Sabouraud Dextrose Agar containing appropriate antibiotics as in the case of the farm workers.

In the case of the 60 female weavers and 40 car washers whose ages range from 30-45 years respectively, examination was only restricted to areas of the body not covered by clothing due to the outdoor nature of their jobs. Skin scrapings interdigital collected from infections of the car washers were subjected to the same treatment for the isolation of dermatophyte fungi as described for farm worker above.

The resultant culture plates from all the sampling except those cultured on sterilized soil were incubated at 37°C and examined on daily basis for the presence of dermatophytes.

dermatophytes that appeared mainly after 10-14 days subjected to series subculturing until pure cultures were obtained. The isolates were then identified with the aid of the microscope and existing cultures. References were made to Rebel and Taplin (8); Rippon (9); Campbell and Stewart Wherever necessary, clides cultures were made. Analysis of variance was used in the interpretation of the results.

RESULTS

The dermatophyte from the various occupational groups as Table Ŧ included shown Trichophyton and **Microsporum** species. Trichophyton tonsurans observed in toe-web infection among the farm workers was also isolated from one of the cows. Among the prison inmates, out of a total of 41 positive cultures, 9 (21.95%) were due to infection caused by T. mentagrophytes and 7 (17.07%) cutaneum. by *T*. infections were recorded among the male inmates, but none in their female counterparts and female hairdressers. In the case of the car washers, only interdigital infections caused by T. mentagrophytes were recorded.

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				SITES	OF IN	FECTI	<u>ON</u>				
Organisms	GROIN		101	E-ME8	4	HAIL		FOLD OF BUTTOCKS		SHOULDERS	
	P.1	F.W	P.J	F.W	P.I	F.W	P,I	F.W	1.9	F.W	
Trickophyton montagrophytes	2(4,87%)	1(8.33%)	0	3(35.0%)	0	ø	0	0	o	0	6
Triphophyton tosurans	0	2(15.66%)	0	2(16.86%)	0	0	٠	0	0	¢	6
Microsporon canis	0	a	•	2(16.86%)	٥	9	•	e	0	G	2
Trichesporon cutaneum	7(17.07%)	o	G	٥	٥	9	Ð	0	0	0	7
Aspendilles niger	3(7.31%)	0	c	0	0	0	•	•	0	จ	3
<u>Рітураро</u> гит	o	•	a	0	Đ	0	0	0	5(12.19%)	o	6
Candida	2(29.26%)	0	G	٥	a	0	٥	0	0	a	2
Mucor sp	0	۵	đ	2(16.66%)	q	Q	•	¢	0	C	2
<u>Dermatitis</u>	0	۵	e	9	0	0	10(24.39%)	0	ů.	0	10
TOTAL	14	3	¢	9	0	0	10		5	đ	41(1001
KEY: P.S		Prison famel	as,								
F.W. =		Farm Worke	_								

Table 2: Analysis of Variance for data on distribution of dermatophytes and related fungi amongst Zalaki Farm Workers

ANOVA Source of Variations SS df P.Value Fcrit MS Rows 0.800 3 0.267 2.285 0.357 3.490 15.300 Columns 4 3.825 17.00 0.000 3.259 **Error** 2.700 0.225 12 TOTAL 18.800 19 Multiple Range Value = 0.129

Table 3: Analysis of Variance for data on distribution of dermatophyte and related fungi amongst Jos Prison Inmates.

ANOVA									
Source of Variations									
	SS ,	df	MS F		P.Value	Fcrit			
Rows	18.971	6	3,162 0.	377	0.886	2.508			
Columns	66.971	4	16.743 1.	999	0.127	2.776			
Error	201.029	24	8.376						
TOTAL	286.97 1	34							
Multiple Range Value =	3.419565								

Pg. 14______

DISCUSSION

The results obtained have that infection with shown species dermatophyte is occupational hazard. This agrees with previous findings from other investigators (2,6,11). Continous exposure of the farm workers to infected cows could result in their being infected with dermatophyte species that are zoontic. The farm workers at Zalaki had 75.0% rate interdigital infections. conditions of service could have enhanced this high infection rate among the workers. The workers were observed to share pithead shower facilities at the end each day's work. Dermatophytes have been isolated from floors pithead baths, swimming pools and changing rooms (3). English (5) has also observed that fungi have been known to survive for months in shed skin scales even after serveral laundering of wool. Also the use of alkaline soap could have enhanced the spread of infection, among the community. survey of a naval In establishment in Cranston England, Broughton (6) found that tinea pedis was particularly seamen common among experiencing much desquamation due to the use of alkaline soap. It would therefore appear that the communal use of shower facilities, the use opf common overall and could soap have all played

significant roles in the spread of ringworm infections amongst those workers. Moreover, the use of heavy footwear in damp hot weather condition may also have provided suitable environment for their infection (11).

The dermatophyte species, T. mentagrophytes and T. tonsurans isolated form the workers known anthropophilic species not restricted to any geographic region. However. the isolation tonsurans in one of the infected cows is worthy of note since this anthropophilic species has not been listed as a zoophilic and neither has it been isolated an from an animal. Its mode of transmission though uncertain could have been from man to animal or vice versa. Infections from infected animal have been known to be acquired building or equipment contaminated by contact infected animals, while sources of infection of farm animals could include barns, fences and soil. Infections among the prisoners were mainly in the groin accounting for 58.53% of total infections. Tinea cruris has been considered an infection of sedentary people and to be more common among males than females. This could account for the non-isolation of species of these fungi from the Jos female prison inmates (7.8%) in view of the prevailing environmental conditions of over-crowding, poor personal hygienic and nutritional conditions. The dry low humidity of the Jos Plateau could have contributed to the low infection rate, as it reduces sweating, which reduces ringworm infection to the barest minimum. This be compared to can situations in India where Tinea corporis and Tinea pedis uncommon but Tinea particularly of the lower covered waistline and groin assumes particular clinical significance due to the not humid climate of the region (12).

Ringworm infections were not recorded among the female hair weavers. The absence of infection among them in spite of their handling and association with people's hair could be due to the fact that the hair weavers wash their hands after attending to each customer, coupled with the age range of their customers who were found to be adults between the age range of 20-60 years. Tinea capitis is an infection that is common among children, the highest incidence occurring between the ages 4-14 years. The risk of contracting infections from other parts of the client's body is reduced, as bodily contact is minimal.

The constant exposure of car washers to water and use of different types of detergent must have been predisposing factors to their tinea pedis and tinea manum infection.

REFERENCES

- 1. Philpot CM. Some aspects of the epidemiology of tinea. *Mycopathologia*. 1977; **62**:3-13.
- Ogbonna CIC, Enweani IB,
 Ogueri SC. The distribution of ringworm infections amongst Nigerian nomadic Fulani herdsmen.
 Mycopathologia. 1986; 96: 101-106.
- 3. Ajello L, Getz ME. Recovery of dermatophytes from shoes and shower stalls. J. Invest. Dermato. 1954; 22:17-21.
- 4. Clayton Y, Midgeley G. A new approach to the investigations of scalp ringworm in London School Children. J. Clin. Path. 1968; 21:291.
- 5. English MP. Trichophyton rubrum infections in families. Brit. Med. J. 1957;11:744-746.
- 6. Broughton RH. Reinfection from socks and shoes in tinea pedis. *Brit. J. Dermato*. 1955; **67**: 249-254.

- 7. Gentles JC, Holmes JG. Feet ringworm in coal miners.

 Brit. J. Industr. Med.
 1957; 14:22- 29.
- 8. Rebel C. Taplin D. Dermatophytes: Their and recognition identification. Univ. of Miami Press. Florida, U.S.A Coral Gables: 1970.
- Rippon JW. Medical Mycology. The pathogenic fungi and the pathogenic Actinomycetes. 2nd edition, W.B. Saunders Company, Philadelphia, 1974.
- Campbell M, Stewart J. The Medical Mycology Handbook. John Whiley and Sons incorporated, New York, 1980.
- 11. Allen A, Taplin D. Epidermis Trichophyton entagrophytes infections in servicemen; source of infection, role of environment, host factors and susceptibility. JAMA. 1973; 226: 864-867.

- 12. Desai SC. Epidemicity and clinical features of Trichophyton rubrum in the Tropics. Dermato.
 Int. 1966; 5: 222-224.
- 13. Randhawa HU. An epizootic of dermatophytosis caused by *Trichophytons* simii in poult. Sabouraudia. 1973; 11:1-3.
- 14. Klobuscicky M, Buchvald J. Workmen Keratinophiler Pilze cut unbeleten organischen material in der umgebung von Trichopyton verrucosum infiziertens kalbern. Mykosen. 1976; 19: 124-127.

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INHIBITION OF SWARMING BY UREA AND ITS DIAGNOSTIC IMPLICATIONS AMONG UROPATHOGENIC PROTEUS SPECIES FROM LAGOS, NIGERIA

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The anti-swarming property of urea and effects on antibiotic susceptibility among 52 uropathogenic *Proteus* strains from Lagos, Nigeria were investigated. Urea caused a reduction in swarming and number of swarmed cells at 0.5% (n = 42, DOCZ = 15.5mm), 0.75% (n= 24, DOCZ = 10.7mm), 1% (n = 17, DOCZ = 3.4mm) and 1.25% (n = 8, DOCZ = 1.7mm). Compared to DOCZ obtained at 0.5% urea, the further reduction in DOCZ at other urea concentrations was found to be significant (p < 0.05). Urea at less than 0.75% allowed identification of *E. coli, K. pneumoniae and S. saprophyticus* in mixed cultures containing *Proteus spp,* while colonies of *Pseudomonas aeruginosa* were distinctly identified at 1% urea with swarming restrained at 1.25% urea. At 1.25% urea, antibiotic susceptibility testing by agar diffusion method revealed significant increase and decrease in the number of *Proteus* strains that showed resistance to amoxicillin and nitrofurantoin. Compared with the control, significant increases in the MICs of gentamicin or nitrofurantoin and streptomycin were found at ≥ 0.5% and ≥ 0.75% urea respectively (p<0.05). The identification of extended spectrum beta lactamases (ESBL) producing strains were unaffected by urea. This study has demonstrated urea induced swarming inhibition of uropathogenic *Proteus in vitro*. However, results suggest the use of urea with great caution in diagnostic practices for optimal clinical and public health benefits in Nigeria.

<u>Keywords</u>: Antibiotic susceptibility, anti-swarming, urea, uropathogenic proteus, Nigeria, DOCZ (= Diameter of outermost colony zone mean value)

INTRODUCTION

Proteus are spp Gramnegative facultative anaerobic rods of tremendous clinical and public health importance particularly in developing countries (1). These bacteria are frequently implicated as aetiologic agents of urinary tract infections (UTI), which may kidney damage lead to complicate pregnancies untreated (2, 3). In Nigeria, several hospital and community based studies have put the isolation rates of Proteus spp at 3-8%, with Proteus mirabilis and P. vulgaris as predominant nosocomial pathogens among patients with indwelling catheters,

benign prostatic hypertrophy, and vesicoureteral reflux (4, 5, 6). Of great concern is the mortality rate report of 17.1% in African neonates with UTIs that are untraceable by the radiologic examination of the urinary tract (7).

the One of unique characteristic features of Proteus spp on tolerable culture media is the ability to spread and form a thin film with distinct colonial surfaces. This zones on agar phenomenon called swarming, involves the differentiation of vegetative motile to hyperflagellated-elongated cells coordinated capable of and

population concerted mass migration (8). The susceptibility of catheterized patients to Proteus associated UTI has been demonstrated in vitro using Foley catherters (9). Swarming has also been shown to be one of the requirements for the colonization of urothelial cells in vivo and in (io. 11). of vitro One drawbacks of Proteus swarming in the laboratory is the inherent difficulty in the detection of other polymicrobial pathogens in infection cases. Fons et al (12), reported difficulty the in distinguishing colonies of Pseudomonas aeruginosa among P. mirabilis swarm cells on agar plates.

In routine diagnostic laboratories, the use of nutrient and blood agar media for Proteus culture storage and antibiotic susceptibility testing, as practiced developing countries, compromise purity of stocks for genetic studies and many cases of polymicrobial infections involving Proteus may not be noticeable. This may hinder the use of drug combination to implement effective clinical cure, provide a reason for the incidence recurrent bacteruria in treated patients and promote antibiotic resistance of 'silent' organisms. False antibiotic susceptibility outcome öf pathogens in vitro also contributes to the spread of drug resistant strains in a community and promotes clinical failure.

P-nitrophenylglycerol Urea. (PNPG), and activated charcoal have experimentally been demonstrated to possess anti-swarming properties for routine recommended (12.13.14).laboratory usage However the use of PNPG in culture media for antibiotic susceptibility testing has been queried (15). In Nigeria, urea is commonly used in culture media designed for the identification of pathogens of UTIs spp (4, 5). including *Proteus* However, reports have been silent on Proteus swarming prevention possibilities and consequences on antibiotic susceptibility outcome. This study investigated the clinical importance associated with the use of urea in Proteus identification media in terms of pattern of swarming inhibition, the effect on antibiotic susceptibility and extended spectrum beta lactamase (ESBL) classification.

MATERIALS AND METHODS Bacterial strains

Fifty-two *Proteus spp* isolated from randomly selected 408 mid stream urine samples of in-patients and outpatients at different clinics and hospitals in Lagos, were used in this study. The isolates were identified on McConkey, Blood agar and composite media using criteria which included non-lactose fermentation, swarming ability,

and phenylalanine urease deaminase production (16). For mixed culture assay, pure strains P.aeruainosa. Klebsiella αf pneumoniae. Staphulococcus saprophyticus and E. coli were obtained from the Microbiology Laboratory of the Nigerian Research Institute of Medical (NIMR), Lagos.

Swarming inhibition assay

Nutrient agar plates containing 5% sheep erythrocytes supplemented with without 0.5-1.25% urea (Sigma, USA) were used to grow the selected Proteus strains. A loopful of standardized inoculum (2 x 104 CFU/spot) of each strain was concentrically inoculated at the center of the agar and incubated at 37°C for 24 hours under aerobic conditions. Urea negative plates were used as controls. The swarming profile of P. mirabilis was examined in ATCC49565 parallel with those of the test organisms. The degree swarming was measured as the diameter of the outermost colonial zone (DOCZ). DOCZs were interpreted as mean + standard deviation. to allow statistical deductions using student's t- test and chi-square analysis. P value less than 0.05 was indicated as significant.

Mixed culture assay

Four swarmed isolates of the tested *Proteus spp* were rapidly

selected and suspended in Mueller-Hinton broth containing at least any two of P.aeruginosa, Klebsiella Staphylococcus pneumoniae. saprophyticus and E. coli. resulting mixed cultures were then used to inoculate Blood supplemented (0.5 with urea 1.25%). Mixed culture plates containing a reference swarming strain of P. mirabilis ATCC49565 was also examined. Mixed culture plates without urea also served as control.

Antibiotic susceptibility testing

The response of untreated Proteus isolates and urea survivors to 13 antibiotics used in Nigeria was investigated by agar diffusion method according to Bauer et al (17). The antibiotics from Abtek Biological Limited, Liverpool, England mounted on inoculated plates were tetracycline 10 µg, ampicillin 25 µg, amoxicillin 25 µg, co-amoxyclav 30 ug, cotrimoxazole μg, nitrofurantoin 200 ceftazidime 30 µg, ceftriazone 30 nalidixic acid 30 μg, μg, streptomycin 25 µg, gentamicin 10 ciprofloxacin μg, 10 μg, ofloxacin 5 µg. Diameters of zones of inhibition were measured to the nearest millimeters and values were interpreted resistant as susceptible based on comparison with antibiotic susceptibility profile of E. coli ATCC 25922, a drug sensitive strain.

Determination of minimum inhibitory concentration (MIC) and selection of Extended Spectrum Beta Lactamase (ESBL) producing strains

The MICs of the selected antibiotics were determined by a microdilution method. strains from urea containing and deficient plates were first grown overnight on cysteine electrolyte deficient (CLED) agar at 37°C under aerobic condition. Four distinct colonies of each strain were then subcultured in 4 ml of IsoSensitest broth (Oxoid, UK) and incubated at 37°C to attain a turbidity that was adjusted to 106 CFU/ml with the broth. Stock solutions of the antibiotics were prepared fresh at 128 mg/L for ampicillin, co-amoxiclav. amoxycillin. nitrofurantoin, streptomycin and tetracycline; 64 mg/L for cotrimoxazole: 32 mg/L for nalidixic. gentamicin, acid, ceftazidime and ceftriaxone; and 4 for ciprofloxacin mg/L ofloxacin. In a 12 by 8 wells tray, 50 uL of antibiotic solution occupied the first two rows and subsequently double diluted to fill the remaining rows except the 12th row, which served as a positive control. 50 µl of inoculum at 5 x 105 CFU/well were then added to the wells in increasing order of concentrations. antibiotic susceptibility Antibiotic of

control strain of P. mirabilis ATCC 49565 was examined in parallel with test organisms. All the plates were sealed and incubated at 37°C for 24 hours. Growth was assessed as turbidity observed transillumination. The MIC of each antibiotic was defined as the lowest concentration that inhibits growth. Interpretation of MICs as resistant or susceptible was in line with NCCLS break points οſ antibiotics tested The {18ì. significance of the mean MIC value differences between urea treated Proteus and untreated isolates was evaluated statistically. Extended beta-lactamase producing strains were selected as those with ceftazidime: ceftazidime-clavulanate ratio greater than or equal to 16 according to Livermore and Yuan (19). Values of 8 were regarded as indeterminate.

RESULTS

Survival and variations in the ability of urea at 0.5- 1.25% to refrain swarming of *Proteus* isolates were highlighted in Figure 1. All the *Proteus* strains studied survived urea at 0.5-1.25% concentrations. At 0.5% urea, 42 out of the 52 isolates swarmed and produced an average diameter of outermost colony zone (DOCZ) value of 15.5 mm. At 0.75% urea, 24 isolates swarmed producing mean DOCZ value of 10.7 mm. The number of swarmed *Proteus* further decreased

from 17 to 8 following 1-1.25% urea treatments. Mean DOCZ values of 3.4 mm and 1.7 mm were produced respectively. Urea at 0.5% was observed to allow distinct identification of E. coli and K. pneumoniae in mixed culture Staphylococcus assay. saprophyticus colonies were identified at 0.75% urea while at 1%. 0.75and Pseudomonas aeruginosa was identified, with swarming inhibited (Table 1).

However, the number of cells that displayed resistance to amoxicillin, gentamicin, nitrofurantoin and ofloxacin by disk diffusion method at 1-1.25% urea differed (Table 2) and those of amoxicillin and nitrofurantoin

statistically significant were (p<0.05). Table 3 summarized data for MICs of the 13 antibiotics tested. Significant increases in the αf nitrofurantoin gentamicin and streptomycin were obtained at ≥ 0.5 and $\geq 0.75\%$ urea respectively (p<0.05). Furthermore, two Proteus strains were identified extended beta lactamases producers in plates devoid of urea and those containing 0.5 - 1.25% However, no isolate urea. identified as indeterminate ESBL production among the strains cultured without urea, whereas, between one and two indeterminate identifications were recorded among the urea treated strains (Table 4).

Table 1: Isolates identification from mixed cultures containing swarmed *Proteus* strains.

Mixed culture assay	Isolate identification scheme	
1.	Proteus strains	_
2.	Proteus strains, E. coli, K. pneumoniae,	
3.	Proteus strains, E. coli, K. pneumoniae,	
	S. saprophyticus, P. aeruginosa	
4.	Proteus strains, E. coli, K. pneumoniae,	
	S. saprophyticus, P. aeruginosab	
5.	Proteus strains, E. coli, K. pneumoniae,	-
	S. saprophyticus, P. aeruginosac.	

Keys: 1 = Assay without urea; 2 = Assay containing 0.5% urea; 3 = Assay containing 0.75% urea; 4 = Assay containing 1% urea; 5 = Assay containing 1.25% urea. a, P. aeruginosa not distinctly identified; b, P. aeruginosa swarmed cells identified distinctly; c, P. aeruginosa identified without swarming.

Table 2: Resistance to antibiotics in the presence and absence of urea by the 52 Proteus strains using disk diffusion method.

E	a	_		
40.43	-	b	c	đ
n (%)	n (%)	n (%)	n (%)	n (%)
32 (61.5)	32 (61.5)	32 (61.5)	32 (61.5)	32 (61.5)
31 (59.6)	31 (59.6)	31 (59.6)	31 (59.6)	36 (69.2)*
29 (55.8)	29 (55.8)	28 (53.8)	28 (53.8)	28 (53.8)
48 (92.3)	48 (92.3)	48 (92.3)	48 (92.3)	46 (88.6)
3 (5.8)	3 (5.8)	3 (5.8)	3 (5.8)	3 (5.8)
3 (5.8)	3 (5.8)	3 (5.8)	3 (5.8)	3 (5.8)
0 (0)	0 (0)	0 (0)	0 (0)	0 (0)
2 (3.8)	2 (3.8)	5 (9.6)	5 (9.6)	7 (13.5)
2 (3.8)	2 (3.8)	2 (3.8)	2 (3.8)	2 (3.8)
7 (13.5)	7 (13.5)	, .	, ,	4 (7.7)*
0 (0)	0 (0)	, ,	, ,	0 (0)
34 (65.4)	. ,			32 (61.5)
5Q (96.2)	50 (96.2)		, ,	50 (96.2)
	n (%) 32 (61.5) 31 (59.6) 29 (55.8) 48 (92.3) 3 (5.8) 3 (5.8) 0 (0) 2 (3.8) 2 (3.8) 7 (13.5) 0 (0) 34 (65.4)	n (%) 32 (61.5) 32 (61.5) 31 (59.6) 31 (59.6) 29 (55.8) 48 (92.3) 3 (5.8) 3 (5.8) 3 (5.8) 3 (5.8) 3 (5.8) 0 (0) 2 (3.8) 2 (3.8) 2 (3.8) 7 (13.5) 0 (0) 34 (65.4) 3 (65.4) 3 (61.5) 32 (61.5) 31 (59.6) 32 (55.8) 33 (5.8) 33 (5.8) 33 (5.8) 33 (5.8) 33 (5.8) 34 (65.4) 34 (65.4)	n (%) 32 (61.5) 32 (61.5) 32 (61.5) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 32 (61.5) 31 (59.6) 31 (59.6) 32 (55.8) 48 (92.3) 48 (92.3) 48 (92.3) 48 (92.3) 48 (92.3) 48 (92.3) 3 (5.8) 3 (5.8) 3 (5.8) 3 (5.8) 3 (5.8) 3 (5.8) 3 (5.8) 5 (9.6) 2 (3.8) 2 (3.8) 2 (3.8) 5 (9.6) 2 (3.8) 7 (13.5) 7 (13.5) 7 (13.5) 7 (13.5) 7 (13.5) 9 (0) 94 (65.4) 94 (65.4)	n (%) 32 (61.5) 32 (61.5) 32 (61.5) 32 (61.5) 32 (61.5) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 31 (59.6) 32 (53.8) 48 (92.3) 5 (8) 5 (9.6) 5 (9.6) 2 (3.8) 2 (3.8) 2 (3.8) 2 (3.8) 2 (3.8) 7 (13.5)

Keywords: Urea supplementation: (E, without urea; a, 0.5% urea; b, 0.75% urea; c, 1.0% urea; d, 1.25% urea), n (%), number and percentage of antibiotic resistant strains.

* = significant at 95% confidence limit by chi square analysis.

Table 3: Minimum inhibitory concentrations of antibiotic resistant Proteus strains by microbroth dilution method.

Proteus spp. MIC (mg/L)									
	E	a	ъ	c	d				
Antibiotics	n ()	n()	n ()	n ()	n ()				
Ampicillin	32 (75+33.1)	32 (74+33.9)	32 (76+32.3)	32 (76+32.3)	32 (77+34.3)				
Amoxicillin	33 (81+33,5)	33 (81+33.5)	33 (82+32.4)	33 (83+34.3)	33 (90+31.5)				
Co-amoxiciav	29 (77.2+33.7)	29 (77.2+33.7)	29 (79.4+34.9)	31(79.5+35.9)	31 (80.5+31.9)				
Cotrimoxazole	48 (82.7+33.6)	48 (83.3+32.8)	48 (84+34.1)	48 (84.7+33.3)	48 (84+34.1)				
Ceftazidime	3 (18.7+12.2)	3 (21.3+9.2)	3 (18.7+12.2)	3 (21.3+9.2)	3 (26.7+9.2)				
Ceftrixone	3 (21.3+9.2)	3 (26.7+9.2)	3 (26.7+9.2)	3 (21.3+9.2)	3 (21.3+9.2)				
Ciprofloxacin	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)				
Gentamicin	2 (8.2+1.8)	2 (12.0+5.7)*	5 (20.8+10.7)*	5 (22.4+8.8)*	7 (21.7+10.0)*				
Nalidixic acid	2 (12.0+5.7)	2 (12.0+5.7)	2 (16.0+0.0)	2 (12.0+5.7)	2 (12.0+5.7)				
Nitrofurantoin	7 (59.4+34.2)	7 (73.1+40.1)*	7 (82.3+31.2)*	7 (73.1+40.1)*	7 (86.9+40.1)*				
Ofloxacin	0 (0)	0 (0)	Ò (O)	0 (0)	0 (0)				
Streptomycin	35 (85.9+33.6)	35 (85.9+33.6)	35 (90.5+33.3)*						
Tetracycline	50 (90.9+33.8)	50 (90.9+33.8)	50 (92.2+34.0)	50 (92.2+34.0)	50 (92.2+34.0)				

Keywords: Urea supplementation: (E. without urea: a, 0.5% urea: b, 0.75% urea: c, 1.0% urea; d, 1.25% urea), n (), number of antibiotic resistant strains, numbers in parentheses are mean \pm SD of MIC values, * = significant at 95% confidence limit by Student's t-lest.

Table 4: Identification of the extended beta lactamases producing strains among the *Proteus spp*.

Proteus spp.

	Е	a	ь	с	d	
ESBLs n (%)	2 (3.8)	2 (3.8)	2 (3.8)	2 (3.8)	2 (3.8)	
Inderminate	0 (0)	1 (1.9)	1 (1.9)	1 (1.9)	2 (3.8)	

Keywords: Urea supplementation: (E, without urea; a, 0.5% urea; b, 0.75% urea; c, 1.0% urea; d, 1.25% urea), n (%), number and percentage of ESBL producing strains or inderterminate result.

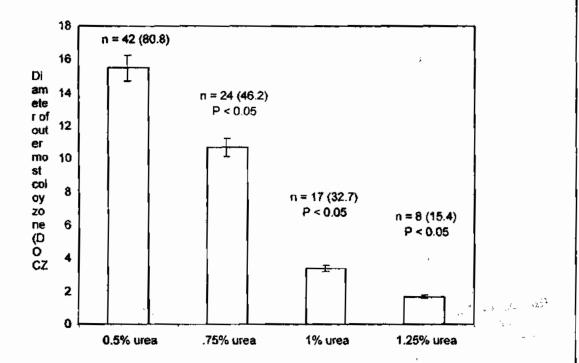


Figure 1: Effect of urea on swarming among *Proteus* isolates from Lagos, Nigeria.

Bars represent mean DOCZ valuesand projections on bars indicate deviations from mean.

n = number of swarmed *Proteus* strains, figures in parentheses indicate percentages.

DISCUSSION

Urea is primarily used in selective and composite media to urease-producing identify microorganisms (20). In recent the possibilities times. anti-swarming exploiting the property of urea to aid isolation and identification of colonies on solid media are being tested (12). In the present study. we found urea useful in refraining swarming of Proteus spp causing urinary tract infections in Lagos. In vitro, urea was also observed to allow isolation and identification of K. pneumoniae, Pseudomonas aeruginosa; S. saprophyticus and E. coli. Pseudomonas aeruginosa identified at 1% urea is higher than the 0.5% urea reported by Fons et al (12). The disparity in concentration could urea attributed to strain variation and difference in swarming ability. Antigenic differences and varying capsular polysaccharide composition have been found among swarming Proteus spp (21). All the Proteus isolates examined survived urea at 1.25% and this suggests that urea is not inhibitory to cell growth at this concentration. This may not be unexpected since many media formulations for identification and speciation of bacteria contain up to 2% urea (22).

In this study, we found that the responses of our isolates to

nitrofurantoin, amoxicillin, gentamicin and streptomycin were compromised following exposure to urea. This became more evident the minimum inhibitory concentrations of these antibiotics were determined. In conducted by Ward et al (15), Pnitrophenylglycerol was found to increase the MICs aminoglycosides; gentamicin and tobramycin and decrease the MICs of ticarcillin, ciprofloxacin colistin against **Pseudomonas** aeruginosa. In Nigeria, several studies prospective have condemned the use of streptomycin amoxicillin for empirical treatment of bacterial infections (23, 24). Recently, high amoxicillin resistant Helicobacter pylori strains were found in the biopsy samples of patients with gastritis and peptic ulcer in Western Nigeria (25). Although MICs of gentamicin and nitrofurantoin among the urea treated and untreated Proteus strains were above their respective break points (17), it is very important to adopt a cautionary use *urea investigating when susceptibility of Proteus to these antibiotics. The p. evalence bacterial pathogens that resistance to these antibiotics in human infections is generally low in Nigeria Therefore, (23).discrepancies associated with the assessment of pathogens to these antibiotics may jeopardize control measures and heighten the risk of multidr g resistant infections. The identification of pathogens ESBL further provides an insight into their mechanisms resistance to beta-lactam drugs. pathogens have experimentally demonstrated to stably express de-repressed, constitutive chromosomal class 1β-lactamases, which hydrolyze most β-lactam antibiotics except carbapenems (26). This study has demonstrated the inability of urea at□1.25% to cause no in discrepancies **ESBL** classification and thus provides an additional credit to its diagnostic usefulness in clinical medicine and public health.

REFERENCES

- 1. Orrett FA. Prevalence of Proteus species in urinary tract infections in a regional hospital in Trinidad. Zhonghua Yi Xue Za Zhi (Tapei). 1999; 62: 438 442.
- Cutner A, Cardoso LD, Benness CJ. Assessment of urinary symptoms in early pregnancy. Br. J. Obstet. Gynaecol. 1991; 98: 1283 - 1286.

- 3. Rasmussen PE, Nielsen FR. Hydronephrosis during pregnancy: A literature survey. Eur. J. Obstet. Gynaecol. Reprod. Biol. 1988; 27: 249.
- 4. Kesah CN, Coker AO, Alabi SA, Olukoya DK. Prevalence, antimicrobial properties and beta-lactamase production of haemolytic enterobacteria in patients with diarrhoea and urinary tract infections in Lagos, Nigeria. Centr. Afr. J. Med. 1996; 42: 147-150.
- 5. Adeyeba OA, Adegoke GO.
 Urinary tract pathogens
 common amongst the
 inhabitants of a state capital,
 SouthWest Nigeria. Int. J.
 Zoonoses. 1986; 13: 153 157.
- Adeyemo AA, Gbadegesin RA, Onyemenem TN, Ekweozor CC. Urinary tract pathogens and antimicrobial sensitivity patterns in children in Ibadan, Nigeria. Ann. Trop. Paediatr. 1994; 14: 271 274.
- 7. Airede AI. Urinary tract infections in African neonates. J. Infect. 1992; 25: 55-62.

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- 8. Rauprich O, Matsushita M, Weijer CJ, et al. Periodic phenomenon in Proteus mirabilis swarming colony development. J. Bacteriol. 1996; 178: 22.
- 9. Sabbuba N, Hughes G, Stickler DJ. The migration of *Proteus mirabilis* and other urinary tract pathogens over Foley catheters. *BJU Int.* 2001; **89**: 55 60.
- 10. Cook, SW, Mody N, Valle J, Hull R. Molecular cloning of Proteus mirabilis uroepithelial cell adherence (uca) genes. Infect. Immun. 1995; 63: 2082 2086.
- 11. Allison C, Coleman N, Jones PL, Hughes C. Ability of Proteus mirabilis to invade human urothelial cells is coupled to motility and swarming differentiation.

 Infect. Immun. 1992; 60: 4740-44746.
- Fons JAM, Asten V, Gaastra W. Urea refrains swarming of Proteus mirabilis. J. Clin. Microbiol. 1999; 37: 1652 1653.
- William FD. Abolition of swarming of *Proteus* by Pnitrophenylglycerin: general

- properties. *Appl. Microbiol.* 1973; **25**: 745 750.
- 14. Smith DG, Alwen J. Effect of activated charcoal on the swarming of *Proteus*. *Nature*. 1966; **212**: 941 942.
- Ward PB, Palladino S, Looker 15. P. P-JC. Feddema nitrophenylglycerol in susceptibility testing media alters the **MICs** of antimicrobials for Pseudomonas aeruginosa. J. Antimicrob. Chemother. 1993; **31**: 489 - 496.
- Cowan ST. Manual for identification of medical bacteria. 2nd ed. Cambridge University Press. London. 1974.
- 17. Bauer AW, Kirby WMM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standard single disk method. Am. J. Clin. Pathol. 1966; 36: 493 496.
- 18. National Committee for Clinical Laboratory Standards. Methods for dilution. Antimicrobial susceptibility tests for bacteria that grow aerobically. Second edition. Approved standard M7 - A2. NCCLS. Villanova PA. 1990:

- 19. Livermore DM, Yuan M. Antibiotic resistance and of extended production spectrum B-lactamase amongst Klebsiella spp from intensive care units Antimicrob. Europe. J. Chemother. 1996; 38: 409 - 424.
- 20. Blaser MJ. Helicobacter pylori and related organisms. In: Mandell CL, Douglas R, Bennett JT (eds). Principle and Practice of Infectious Diseases. 5th edition. Churchill Livingstone. 2000.
- 21. Rahman MM, Guard-Petter J, Asokan K, et al. The structure of the colony migration factor from pathogenic Proteus mirabilis. A capsular polysaccharide that facilitates swarming. J. Biol. Chem. 1999; 274: 22993-22998.
- Oxoid. Manual for culture media preparation. Oxoid. 1995.

- Iwalokun BA, Gbenle GO, 23. Smith SI. Ogunledun A, Akinsinde KA. Epidemiology of shigellosis in Lagos, Trends Nigeria: in antimicrobial resistance. J. Health Popul. Nutr. 2001; 19 : 183 - 190.
- 24. Akinyemi KO, Olukoya DK, Oyefolu AO, Amorighoye EP, Omonigbehin EA. Prevalence of multidrug resistance Salmonella typhi among clinically diagnosed typhoid fever patients in Lagos, Nigeria. Z. Naturforsch. 2000; 55: 489 - 493.
- 25. Smith SI, Oyedeji KS. Arigbabu AO, Atinomo C, Coker AO. High amoxicillin resistance Helicobacter in pylori isolated from gastritis and peptic ulcer patients in Western Nigeria. Gastroenterol. 2001; 36:67 - 68.
- Livermore DM. β-lactamases in laboratory and clinical resistance. Clin. Microbiol. Rev. 1995; 8:557 584.

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THE EFFECT OF SOME NIGERIAN LOCAL HERBS ON HELICOBACTER PYLORI

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Four Nigerian medicinal plants commonly used in the treatment of bacterial infections were tested for antimicrobial activity against twenty local strains of Helicobacter pylori recovered from patients with gastro-duodenal ulcers and gastritis. In vitro agar diffusion assay revealed anti-Helicobacter pylori activity of ethanolic extracts of C. papaya and M. lucida to 80% (16/20) of the isolates tested. While the ethanolic extracts of O. gratissimum and P. amarus inhibited the colonial growth of 35% (7/20) of these strains. The zones of inhibition ranged from 5 - 20 mm in diameter. Contrastingly, the aqueous extracts of these plants appeared to lack anti-Helicobacter pylori activity except in M. lucida and O. gratissimum where inhibition of a total of three isolates was observed. The present results suggest the presence of anti-Helicobacter pylori principles in ethanolic extracts of C. papaya and M. lucidaand support their future use in the treatment of ulcers and gastritis in Nigeria.

INTRODUCTION

It is over a decade that Helicobacter pylori infections were known as a major cause of gastroduodenal ulcers. gastritis and stomach cancer with burden of cases documented in developing countries (1, 2). Most effective therapies employ synergic action between a gastric acid release inhibitor and one or antibiotics eradicate more to Helicobacter pylori, its urease and associated diseases (3, However, the implementation of these therapies in communities where the poor bears the greater brunt of the disease is cost ineffective as concerned drugs are poorly patronized (5).

In Nigeria, Helicobacter pylori is fastly replacing non streroidal anti-inflammatory drugs (NSAIDs) as causal agent of gastro-duodenal ulcers and duodenal perforation cases are also of significant increase (6). More worrisome is the increased resistance Helicobacter pylori isolates to amoxycillin metronidazole, and tetracycline in vitro (7) in a manner that discourage their future clinical against Helicobacter pylori infection in the country. In Nigeria, triple therapy involving omeperazole, metronidazole amoxycillin are widely used in severe cases and treatment failures with this combination have been reported (8). Alternative

therapies that could be used are expensive, have undesirable side effects (9) and a long list of contraindications (10).The discoveries that Allium vegetables anti-Helicobacter habour substances (11,12) have further heightened the global search for similar compounds in other medicinal plants and this to some extent has yielded encouraging results that have the therapeutic applications in areas of discovery as a barrier (13). Morinda lucida (Rubiaceae), Ocimum gratissimum (Lamiaceae), Carica papaya (Caricaceae) and **Phyllanthus** amarus (Euphorbiaceae) extracts are among the folkloric remedies have been confirmed scientifically to possess clinical values against protozoal bacterial infections in Nigeria (14, 15, 16, 17). These plants grow abundantly in Nigerian soils and are not ethnically or age group biased in use. This further explains why their potentials in the future treatment of gastritis and ulcers in Nigeria are investigated.

MATERIALS AND METHOD Plant materials

The plants, were collected from various local markets in Lagos, identified and confirmed by Mrs. B. Opere of the Department of Botany, Lagos State University. Voucher samples of these plants were subsequently deposited in the Department. The plants used were listed in Table 1.

Extraction

A simple extraction procedure of Olukoya et al (18) was adopted to aqueous prepare and organic extracts of the plants tested. To prepare aqueous extracts, 1.1 g of plants' leaves (previously dried at 50°C and ground into fine powders) were steeped in 10 ml of steriledistilled water at 30 - 32°C for five days. The organic extracts were prepared by steeping 1.2 g of plant materials in 5 ml of 40% ethanol. Extracts were then passed through Hemmings filters (BTI UK) and the resulting sterile filtrates aseptically transferred to sterile bottles and labeled as crude extracts of individual plants. The organic extracts were subsequently reconstituted with phosphate buffered saline solution (pH 7.2) to nullify the effect of ethanol on the tested organisms. A mixture of 0.1 ml of sterile water and 5 ml of 40% ethanol was prepared as a control.

Microbial cultures

Twenty strains of Helicobacter pylori recovered from the biopsy samples of patients with gastritis and gastro-duodenal ulcers from Western Nigeria were used as test organisms. Helicobacter pylori ATCC 49503 was used as control. All organisms were cultured on Columbia agar base (Oxoid, CM331) containing 7% sheep blood.

Sensitivity Testing

Antimicrobial susceptibility testing was carried out using the agar diffusion technique. In brief, (Oxoid. Isosensitest agar plates were holed (6 mm diameter) with the aid of a sterile cork-borer and seeded with 10 µL of H. pylori suspension (McFarland 3). The plates were dried in the air and 100 µL of plant extract was introduced into the wells. The plates were incubated microaerophilically (5% O2, 10% CO₂) at 37°C for 4 days. Holes containing bacterial suspension (10 μ L of 9 x 108 CFU/ml) and sterile water or ethanol (100 µL) were used as controls. Diameters of zones of inhibition of both the tested organisms and standard strain (H. pylori ATCC 45903) were measured in millimeters (mm) and recorded.

RESULTS.

Table 2 gave the summary of antimicrobial activity aqueous and ethanolic extracts of the plants against the twenty H. pylori isolates and the standard strain. The ethanolic extracts of C. papaya, M. lucida, O. gratissimum and P. amarus provided evidence of anti-Helicobacter pylori activity in 80% (16/20) and 30% (7/20) of the isolates tested. Helicobacter pylori ATCC 49503 was observed to be susceptible to the organic extracts of C. papaya and M. lucida only. The zones of growth inhibition were 5 - 20 mm in diameter. Apart from the water extracts of M. lucida and O. gratissimum, which inhibited the growth of 10% (2/20, 5 - 15 mm)and 5% (1/20, 10 - 15 mm) of the isolates tested, aqueous extracts of other plants were found susceptible to these isolates. The standard strain was resistant to all aqueous preparations. Unlike sterile water wells, no colonial growth was found in 40% ethanol control wells.

Table 1: The local h	erbs selected for testing	g	
Botanical name	Local name*	Plant part tested	
Carica papaya	Ibepe	Leaf	
Morinda lucida	Ejirin	Leaf	
Octimum gratissimum	Efirin	Leaf	
Phylllanthus amarus	Ehinolubinsowo	Leaf	

^{*} Nigerian (Yoruba) names.

Table 2:	:	Antii	nicro	bial a	ial activity of ethanol and water extracts of the four local herbs					
Strain code no	C. j	рарауа	М. і	lucida	O. gra	tissimum	P. a .	marus	Sterile wat	er 40% Ethano
	E	W	E	W	E	W	E	W		
Hp 1	0	O	2+	0	O	O	0	0	O	2+
Нр 2	2+	O	2+	0	1+	0	0	0	0	3+
Hp 3	1+	0	2+	0	O	0	0	O	O	1+
Hp 4	2+	O	2+	2+	0	0	0	O	0	2+
Hp 5	1+	0	3+	0	2+	0	2+	0	0	3+
Нр 6	3+	0	1+	0	0	O	0	0	0	+1
Нр 7	2+	0	2+	0	2+	0	0	0	O	2+
Hp 8	0	0	0	0	0	0	0	0	O	1+
Hp 9	2+	0	0	0	O	0	0	0	0	2+
Hp 10	2+	O	2+	0	0	O	<u>t</u> +	0	0	2+
Hp 11	2+	0	2+	0	2+	0	2+	0	0	3+
Hp 12	1+	0	3+	0	2+	0	2+	0	0	2+
Hp 13	2+	0	1+	1+	0	0	0	0	0	3+
Hp 14	0	0	0	0	0	0	0	0	0	2+
Hp 15	2+	0	2+	0	0	2+	1+	0	0	3+
Hp 16	l+	0	2+	0	1+	0	1+	0	0	2+
Hp 17	2+	0	3+	0	0	0	0	0	0	2+
Hp 18	0	0	0	0	0	0	0	0	0	2+
Hp 19	2+	0	2+	0	1+	0	1+	0	0	3+
Hp 20	3+	0	3+	0	0	0	0	0	0	2+
Hp 49305	2+	0	2+	0	0	0	0	0	0	3+

Keys: W = water extract; E = Ethanolic extract; 0 = No inhibition; 1+=5-9 mm diameter zone of inhibition; 2+=10-15mm; 3+=16-20mm; Hp = Helicobacter pylori strains.

DISCUSSION

In this study, ethanolic extracts of C. papaya and M. lucida were observed to prevent the growth of 80% of Helicobacter pylori strains tested in vitro. Similar extracts of O. gratissimum and P. amarus also demonstrated anti-Helicobacter pylori activity but in only 35% (7/20) of the isolates. The lack of inhibition observed in water wells confirmed the viability of all the isolates tested. While the inhibitory effect of 40% ethanol attested to the appropriateness of the reconstitution procedure, the

poor antibacterial activity of the water extracts of these plants implies that water has inadequate power to extract anti-Helicobacter pylori principles from these plants. However, With respect to organic extraction, this finding has provided scientific evidence antibacterial activity of Morinda lucida in vitro as most scientific findings described the plant as an anti-malaria herb (14). The study of which Agomo et al(19).demonstrated a complex array of cellular responses to M. lucida

administered to mice infected with Plasmodium uoeli niaeriensis seemed to provide an indication that biological numerous properties are inherent in this plant. Ocimum gratissimum has been extensively demonstrated to inhibit aetiologic agents αf diarrhoeal in vitro and in vivo (15. 20). However, the result obtained study this should interpreted with caution as strains of aratissimum 0. characterized by varying chemical composition (21). The present study has also extended the biological functions Carica of whose seeds papaya have demonstratable evidence of having antifertility effects in rats (22). Although, antimalaria activity of P. amarus in mice and rats has been observed in the laboratory (Personnal communication), this is first time anti-Helicobacter pulori activity will be ascribed to this plant in Nigeria. Based on our findings, there is no doubt that these plants hold tremendous clinical promise especially in rural communities, which provide the greater number of patients and severe cases of gastro-duodenal ulcer. The present study is still in its infancy and therefore invite more research studies to elucidate the active anti-Helicobacter pylori substances in these plants. investigate synergy associated with extract combination leading

to ultimate suggestion of whether these plants can be combined with orthodox drugs to met the criteria of gastric acid suppression, *H. pylori* eradication and stomach protection in the treatment of ulcers and gastritis. In conclusion, the present study has revealed, the tremendous potentials inherent in ethanolic preparations of *C. papaya* and *M. lucida* if adopted for future treatment of ulcers and gastritis in Nigeria.

REFERENCES

- 1. Dixon MF. Helicobacter pylori and gastritis. In: Rathbone RV BJ, Heatley (eds). Helicobacter pylori and gastro-duodenal disease. Blackwell Scientific Publications. Oxford. UK. 1992: 124 - 129.
- Feldman RA, Eccersley AJ, Hardie JM. Epidemiology of Helicobacter pylori, acquisition, population prevalence and disease to infection ratio. Br. Med. Bull. 1998; 54: 39 53.
- 3. Mansour-Ghanael F, Fallah M, Shafaghi A. Eradication of Helicobacter pylori in duodenal ulcer disease. Tetracycline & Furozolidone vs. Metronidazole & Amoxicillin in

omeperazole based therapy. *Med. Sci. Monit.* 2000; **8**: 127 - 130.

- 4. **Jonkers Broek** D. DE. IV. Dooren et al. Antibacterial effect of garlic omeperazole and Helicobacter pulori. J. Antimicrob.Chemother. 1999 : **43** : 837 - 839.
- 5. Drouin E. Helicobacter pylori: novel therapies. Can.
 J. Gastroenterol. 1999; 13:
 581 583.
- 6. Smith SI, Oyedeji KS,
 Arigbabu O, et al. The
 prevalence of Helicobacter
 pylori from gastritis and
 ulcer patients in Western
 Nigeria. Biomedical Letters.
 1999; 60: 115-120.
- Smith SI, Oyedeji KS;
 Arigbabu AO, Atinomo C,
 Coker AO. High amoxycillin resistance in Helicobacter pylori isolated from gastritis and peptic ulcer patients in Western Nigeria. J. Gastroenterol. 2001; 36:67-68.
- Holcombe C, Kaluba J, Lucas SB. Helicobacter pylori infection and gastritis in healthy Nigerians. Eur.

- J. Epidemiol.1994; **10**: 223 225.
- 9. Bell GD, Powell KU, Burridge SM, et al. Helicobacter pylori eradication efficacy and side effect profile of a combination of omeperazole, amoxycillin and metronidazole compared with four alternative regimens. Q. J. Med.J. Assoc. Phys. 1993; 86:743-750.
- 10. O'Gara EA, Hill DJ, Maslin DJ. Activities of garlic oil, garlic powder and their diallyl constituents against Helicobacter pylori. Appl. Environ. Microbiol. 2000; 66: 2269 2273:
- 11. Cavallito CJ, Bouley JH. Allicin, the antibacterial principle of Allium sativum. I. Isolation, physical properties and antibacterial action. J. Am. Chem. Soc. 1944; 66: 1950-1954.
- 12. Cellini L, DiCampli E, Masuli M, DiBartolomeo S, Allocati N. Inhibition of Helicobacter pylori by garlic extract (Allium sativum). FEMS. Immunol. Med. Microbiol. 1996; 13: 273-277.

- 13. Fabry W, Okemo P, Ansorg R. Activity of East African medicinal plants against Helicobacter pylori. Chemother. 1996; 42:315-317.
- 14. Makinde JM, Obih PO. Screening of Morinda lucida leaf extract for antimalaria action on Plasmodium berghei berghei mice. Afr. J. Med. Med. Sci. 1985; 14: 59-63.
- 15. Ilori MO, Sheteolu AO, Omonigbehin EA, Adeneye AA. Antidiarrhoeal activities of Ocimum gratissimum (Lamiaceae). J. Diarrh. Dis. Res. 1996; 14: 283-285.
- Emeruwa, AC. Antibacterial substance from Carica papaya fruit extract. J. Natur. Products. 1988;
 45: 123-127.
- 17. Sofowora A. The present status of knowledge of the plants used in traditional medicine in West Africa. A medical approach and a chemical evaluation. J. Ethnopharmacol. 1980; 2: 109-118.

- 18. Olukoya DK, Idika N, Odugbemi TO. Antibacterial activity of some medicinal plants from Nigeria. J. Ethnopharmacol. 1993; 36: 69-72.
 - 19. Agomo PU, Idigo JC, Afolabi BM. "Antimalarial" medicinal plants and their impact in cell populations in various organs of mice. Afr. J. Med. Med. Sci. 1992; 21:39 –46.
- 20. Offiah VN, Chikwendu UA. Antidiarrhoeal effects of Ocimum gratissimum leaf extract in experimental animals. J. Ethnopharmacol. 1999; 68: 327 330.
- 21. Angers P, Morales MR, Simon JE. Fatty acid variation in seed oil among Ocimum species. J. Am. Oil. Chem. Soc.1996; 73: 393-395.
- 22. Udoh P, Kehinde A. Studies on antifertility effect of pawpaw seeds (*Carica papaya*) on gonads of male albino rats. *Phytother. Res.* 1999; **13**: 226 228.

BACTERIOLOGICAL QUALITY OF SOBO DRINKS RETAILED WITHIN ILORIN METROPOLIS

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Sobo drinks retailed within llorin metropolis were investigated for their bacteriological quality, in terms of the total bacterial count and types of bacterial species present. The sobo drinks were found to have an average pH of 3.2. The bacterial counts were generally high ranging from 5.0x104 to 24x104 CFU/ml. Six bacterial species; Bacillus subtilis, Staphylococcus aureus, Streptococcus faecalis, Klebsiella spp, Enterobacter spp, and Escherichia coli were isolated. Consideration of the distribution pattern showed that samples from Unilorin Main Campus and General Post Office contained all the isolates, while three of the isolates; Escherichia coli, Bacillus subtilis and Staphylococcus aureus were common to all samples. Possible sources of the contaminants and their public health implications are discussed and suggestions offered on ways of ensuring safety of the sobo drinks.

INTRODUCTION

Sobo is a drink produced from carefully selected bright red sorrels of Rossell hemp (Hibiscus sabdariffa). It is a hot water extract of sorrel of the Rossell hemp: a shrub with distinct bright red coloured sorrels that is fibrous, and is native to West Africa (1). The drink is commonly used as refreshment and in the entertainment of guest at social occasions. The sorrels are sorted and cleaned before soaking in water. It is then heated to boiling after which the plant materials are removed by sieving. Sugar and other additives (flavours) are then added as desired. The hot extract is allowed to cool after which it is dispensed into nylons or plastic containers. It is the cooled extract. which is usually refrigerated before retailing that is taken as

'sobo'. The consumption of sobo has continued to spread across Nigeria been helped mainly by the worsening economic fortune of the majority of people and the escalating price of soft drinks and beverages that are commonly patronized.

Foods frequently serve as vehicle for spread οf several organisms some of which pathogenic. Many (2).suppers and banquets have come to a disastrous end when homeprepared foods and drink serve not only as food and drinks for guest the vehicle but also as for transmitting Staphylococcal food poisoning (3). In view of the fact that sobo is never subjected to any form of post-production treatment that can eliminate, or at least reduce the bacterial load in the drink, it could be a potential source

of health hazard. Also the activities involved in the cooling. and subsequent dispensing of the drink into containers represent potential sources of health hazards. Some researchers have reported that gastrointestinal illness characterized by diarrhoea. abdominal cramps, and vomiting which may be assumed as been of unknown actiology may arise from drinking drinks contaminated with microorganisms. In this retailed drinks sobo were purchased from various location within Ilorin. They were analyzed for their bacteriological quality as indicated by the number (counts) and kinds of bacteria thev harbour.

MATERIALS AND METHODS

Samples of sobo drinks were purchased from retailers from five locations within the metropolis; these were Unilorin Secondary School, Unilorin Main Campus, Market, Oja-Oba, Ipata General Post Office. These were immediately taken to laboratory for analysis. The pH of all the samples was determined using Pye Unican pH meter (Model 292 MK2). The bacterial populations were determined by preparing tenfold serial dilution of the samples, and then plating 1 ml of desired dilution on nutrient agar using pour plate count method (5). Representative colonies of bacterial isolates were selected and purified by subculturing on nutrient agar using the streaking method. Pure cultures were then characterized and subsequently identified according to Bergy's Manual ofDeterminative Bacteriology (6).

RESULTS

The pH of the samples ranged from 2.6 to 4.1 (Table 1), the mean pH was 3.04. The bacterial counts were generally high; it ranged from 5 x 10^4 to 24×10^4 CFU/ml (Table 2). A sample from Unilorin Secondary School had the highest count (24 x 104 CFU/ml) while a sample from Ipata Market had the lowest count (5 x 104 CFU/ ml). A total of six bacterial species were identified. These were Bacillus subtilus. Staphylococcus aureus. Streptococcus faecalis, Klebsiella Enterobacter and spp, spp Escherichia coli. Their distribution among the collection points is shown in Table 3. Samples from Unilorin Main Campus and General Post Office were observed to contain all the isolates and three of the isolates; Escherichia coli, Bacillus subtilus and Staphylococcus aureus were common to all samples. least Klebsiella the was 🦏 encountered; been found only in some samples from Unilorin Main Campus and General Post Office.

Table 1: pH of Sobo Samples Examined

Collection Point.					
	Collection Time				
	1	2	3	4	5
Unilorin Secondary School.	2.6	3.0	3.2	3.3	3.0
Unilorin Main Campus.	3.2	3.2	3.3	3.2	3.1
Ipata Market.	3.2	3.1	4.1	3.1	3.2
Oja-Oba	3.0	3.2	3.2	3.2	3.2
General Post Office.	3.2	3.2	3.0	3.2	3.2

Table 2: Population of Bacteria in Sobo Samples Examined

Collection Point.	Mean Total Bacterial Count (cfu/ml) x 10 ⁴ Collection Time						
	1	2	3 -	4	5		
Unilorin Secondary School.	9.0	16.0	24.0	14.0	13.0		
Unilorin Main Campus.	21.0	12.0	6.0	13.0	11.0		
Ipata Market.	5.0	16.0	8.0	11.0	7.0		
Oja-Oba	16.0	9.0	11.0	16.0	8.0		
General Post Office.	13.0	7.0	8.0	11.0	14.0		

Table 3: Distribution of Bacterial Isolates Among Collection Points.

Bacterial species				Source of Sample			
	USS	UMC	IMT	OJA	GPO		
Staphylococcus aureus	+	+	+	+	+		
Streptococcus faecalis	+	+	+	-	+		
Bacillus subtilis	+	+	+	+	+		
Klebsiella sp		+		-	+		
Enterobacter sp	-	+	-	+	+		
Escherichia coli	+	+	+	+	+		

Кеу **Us**s:

Unilorin Secondary School.

IMT: Ipata Market

GPO: General Post Office.

UMC: Unilorin Main Campus.

OJA: Oja Oba

DISCUSSION

The pH values obtained indicate that the sobo drinks were acidic in nature and hence normally not favour the proliferation of bacteria. However pH alone is not a sufficient parameter to predict the chances of survival and proliferation of

bacteria in the sobo drinks. The high bacterial counts obtained are indicative of poor production conditions. In the light of the amount of heating that goes into sobo production, the presence of bacteria can only be as a result of post heating contamination. Simlar post treatment contamination of water has been reported (7). This can occur during cooling of the hot extract, addition of flavours and sweetener, or dispensing of the extract into nylons. Utensils and water used during the post heating stages can also serve as sources of contamination. Water used in processing has been identified as the major source of contamination of locally made drinks (8).

The presence of Staphylococcus aureus in all samples is indicative of human contamination after heating. This could be from direct human contact such as fingers dipped into the extract to taste the sweetness, or indirectly through additives or utensils. The organism is associated with enterotoxin characterized by short incubation period (1-8 hours). violent nausea, vomiting diarrhoea. There is no fever (9). The presence of Bacillus subtilis is indicative οf environmental contamination, which could have resulted from exposure of the extract to air or contact of utensils used with the soil or from the water used in the post heating stages. The presence of E. coli. Klebsiella and Enterobacter spp suggests faecal contamination, the while presence Streptococcus faecalis particularly indicates a fairly recent faecal

contamination (10). Some strains of with coli are associated production stable of heat enterotoxins, hence, their presence constitute a health risk (11). Although there are no specific guidelines as at present to give a standard for assessing sobo quality, the WHO standard for drinking water could be used since sobo serves as a drink. The standard requires that drinking water should not contain pathogenic organisms and should be free of bacteria indicative of polution with excreta (11). Hence it can be adjudged that the sobo drinks retailed in most location within Ilorin, as obtained this study, are bacteriologically fit for consumption.

Generally the drinks appear to have been contaminated due to saniatary conditions, unhygienic production practices and prolonged exposure to the environment. Therefore to ensure the safety of the sobo drinks, producers must maintain a clean environment, minimize contact with the extract and exposure to the environment after heating and also maintain a high personal hygiene level. Also utensils should properly washed before use and clean water should be used at all stages of production. On a larger note further studies examining the chemistry of the extract and its effect(s) on visceral organs and body system are desirable before an overall acceptability can be placed on the sobo drink.

REFERENCES

- 1. Wilson FD, Menzel MY. Kenaf (Hibiscus cannabis), Rosselle (Hibiscus sabdariffa). Economic Botany. 1974; 18: 80-81.
- Singleton P. Applied Bacteriology I: food. Bacteria in Biology, Biotechnology and Medicine. 4th edition. John Wiley & Sons Ltd, West Sussex, England. 1999 : 267-273.
- 3. Alabi DA, Akinsiyun AA. Studies of the Microbial Quality of Filtered Water in Households of A University Community in Nigeria. *J. Hyg.* 1996; **96**: 239-248.
- Cruck VF, Shank OO. Code of Practice on General Principle of Food Hygiene. Kesington University Press. 1984: 99-104.
- Collins EH, Lyne PM, Greange JM. Microbiological Methods. 6th edition. Butterworth and Co. 1989: 102-106.
- Buccanhan KE, Gibbon NE.
 Bergy's Manual of

Determinative Bacteriology. 8th edition. The William and Wilkins Co. Baltimore. 1974.

- 7. Akinyosoye FA, Nwosis NI.
 Microorganisms Associated
 With The Production of
 Kunun Zaki, A Nigerian
 cereal drink. 1994; 6(2):
 133-137.
- Okeke IN, Lamikanra 8. A, Czeczulin J, etal. Heterogenous virulence of enteroaggregative E. coliisolated from children in South Western Nigeria. J. Infect. Dis. 2000; 181(1): 252-260.
- Brock TD, Madigan MT, Martinko JM, Parker J. Biology of microorganisms. 7th edition. Prentice Hall, Englewood Cliffs, New Jersey. 1994.
- 10. Galvani MW. Faecal
 Cotamination- Analyst
 Responsibility Sewage Works.
 National Academy Press,
 Washington DC. 1974: 6669.
- 11. WHO. World Health Organization guidelines for drinking water 1 (Recommnedation).

 Macmillian. Ceuterick .1984a : 17-38.

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BACTERIOLOGICAL EXAMINATION OF CHRONIC OSTEOMYELITIS CASES IN ILE-IFE, SOUTHWESTERN NIGERIA

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The bacteriological examination of chronic osteomyelitis cases in Ile-Ife, revealed Proteus mirabilis as the predominant isolate encountered (23.1%), Staphylococci constituted 30.7% of the total bacterial isolates with Staphylococcus aureus accounting for only 10.2%, Other Gram-positive cocci cultured include, Staphylococcus spp (20.5%), coagulase negative staphylococci (CONS) (12.8%) and Streptococcus spp (1.2%). Gram-negative rods constituted 55.1% of the total isolates with Pseudomonas aeruginosa being 8.5%, Eschericia coli 5.1% Citrobacter freundii, Salmonella spp 2.5% each. Antibiotic sensitivity test revealed all isolates to be multi-resisfant to traditional antimicrobials. which is of epidemiological importance in treating cases of chronic osteomyelitis in this environment. The study suggests institution of aggressive therapeutic interventions to evert possible segualae.

INTRODUCTION

Osteomyelitis is an inflammation of the bone caused mainly by microorganisms and can be categorized into acute and chronic forms. According to Ofiaeli osteomyelitis (1). chronic perhaps one of the commonest orthopaedic diseases in the tropics among children and adolescents under the age of twenty years. Multiple factors predispose individuals to chronic osteomyelitis among which are mismanagement of acute osteomyelitis (2),haemoglobinopathies such as sickle cell diseases (3.4.5).Haematogenous bone infections. which persist and defy antibiotic environmental therapy. personal hygiene (6). Prompt

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bacteriological investigations and aggressive management of patients chronic ostomyelitis essential for effective resolution and prognosis of this condition. While treatment modalities of chronic involve osteomyelitis various surgical interventions (7), there is no consensus as to the best although surgical option appropriate, effective and prompt intervention of antimicrobials is desirable when the microbial agents are cultured from patients. Reports from the literature indicate paucity of information on the incidence of chronic osteomyelitis cases in Ile-Ife and its environs except for a study undertaken Ogunjumo by 1981(6). The present prospective study reports on the current status

of this condition in Ile-Ife and its with environs regard ťΟ profile, patients predisposing factors and the treatment modalities in an effort to assist clinicians in the effective management of this condition to reduce cost and period hospitalization of these patients.

MATERIALS AND METHOD Study population

Criteria for inclusion in establishment study were: osteomyelitis by radiography and only patients who presented at the hospital and were willing submit themselves for treatment were included. All patients described above in all age groups were considered, 82 consecutive in-patients who satisfied these criteria were admitted into the study. Their ages ranged from 6 months to 90 years, of which 58 were males and 24 were females. The study was conducted between April 1995 to April 1997 at the Obafemi Awolowo University Teaching Hospital Complex (OAUTHC) Ile-Ife, Osun State. The SouthWestern Nigeria. complex is a referral centre for more than half a million people in Ile-Ife and its environs.

Collection of samples

Samples were collected by clinicians from each patient either during routine visit to the hospitals or at the operating table with sterile cotton-tipped applicators or sterile needles and syringes. At least 2-5 ml of venous blood was similarly obtained from each patient. All samples were collected with care to prevent any contamination during transit. Some of these specimens were introduced into transport medium containing Brain Heart Infusion (BHI) broth and Thioglycolate broth (TB) (Difco, Detriot, Michigan, USA).

Processing of samples

Each sample was collected with sterile cotton-tipped applicators and applied unto freshly prepared Mannitol Salt Agar (MSA), Blood Agar (BA) plate and Eosin Methylene Blue (EMB) agar. Each sample was also introduced into Cooked Meat medium and Thioglycolate broth (TB). inoculated plates were thereafter incubated at 37°C for growth. Approximately 2 ml of venous blood was obtained from each subject and introduced into Thioglycolate broth (TB), which was subsequently incubated at 37°C for growth. The bacterial colonies that grew on the plates were picked and classified into different groups based on their Those colonies Gram reaction. resembling staphylococci were inoculated onto Mannitol Salt Agar and those isolates that fermented mannitol were picked. These were tested for coagulase production using both the slide and tube agglutination test employing pooled

human plasma. Gram-negative isolates that grew on EMB agar were further tested on Triple Sugar Iron (TSI agar), Koser's Citrate and Sulphide Indole Motility (SIM) agar and further characterized on other conventional media.

Antibiotic susceptibility test

Antibiotic susceptibility of organisms the isolated determined by the standard agar dilution method of Ericsson and Sherris (8). The antibiotic discs employed included Ampicillin (Amp) 10 ug, Chloramphenicol (Chl) 10 µg, Cloxacillin (Cxc) 5 µg, Erythromycin (Ery) 5 µg, Nalidixic acid (Nal) 10 µg, Gentamicin (Gen) 10 ug, Penicillin (Pen), 1 unit, Ofloxacin (Ofx)10 10 µg Streptomycin (Str) and Tetracycline (Tet) 10 ali μg, obtained from Abtek Biological Ltd, Liverpool, U.K. Lot 5B02/P. The test medium was Mueller-Hinton agar (Oxoid, UK). S **ATCC** 25923 aureus and **ATCC** Enterobacter aerogenes 13042 were run control* as organisms.

RESULTS

A total of 82 patients were seen. 58(70.7%) were males and 24(29.2%) were females. The incidence of chronic osteomyelitis was lowest among subjects aged fifteen years and below with 11(13.4%) patients, of whom

8(72.7 %) were males. Overall the incidence of the condition was more than double in males as against the females (See Table 1).

Table 2 shows the types of bone commonly involved in cases of chronic osteomyelitis during the study. Forty-six (56.1%) of these involved the tibia followed by femur (13.4%), humerus (8.5%), ulna (6.1%) and radius (4.8%).

The type of surgical interventions carried out on the patients is shown in Table 3. Wound debridement constituted a proportion of surgery occurring in 32 cases followed by sequestrectomy, saucerization and curettage in 22 cases. Local muscle for 10; accounted grafting, skin grafting and antibiotic impregnation constituted 2 cases each, while plating was done for non-union in only one case. The sources of infections in the chronic osteomyelitis cases varied. Posttraumatic sources were seen in 50 for cases and accounted а considerable of proportion infections. Decubitus ulcers contributed 5 cases. This was followed by diabetic foot with 4 cases. Leg ulcer, pin-tract infection, and crush injury accounted for 3 each. Gunshot cases injuries occurred in 2 cases and acute osteomyelitis occurred in only one case.

Bacterial isolates cultured from chronic osteomyelitis specimens

Seventy-eight bacterial isolates were cultured from 82 cases of chronic osteomyelitis microbe averaging 1.05 per Gram-positive subject. cocci constituted 44.9% of the entire isolates. Staphylococci accounted for 97.1% of these isolates. Among the staphylococci, Staphylococcus accounted for 47.1%, coagulase negative staphylococci (CONS) 29.4% and Staphylococcus aureus 23.5%. Gram-negative rods accounted for 55.1% of bacterial isolates with Proteus mirabilis predominating (41.9%) followed by Pseudomonas aeruginosa (25.9%), Escherichia coli (13.9%), Klebsiella aerogenes (9.3%) and Citrobacter freundii and Salmonella constituting 4.6% each of the Gram-negative short rods. None of the blood culture showed any growth.

Antibiotic susceptibility profile of some of the bacterial isolates

The antibiotic susceptibility pattern of the bacterial isolates is shown in Table 6. The staphylococci all virtually resistant Benzylpenicillin and Ampicillin. All the Staphylococcus aureus and CONS isolates were resistant to these two penicillins but sensitive to Cloxacillin and Ofloxacin both of which are beta-lactamase resistant drugs. Pseudomonas aeruginosa isolates were resistant to all drugs used except Gentamicin which 45% of P. aeruginosa isolates were sensitive. similar Α trend resistance was seen with Proteus mirabilis and E. coli isolates (Table 6).

TABLE 1: PROILE OF CHRONIC OSTEOMYELITIS CASES SEEN AT OAUTHC BETWEEN 1995-1997

Age	Number (%)	Number (%)	
	Male	Female	
0-15	8(13.8)	3-(12.5)	
16-30	20(34.5)	10-(41.7)	
31-45	17(29.3)	5-(20.8)	
>46	13(22.4)	6 - (25.0)	
Total	58 (70.7)	24 (29.2)	

TABLE 2: TYPES OF BONE INVOLVED IN CASES OF CHRONIC OSTEOMYELITIS AT OAUTHC, ILE-IFE

BONE TYPE	FREQUENCY (%)
Tibia	46(56.1)
Femur	11(13.4)
Humerus	7(8.5)
Ulna	5(6.1)
Radius	4(4.8)
Phalanges	2(2.4)
Sacral Bone	2(2.4)
Tarsal	1(1.2)
Clavicle	1(1.2)
Клее	1(1.2)
Elbow	1(1.2)
Ribs	1(1.2)

TABLE 3: TYPES OF SURGICAL INTERVENTIONS EMPLOYED AT THE OAUTHC, ILE-IFE

PROCEDURE	FREQUENCY
Sequestrectomy, saucerization and curettage	22
Wound debridement	32
Local Muscle flaps	10
Amputation	7
Drainage of Pus	5
Bone grafting	2
Skin grafting	2
Antibiotic impregnation	2
Plating (after control of infection)	1

TABLE 4: SOURCES OF INFECTION IN CHRONIC OSTEOMYELITIS AT OAUTHC, ILE-IFE

ROUTE OF INFECTION	FREQUENCY
Post traumatic	50
Decubitus nicers	5
Diabetic foot	4
Leg ulcer	3
Pintract infection	3
Sacral pressure (bed sore)	3
Crush injury	2
Gunshot injury	2
Acute osteomyelitis	1

: .

TABLE 5: BACTERIAL SPECIES ISOLATED FROM CHRONIC OSTEOMYELITIS PATIENTS AT THE OAUTHC, ILE-IFE

BACTERIAL ISOLATES	FREQUENCY (%)
Gram Positive	
Staphylococcus spp.	16(20.5)
Coagulase nagative Staphylococci (CONS)	10(12.8)
Staphylococcus aureus	8(10.0)
Streptococcus spp.	1(1.2)
Gram Negative rods	
Proteus mirabilis	18(23.1)
Pseudomonas aeruginosa	11(14.1)
E.coli	6(8.5)
Klebsiella aerogenes	4(5.1)
Citrobacter freundii	2(2.5)
Salmonella spp.	2(2.5)
Total	78

TABLE 6: ANTIBIOTIC RESISTANT PROFILE OF BACTERIAL ISOLATES

Organism	Total No. Tested	% Resistant								
	rested	PEN	AMP	ERY	CHL	STR	TET	GEN	схс	OFX
Staphylococcus aureus	8	100	100	450	25	25	100	25	0	0
Staphylococcus spp	16	75	44	13	44	100	100	13	0	0
Coagulase Negative Staphylococci (CONS)	10	100*	100	20	20	20	100	0 ; * ,	0	0
Streptococcus spp	1	0	0	0	0	0	0	0	0	0
Pseudomonas aeruginosa	11	100	100	100	100	100	100	45	0	0
Proteus mirabilis	18	100	100	100	100	100	100	33	0	0
Klebsiella aerogenes	4	100	100	100	100	100	100	25	0	0
Escherichia coli	6	100	100	100	100	100	100	33	0	0
Salmonella spp	2	100	0	0	O 1,	0	ó	0	Ó	0

^{*} Only major organisms were tested

DISCUSSION

This work was prompted paucity of the because regarding chronic information osteomyeliitis in Ile-Ife and its environs. Except for the study carried out by Ogunjumo in 1981 (6), there is no other report of chronic osteomyelistis in this environment. Unlike acute osteomyelitis. substantial \mathbf{a} proportion of the patients with chronic osteomyelitis were adults (78%) and a number of these were males (67%)indicating predominance of this condition amongst males (2.41:1). Our results show that the tibia was the affected predominant bone followed the femur bv humerus. The majority of the chronic osteomyelitis cases and others were as a result of posttraumatic infections due compound fracture, crush injury and pin tract infection. Although other organisms have been implicated chronic osteomyelitis, bacterial species are the most common. The bacterial aetiology of chronic osteomyelitis varies but according to most Staphylococcus studies (9,16),aureus to be the seems predominant organism encountered. In a study carried out at this Centre by Ogunjumo (5), S. aureus was the predominant organism isolated from patients. While staphylococci still remain the significant organisms cultured from chronic osteomylitis patients, study reveals that our Staphylococcus spp (20.5%) has become the predominant Gram positive cocci recovered, followed by coagulase negative staphylococci (CONS) (12.8%). S. aureus came a distant third (10.3%).This observation underscores the changing pattern of bacterial actiology of this condition even within the same environment.

However, Gram negative short rods constituted 55.1% of the total bacterial isolates cultured various specimens with from Proteus mirabilis accounting 41.9%) of the Gram negative rods. This was followed by Pseudomonas aeruginosa (25.9%). Overall, Proteus mirabilis was the predominant bacterial isolates chronic in osteomyelities at this centre. While Ogunjumo (6) previously reported S. aureus as the predominant organism in chronic osteomyelitis at this centre, our study however, corroborates Oguachuba (10) who reported Proteus spp predominance Northern Nigeria. in Jos, The increasing prominence of Pseudomonas aeruginosa in chronic osteomyelitis is of concern because of this organism resistance antimicrobial various agents. However, none of the subjects from whom pseudomonas were cultured had sickle cell disease (11). An average of two types of organisms

each was cultured from patients that participated in the Seven of the specimens cultured did not grow on all the media employed. The occurrence of multiple organisms specimens cultured, in some suggests polymicrobial nature of chronic osteomyelitis appears to be a feature of this condition compared with acute osteomyelitis cases, where over 50% of clinical specimens cultured contained a single organism (12).

Many factors other than the age affect the development of chronic osteomyelitis (1.5, 13).Osteomyelitis resulting from direct extension of a decubitus ulcer. wound infection or open fracture may be difficult to detect clinically before the infection progresses to .chronic osteomyelitis. Therefore early recognition of osteomyelitis is highly desirable for favourable prognosis. Even roentgenographic when abnormalities of the bone may be helpful in confirming the presence of infection, if roeritgenographically diagnostic changes are delayed, the process may often progress to chronic form. In patients with chronic recurrent osteomyelitis, antimicrobial therapy is usually dictated by the microflora present within the infected bone. Even when cultures of sinus drainage yield bacteria, which are

not present in the bone or fail to yield organisms, such cultures of sinus tract are only useful as guidelines for initial therapy.

The selection of antimicrobial agents for the treatment of chronic osteomyelitis caused by Gramnegative rods may be difficult. The in vitro sensitivity testings carried out with these isolates underscores this point. Virtually all the gramnegative rods recovered from the clinical source were multiply resistant to the various antibiotics used at least in vitro. Except for four isolates of Staphylococcus spp, all the other staphylococcal isolates were resistant to Benzylpenicillin and Ampicillin, suggesting these beta-lactamase were producers. Such observation with S. aureus isolates had been reported Mollan and Piggot (12), in which seventy-four percent of were resistant strains to Benzylpenicillin. Gentamicin was marginally effective in vitro with only five isolates of Pseudomonas aeruginosa sensitive. Similarly the Proteus mirabilis recovered from the patients were resistant to most antibiotics used in treating Graminfections negative except Gentamicin. The ineffectiveness of many of these antibiotics in vitro against the majority of our isolates corroborates reports of investigators of the existence of multiply resistant enteric rods even amongst apparently healthy

individuals in this environment and elsewhere (14,15).

We recommend that this clinicians practicing in environment should be mindful of this phenomenon and therefore employ in their treatment modalities as combination effective drugs and also follow-up patients' treatment considerable length of time in order to resolve the condition. According to Jensen and Lassen Fusidic acid given combination with other antibiotics effective preventing were in emergence of resistant strains of S.aureus when administered as first line treatment. Ĩt interesting that the bacterial isolates cultured in our study responded very well to Cloxacillin and Ofloxacin both of which are known to be effective against betalactamase producers.

While antimicrobial therapy is desirable in the control of chronic osteomyelitis, surgery remains the therapeutic and diagnostic procedure, which should be carried out early to resolve the condition (1,18). Our therefore study recommends detection of prompt acute osteomyelitis cases so that these do not progress to chronic form. chemotherapeutic Aggressive of management acute osteomyelitis is suggested. Early bacteriological investigations

determine bacterial aetiology are highly desirable so that first line antimicrobials are rigorously administered. Since the majority of chronic osteomyelitis cases are posttraumatic, orthopaedic surgeons should be more conscious the risk of such injuries progressing to chronic forms and educate patients of prompt and proper management at initial stages to avert subsequent complications. prevalence of The multiply antibiotic resistant organisms in this study should be brought to the notice of Health Administrators in the Ministry of Health who should educate the public and dissuade communities in engaging in selfmedication.

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REFERENCES

- Ofiaeli RO. Chronic osteomyelitis in Nigerians. Experience with policy of microbial evaluation.
 Orient J. Med. 1993; 5:87
 –90.
- 2. Ofiaeli RO. Radiological features of chronic osteomyelitis in long bones and the effects of diaphysical sequestration on linear growth. Orient J. Med. 1991; 3:20-22.
- 3. Epps CH, et al.
 Osteomyelities in patients
 who have sickle cell disease.

 J. Bone Joint Surg.
 1991; 75: 1281-1293.
- Adeyokun AA, Hendrickse 4. RG. Salmonella osteomyelitis in childhood-A report of 63 cases 1000 seen in Nigerian children whom 57 had sickle cell anaemia. Arch. Dis. 1980 ; **55** : 175 Childhood - 184.
- 5. Ebong WW. Acute osteomyelitis three years after a close fracture in an adult with sickle- cell anaemia. J. Bone Joint Surg. 1980; 62-A(2): 1196 1198.

- 6. Ogunjumo DO. Aetiology and control of chronic pyogenic osteomyelitis in Ile-Ife, Nigeria. *Tropical Doctor*. 1981; 11:155-159.
- 7. West WF, Kelly PJ, Martin WJ. Chronic osteomyelitis 1: Factors affecting the results of treatment in 186 patients. *JAMA*. 1970; 213: 1837-1842.
- Ericsson HM, Sherris JC. The agar dilution method. Acta. Pathol. Microbiol. Scand. 1971
 ; 217 (Sect. B.Suppl): 11-19.
- 9. Essien IAJ, Ndukwe AU.
 Chronic osteomyelitis A 12month experience at the
 National Orthopaedic
 Hospital, Enugu. Nig.
 Postgrad. Med. J. 1994; 1 (3)
 : 22-25.
- Oguachubam HN, Egere JU.
 The bacteriology of osteomyelitis in sicklers in Eastern and Northern Nigeria. Nig. Med. J. 1989;

 19(3): 127-129.
- Oguachubam HN. Hospital infection in Orthopaedic Traumatological Department of the Jos University Teaching Hospital. Nig. Med. J. 1985; 9(4): 99-101.

- Mollan RAB, Piggot J. Acute osteomyelitis in children. J. Bone Joint Surg. 1977; 59-B(1): 2-7.
- 13. Overton CM, Tully WP. "Surgical Treatment of Chronic Osteomyelitis in Long Bones". Am. J. Surg. 1973; 126:736-741.
- 14. Lamikanra A, Ako-Nai AK, Ogunbiyi DA. Transferable Antibiotic Resistance in Escherichia coli isolated from Healthy Nigerian, School Children. Int. J. Antimicrob. Agents. 1996; 7:59-64.
- 15. Lamikanra A, Ako-Nai AK, ... Ola JB. Transmissible Trimethoprim resistance in strains of Escherichia coli isolated from cases of infantile diarrhoea in Nigerians oral rehydration therapy clinic. Ann. Trop. Paediatr. 1989; 9: 256-260

- 16. Lester SC, Pla MP, Wang F, et al. The carriage of Escherichia coli resistant to antimicrobial agents by healthy children in Boston, Caracas, Venezuela and Qui Pu, China, N. Engl. J. Med. 1990; 323:
- Jensen K, Lassen HCA.
 Combined Treatment with Antibacterial
 Chemotherapeutical Agents in Staphylococcal Infections.
 Quat. J. Med. 1969; 38: 91-106.
- McNally MA, Small JO, Tofighi HG, Mollan RAB. Twostage Management of Chronic Osteomyelitis. The Long Bones. J. Bone. Joint. Surg. 1993; 758: 375-380.

STUDIES ON DIABETIC FOOT ULCERS IN PATIENTS AT JOS UNIVERSITY TEACHING HOSPITAL, NIGERIA

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An epidemiological and microbiological studies of diabetic foot ulcers were carried out in our hospital, with a view to reducing the amputation and mortality rate associated with the disease. Wound swabs from 38 Diabetes Mellitus (DM) foot ulcer patients were investigated using culture methods for both strict aerobes and anaerobes. The bacterial isolates were subjected to antibiotic susceptibility tests using the disc diffusion method. Baseline biochemical and haematological analysis were also carried out. The prevalence of the disease was stratified in relation to some clinical and laboratory parameters, gender, age, educational and occupational status of the patients. The prevalence of the diasese was 24.7%, with an amputation and mortality rates of 18.4% and 15.8% respectively. Only 13% had DM for less than 1 year, while 53% for more than 10 years. 28.9% have regular shoe-wearing habits. Duration of healing ranged from 2 weeks to 24 weeks (mean = 2.7months). 31% of the patients with marked periosteal reaction had lower extremity amputation or died before amoutation could be done. Staphylococus aureus (31%), Proteus spp (16%), Pseudomonas aeruginosa (10%), Klebsiella spp (6%), Peptococcus spp (6%), Bacteroides fragilis (3%), Streptococcus pyogenes (3%), Escherichia coli (3%), Candida albicans (3%), Streptococcus viridans (1%), Flavobacterium spp (1.5%) and Bacteroides melaninogenicus (1%) were isolated. Most of the bacteria isolates were sensitive to pefloxacin. Our results demonstrate a very high rate of diabetic foot ulcer with the corresponding high rate of amputation and mortality. A multi-disciplinary approach to the management of DM foot ulcers is advocated.

Efforts should be made to carry out cultures of samples from refractory ulcers to rule out yeast colonization which if not treated will delay wound healing.

Key words: Diabetic foot ulcers, Microbial and antimicrobial surveillance, Refractory ulcers.

INTRODUCTION

is **Mellitus Diabetes** increasingly being recognized as a major medical problem in our environment, and the threat of lower extremity amputation (LEA) cast's an ominous shadow over the lives of diabetic patients. Diabetic ulcers are the most common foot lesions leading to lower extremity amputation (1). Management of diabetic foot requires thorough knowledge of the major factors for amputation, frequent routine evaluation and meticulous preventive

maintenance (2). The etiology of lower extremity diabetic ulcer includes injury complicated by underlying neuropathy, ischaema or both. The use of recombinant growth factor is becoming more widespread as more is learnt about their essential role in wound healing (3).

According to a study by Bild (4), more than 50,000 amputations were performed on diabetic patients each year in the United States of America. Reported prevalence rates of diabetic foot ulcers in different

groups of Nigeria diabetics range from 0.9% to 8.3% (5-8). complication led to limb amputation in 40% of cases (8). Foot infections caused a third of all skin infections in 1,050 Nigeria diabetics (9), and accounted for 25% of deaths in ketocidosis (8). In a study by Walters et al (10) in United the Kingdom, the diabetic foot prevalence of ulceration was 7.4%.

Psychosocial problems from limb amputation persistently retard rapid rehabilitation. Nigeria with inadequate health care facilities and uniform lack of comprehensive rehabilitation progammes, it is important to recognize foot lesions early and thereby offer early and effective treatment. **Amputation** expensive (about \$500 per case, including the not costs and there rehabilitation) is difficulty psychological readjustment.

When managing diabetic foot ulcers, one is faced with the dilemma of whether to opt for radical surgery or to continue with conservative measures. We therefore, decided to carry out a prospective study to see how we can effectively. reduce amputation rate and the mortality associaed with diabetic ulcers in our environment.

PATIENTS AND METHODS Patients

One hundred and fifty four from diabeties were admitted January 1998 to December 1999 for various reasons ranging from diabetes poor control of septicaemia. All were prospectively studied. Each of the patients with foot diabetic ulcer(s) interviewed using the close ended questionnaire and then examined clinically. For example, with respect to shoe wearing habit, patients were asked whether it is regular or none. In this study, diabetic is defined according to WHO guidelines of glucose more than 7.0 mmo1/L (>126 mg/dl) for fasting blood sugar or more than 11.1 mmo1/L (>200 mg/dl) 2 hours postprandial (11).

Bacteriology

The wound swabs were collected as much as possible before any antibiotic therapy and inoculated immediately onto freshly prepared Blood Agar (BA), Chocolate (Choc.) Agar and MaConkey (McC) Agar plates. Preheated Robertson's Cooked Meat medium (RCM) was also inoculated and incubated aerobically at 35°C. One set of BA, Choc and McC agar plates was incubated aerobically at 35°C for 18-24 hours. The other set was incubated anaerobically at 35°C for 48 to 72 hours. The RCM broth cultures were subcultured after 48 hours and 72 hours onto the solid media and incubated appropriately. The identifications of the isolates were done using the standard methods (12). Antibiotic susceptibility tests were carried out on the pathogenic isolates using the NCCLS techniques (13).

Haematology

Full blood count and Erythrocyte Sedimentation Rate (ESR) values were carried out on the patients' blood samples collected on admission (14).

Chemical pathology

Short-term glycaemic control was assessed by fasting glycosuria, acetonuria and fasting blood glucose levels. Urinalysis of the patients' samples were carried out using the MediTest strips and proteinuria was defined as the presence of greater than 50 mg/dl of protein.

Plain foot radiographs

Serial plain radiographs on the affected limbs were carried out on 32 of the 38 patients with diabetic foot ulcers. Two of the six patients that did not do X-ray could not afford the cost and their ulcers healed within six weeks, while the remaining four had above knee amputation immediately they were brought to the hospital so as to save their lives.

Surgery

Prophyhlactic surgery was done when necessary by serial debridement and large lesions were skin-grafted. In a few cases, where the life of the patients was in danger, amputation (especially above the knee) was inevitably carried out.

Antibotic therapy

The patients were treated based on the *in vitro* susceptibility pattern of the isolates, in addition to Metronidazole, Vitamin C, Vitamin B-complex and Insulin therapy. This was in addition to the scrupulous twice, daily dressing of the ulcers with hydrogen peroxide and eusol after warm hypertonic saline immersion of the affected whole foot for 30 minutes

RESULTS

Table 1 shows the duration of diabetes mellitus at the time of presentation to the hospital with diabetic foot ulcers. Only 5 (13%) of the 38 patients studied had duration of diabetes mellitus for less than one vear. All the ulcers were located on the hind limbs and 7 (18%) had bilateral ulcers on both hind limbs while 31 (82%) had unilateral ulcers. The duration of healing ranged from 0.5 month to 6 months with a mean of 2.73 months. None of the patients was hospitalised for the entire duration of the healing. As soon as the blood glucose level is stabilised, they were discharged and then followed up in an ambulatory setting. The fasting blood glucose at the time of admission ranged from mmol/L to 28 mmol/L. The packed cell volume (PCV) ranged from 18% to 45%; white blood cell (WBC) count $3,500/\text{mm}^3$ ranged from 15,700/mm³; differential neutrophil

counts ranged from 48% to 78%; lymhocyte counts from 19% to 50% and the erythrocyle sedimentation rate (ESR) ranged from 6 mm/hour to 150 mm/hour. All these were the baseline data at the time of admission.

The plain foot x-rays showed periosteal reaction, osteomyelitis and soft tissue swelling. A few of the x-rays were normal while 31% of the patients who had marked periosteal reaction had their limbs amputated or died before amputation could be done

The sex distribution of the diabetes patients that attended the hospital for the 2 year period is shown in Table 2. There is no significant difference in prevalence of diabetic foot ulcers in relation to gender (p>0.05). For the 2-year period, the prevalence of the diabetic foot ulcers in our hospital was 24.7%. Table 3 shows the prevalence of some important clinical and laboratory parameters in the patients with diabetes foot ulcers while Table 4 shows the prevalence in relation to the ages of the patients. No ulcer was seen in patients of less than 20 years and more than 80 years of age while the · 51-60 years group had the highest with 44.7%.

The prevalence of diabetes foot ulcers in relation to educational status of the patients is shown on Table 5. Patients with no formal education recorded the highest with 36.8% (P<0.05). Table

6 shows the prevalence of the diseases in rotation to the occupational status of the patients. Civil servants recorded the highest with 39.2%, but there is no statistical difference (P>0.05).

The microorganisms isolated from the diabetic foot ulcers foot ulcers are shown on Table 7. Twenty-two (58%) of the patients poly-microbial flora, 4(6%) showed no growth and strict anaerobes, aerobes and yeasts were encountered. Table 8 summarizes the percentage susceptibility patterns of the bacterial isolates to some of the commonly prescribed antibiotics in our hospital. Most of the isolates were sensitive pefloxaclin, except for Klebsiella spp that recorded only 25% sensitivity.

The summary of the laboratory comparing patients ulcers healed and those who went home for amputation or died is shown in Table 9. There were no significant differences in the haematocrit values, white blood cell erythrocyte sedimentation counts. rates, differential neutrophil counts, fasting blood sugar and fasting glycosuria first on visits. Only 8(21.1%) of the patients had acetonuria on their first visit; out of which 7(87.5%) under went amputation or died.

Table 1: Duration of Diabetes Mellitus at the time of presentation with ulcer.

1 (1)

Duration (years)	No of Patients	% of Total
<1	5	13
1-5	7	18
6-10	6	16
>10	20	53

Table 2: Sex Distribution of Diabetic Foot Ulcers in JUTH [January 1998-December 1999].

Admission	Males	Females	Total
Total DM	96	58	154
Total DM Foot	24	14	38
Relative frequency of	25%	24.1%	24.7%
DM Foot			

Table 3: Prevalence of some Important Clinical and Laboratory Parameters in the Patients.

Parameter	Number	Number	%Positive
	Examined	positive	
Positive knowledge of foot care.	38	11	28.9
Regular shoe wearing habit	38	30	78.9
Presence of previous foot ulcer	38	12	31.6
Presence of unilateral lesion	38	31	81.6
Spontaneous initiating factor	38	30	78'9
Presence of foot pain	38	26	68.4
Presence of shooting or lancinating	38	27	71.1
pain	38	24	63.2
Presence of pain at rest	38	11	28.9
Presence of intermittent claudication	37	14	36.8
Presence or nocturnal diarrhoea	38	15	39.5
Presence of postural dizziness	24	21	87.5
Presence of impotence	38	22	57.9
Presence of fever	38	8	21.1
Presence of acetonuria	38	24	63.2
Presence of albuminuria	38	7	18.4
Amputation	38	6	15.8
Mortality		1	
•			

Table 4: Prevalence of Diabetes Foot ulcers in relation to Age at JUTH, Nigeria

Age Group (years)	Number with Ulcer	% with Ulcer
21-30	2	5.3
31-40	4	10.5
41-50	9	23.7
51-60	17	44.7
61-70	4 .	10.5
71-80	2	5.3
	.	

Tables 5: Prevalence of Diabetes Fogt in Relation to Educational Status

Educational Level	No. of Ulcer	% with Ulcer
None	14	36.8
Primary School	10	26.3
Secondary school	6	15.8
Tertiary Institution	8	21.1

Table 6: Prevalence of Diabetes Foot Ulcers in Relation to Occupational Status of the Patients

Occupational Status	No with Ulcer	% with Ulcer
Farmers	5	13.2
Traders	8	21.1
Civil servants	13	39.2
House-wives	7	18.4
Others (like Butchers, motor	5	13.2
mechanics, applicants)		

Table 7: Microbial Isolates from Diabetic Foot Ulcers at JUTH, Nigeria

Microorganism	No of Isolates	% of Isolates
S. aureus	21	31
S. pyogenes	. 2	3
S. viridans	i	- 1
Cl. perfringens	4	6
Peptococcus spp	4	6
Klebsiella spp	4	6
Proteus spp	11	16
B. fragilis	2	3
B. metaninogenicus	1	1
Flavobacterium spp	1	1
P. aeruginosa	7	10
Candida albican	2	3
Acinetobacter spp	1	1
Esherichia coli	2	3
No growth	4	6

Table 8: Percentage Susceptibility Patterns of the Bacterial Isolates to Clindamycin (CLN), Augumentin (AMC), Ampicillin (PN), Cotrimoxazole (SXT), Tetracycline (Te) and Pefloxacine (Pef).

Antibiotics							
	CLN	AMC	PN	SXT	Te	Pef	
Bacteria		_				_	
S. aureus	29*	48	0	10	48	90	
S. pyogenes	100	100	100	0	Ó	100	
S.viridans	100	100	100	0	0	100	
CL. Perfringens	75	100	50	0	75	100	
Peptococcus sp.	100	75	25	0	25	100	
Klebsiella sp.	NA+	25	0	25	0	25	
Proteus sp	ŇA	36	0		9	73	
B. fragilis	50	50	50	50	50	100	
B. melaninogenicus	0	0	0	0	0	100	
Flavobacterium sp.	0	O	0	0	100	100	
P. aeruginosa	NA	NA	NA	NA	NA	86	
Acinetobacter sp.	NA	0	0	0	0	100	
E. coli	NA	100	0	0	0	100	

^{* %} susceptible

Table 9: Laboratory Parameters of Patients with healed Diabeti Ulcers
Amputated/Dead Patients.

	PCV (Mean + SE)	WBC (Mean + SE)	ESR (Mean + SE)	Neutrophils (Mean + SE)	FBS (Mean + SE)	Aceton uria Num- ber	PG Number
Healed Ulcers	37.88_+1.155	6816 <u>+</u> 558.62	41,167 <u>+</u> 6.574	57.12 <u>+</u> 1.365	12.748±0.832	01	21
Limb/death P-Value	30.538+1.96	1111±1086.623	81.154±10.837	67.846±2.412	19.269 <u>+</u> 1.45	07	13
(using kruskal-							
Wallis ANOVA	P > 0.05	P > 0.05	P > 0.05	P > 0.05	P > 0.05	P>0.05	P > 0.05

DISCUSSION

This study has shown a relatively high prevalence rate of diabetic foot ulcers (24.7%) among the DM patients in our hospital with 18.4% and 15.8% amputation and mortality rates. The population was limited to patients who attended the hospital during the study period either as in patients or ambulatory patients. It is very likely that the burden of diabetic foot ulcers may be more

in the general population (15 years and above) within the metropolis with a DM prevalence of 3.1% (16). We believe that a combination of conservative approach and aggressive prophylactic surgery in high-risk patients, if started very early in the course of the disease will reduce the amputation and mortality rates in our environment, which hitherto was high. Previously many limbs that should have been

⁺ Not applicable.

saved were amputated without combination trying the of conservative approach and aggressive prophylactic surgery. Also some DM foot ulcers patients shy away from attending the hospital for fear of amputation and eventually some of them die due to infections. We believe that with adequate health education. which should be given to the DM patients particularly on their clinic days, they will be willing to report any ulcers to the physician no matter how trivial. **Proper** management of the ulcers includes the use of proper footwear, non-weight-bearing limb support, use of appropriate antibiotics, debridement. aggressive revascularization. control of serum glucose levels and careful monitoring of the **Empirical** ulcers. antibiotic selection was replaced by cultureguided definitive therapy as soon possible and this greatly improved the condition of the patients. It was discovered in the study, that the prevalence of diabetic ulcers was low in those with positive knowledge of foot and this care confirms the of advantage regular health education for the diabetics. The of the disease prevalence relation to age is in agreement with the known knowledge that diabetes mellitus is highest in the 51-60 year age group. A few

diabetic with no formal education had the highest prevalence of ulcers due mostly to ignorance.

Some ofthe important parameters which are relevant to development of foot ulcers diabetic patients such as peripheral neuropathy, peripheral pulses. presence of calluses, blisters, corns and trophic changes are being prepared for publication. Although new therapies, such as the use of recombinant exogenous growth factors are being developed for refractory ulcers (17), care must be taken to isolate and identify all the microbial flora of the ulcers.

The bacterial isolates showed a high rate of mulitiple resistance to commonly used antibiotics. This is mainly due to the ready availability and the easy access access to those antibiotic by the general population without medical prescription. It is most likely that majority of the Staphylococcus aureus strains isolated in this study are MRSA as earlier reported by Ikeh (18), who got a prevalence of 43% out of the 180 consecutive isolate of S. aureus in our hospital. In this study, we encountered few isolates of Candida albicans as part of the mixed microbial flora and the patients responded very well on the inclusion of ketoconazole to the treatment regimen. Diabetic ulcers shoud be thoroughly evaluated and appropriately treated through established protocols utilizing all members of the team. This certainly

reduces the duration of hospital admission, morbidity and loss of limbs. Although foot problems in diabetics cannot be eradicated completely, it is always better to diagnose and manage diabetic ulcers effectively, educate and motivate patients to care for their so as to minimize complications and decrease health care costs. In the absence of facilities for culture and antibiotic susceptibility tests, pefloxacine may be useful as a monotherapy in diabetic foot ulcers due to the marked clinical and laboratory reponse in our study.

Except for acetonuria, none of the conducted laboratory parameters determined outcome of the diabetic foot ulcers This shows that with disease. proper managemnet of the patients, the morbidity, amputation and the mortality rate can be drastically reduced if the management is started early. The study population was limited to patients who attented the hospital either as inpatients or ambulatory patients, and it is very likely that the burden of diabetic foot ulcers is more in the general population.

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REFERENCES

- 1. Armstrong DG, Lavery LA. Diabetic foot ulcers: prevention, diagnosis and classification. Am. Fam. Physician. 1998; 57(6): 1325-32.1337-38.
- Armstrong DG, Lavery LA, Harkless LB. Who is at risk for diabetic foot ulceration? Clin. Paediatr. Med. Surge. 1998; 15(1): 11-19.
- 3. Krasner D. Diabetic ulcers of the lower extremity: a review of comprehensive management. Ostomy Wound Manage. 1998; 44(4): 56-8,60-2,64.
- 4. Bild DE. Lower extremity amputation in people with diabetic: epidemiolgy and prevention. Diabetes Care. 1989; 12:24.
- Kinner TWG. The pattern of diabetes mellitus in a Nigerian Teaching Hospital. East Afr. Med. J. 1963; 40(5): 288.
- Greenwood BM, Taylor JR. The complication of diabetes in Nigerians. Trop. Geograph. Med. 1968; 20: 1.
- Osuntokun BO, Akinkugbe OO, Francis TI. Diabetes Mellitus in Nigerians: a study of 832 patients. West Afr. Med. J. 1978; 8(5): 295.

- 8. Lawson EAL, Oyemade GAO, Adetuyibi A. The foot complication in the Nigeria diabetic patient. Nig. Med. J. 1978; 8(5): 401.
- 9. Bell ET. Atherosclerotic gangrene of the lower extremities in diabetic and non-diabetic persons.

 Am. J. Clin. Pathol. 1957;
 28:27.
- 10. Walters DP, Gatling WA, Muller MA, Hill RD. The distribution and severity of diabetic foot disease: a community study with comparison to a non-diabetic group. Diabet. Med. 1992; 9(4): 352-58.
- 11. World Health Organization guidelines. Expert Committee on diagnosis and classification of Diabetes Mellitus. Report of the Expert committee on the diagnosis and classification of Diabetes Mellitus. Diabetes Care. 1997; 20: 1183-97.
- 12. EII, Balows A, Lennete WJ, Truant JA. Hausler Manual Clinical of Microbiology 3rd edition. American Society of Microbiology, Churchill Livingstone, Edinburgh and London. Washington DC. 399-416.

- 13. National Committee for Clinical Laboratory Standards. Methods for dilution antimicrobial susceptibility tests for bacteria grow that aerobically. 4th edition Appproved Standard M7- A4, Wayne, PA. NCCLS, 1997.
- 14. Dacie JV, Lewis SM. Practical Haematology, Churchill Livingstone, Edinburgh, 1991:321-468;
- 15. Trinder P.Determination of blood glucose using 4-aminophenazone as oxygen acceptor. *J. Clin. Pathol.* 1969; **22**: 246.
- 16. Puepet FH. The prevalence of diabetes mellitus in adults in Jos, Nigeria. Dissertation for fellowship in Internal Medicine (FMCP)-National Postgraduate Medical College of Nigeria 1996.
- 17. Steed DC. Foundations of good ulcers care. Am. J. Surge. 1998; 176(2A suppl): 1205-255.
- 18. Ikeh EI. A twelve month surveillance of methicilim resistant Staphylococcus aureus (MRSA) at Jos University Teaching Hospital, Nigeria. Afr. J. Clin. Exp. Microbiol. 2003; 4(1): 52-55.

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AETIOLOGIC AGENTS OF DIARRHOEA IN CHILDREN UNDER FIVE YEARS OF AGE IN OSOGBO, OSUN STATE.

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A survey of the aetiological agents of diarrhoea in children under 5 years of age was carried out in Osogbo, Osun State. A total of 135 patients visiting the outpatient and children emergency units of LAUTECH Teaching Hospital were examined. Strains of *Shigella* isolated were tested for antibiotic sensitivity. Consideration of the distribution showed that *Escherichia coli* was more prevalent in children aged between 3 to 5 years (57%) and was the most encountered of the organisms isolated (present in 77.8% of all samples) white *Shigella spp* was found in 16.3% of samples, *Vibro cholerae* 0.7% and other coliforms 5.2%. Statistical analysis showed that *E. coli* was significantly associated with diarrhoea in the patients aged 3-5 years (P< 0.05). *Shigella spp* was shown to show some resistance to tetracycline and high sensitivity to ofloxacin.

INTRODUCTION

Diarrhoea is a leading cause of morbidity and mortality among children in developing countries (1). It has been found to be associated with bacteria, viruses However the and protozoa. pathogens most commonly associated with children diarrhoea werefound to include E. coli, Vibrio spp, Salmonella, Shigella spp, and parasitic agent, Giardia lamblia (3). Rotavirus has been implicated recent study, being more pronounced during the dry season (4). Some factors such as poor food hygiene, water and sanitation combine to facilitate the spread of enteropathogens and epidemics in such areas (5). E. coli has been implicated as the prevalent aetiological agent of Infantile

Nigeria; exhibiting diarrhoea in numerous strains(6). epidemiological significance of each E. coli strain in childhood diarrhoea varies with geographical area (7). Although extensive investigation of diarrhoea in Nigeria have not been the diarrhoea-specific reported. mortality in children less than 5 years of age in Africa has been estimated at about 10 6 per 1000. Although oral rehydration has been shown to reduce early child mortality (8), it is a general belief that malnutrition may increase the duration or severity of diarrhoea in young children, while diarrhoea lead to symptoms malnutrition in them (8). This study investigates the aetiological agents of childhood diarrhoea in Osogbo and their susceptiblity to some antibacterial drug.

MATERIALS AND METHODS Study area/ sample source

The study was carried out at the children and emergency and out patient departments of the Teaching Hospital, LAUTECH CHS Osogbo. Stool samples were collected from patients presenting with diarrhoea into sterile sample bottle. One hundred and five stool samples were collected over a period 10 months (from middle of July 2001 to April 2002) and analyzed.

Microbial Analysis

The samples were examined for signs of diarrhoea (9). Tenfold serial dilution was carried out on those samples found consistent with the description for diarrhoea using sterile normal saline as diluent. 1.0mlof appropriate dilutions were inoculated onto Thiosulfate-Citrate-Bile Salt-Sucrose agar medium. Salmonella-Shigella agar medium and MacConkey agar, according to described method (6). Rectal swab specimens were directly inoculated onto each agar plate by routine techniques. The colonies that grew on each medium were examined and the organisms characterized. The Shigella strains isolated were tested for susceptibility to some antibiotics (ampicillin,

tetracycline, cefdinir, ofloxacin and erythromycin.) and the minimum inhibitory concentration (MIC) of each drug was determined by plate dilution technique.

RESULTS

All the samples were positively identified as diarrheal Bacteriological analysis showed that of a total of 135 isolates recovered from the stool samples; 77 were strains Escherichia coli, others are Shigella 22, Vibrio cholerae 1, and other coliforms (Table 1). 7 consideration of the age distribution shows that E. coli was more prevalent in children aged between 3 to 5 years (57%) and was most encountered of the organisms isolated. A strain of Vibrio cholerae was also observed in a sample from children within the ages of 3-5yrs. Seasonal variations in incidence of bacterial diarrhoea is shown on Table 2. It was found to be prevalent during wet season (P>0.05). A total of 93 (68.9%) isolates were recovered during the wet season compared to 42 (31.1%) recovered in the dry season. The frequency of isolation of E. coli was also higher in the wet season than dry season (52.6% and 25.2% respectively).

Table 3 shows the susceptibility pattern of the 22 strains of *Shigella spp* examined. All the shigella isolates were

resistant to tetracycline and erythromycin but were susceptible to ofloxacin. Two peaks of MICs were found for ampicillin 3.1 and 100 mg/ml or greater. Almost all the MICs of cefdinir were distributed in a narrow range, from 0.1 to 0.4 mg/ml.

Table 1: Incidence and age distribution of enteropathogens isolated

	Yr	Yr	Yr	
Pathogen	< 1yr	1-2%	3-5%	(Total n = 135)
E.coli	16(11.9%)	12(8.9)%	77(57%)	
Shigella	2(1.5) %	12(8.9)%	8(59%)	
Other coliforms	2(1.5) %	3(2.2) %	2(1.5) %	
Vibro cholerae	-{-}	(-)	1(0.7%)	
	20	27	88	135

Table 2: Incidence and Seasonal Variation of Enteropathogens Isolated
From Patients presenting at Hospital in Osogbo

1.	Nos of Isolates.					
Organism	Dry	Rainy	Total%			
Shigella	6(4.4)%	16(11.9)%	22(16.3)%			
E.coli	34(25.2)%	71(52.6)%	105(77.8)%			
Other coliforms	2(1.5)%	5(3.7)%	7(5.2)%			
Vibro cholerae	0(-)	1(0.7)%	1(0.7)%			

Table 3: Drug susceptibility pattern of shigella spp isolated from patients

Presenting at hospital in Osogbo.

Minimum Inhibitory Concentration (MIC) mg/ml					
	ABPC	AT	EM	CFDN	OFLX
< 0.0.25	0	0	0	0 .	O.
0.05	0	0	Q	0	8
0.1	0	0	0	1	11
0.4	0	0	0	12	2
0.8	0	0	0	9	ī
1.6	1	1	0	0	0
3.1	5	0	0	0	0
6.3	3	0	2	0	0
12.5	1	0	11	0	0
,· 25	0	0	3	Ö	0
50	0	6	6	Ö	0
> 100	12	13	10 ,	0	0

ABPC: Ampicilin, AT: Tetracyclin CFDN: Cefdinir

EM: Erythromycin OFLX: Ofloxacin

DISCUSSION

Diarrhoeal diseases are part of the main social problems in Osogbo, as in other developing countries in tropics (1.11).Clarification of the enteropathogens involved in diarrhoeal diseases in the country is an essential step toward the implementation of effective primary health activities care against the diseases. This study information provides on prevalence of enteropathogens in a distinctive area, Osogbo, Osun State. Although it may not give a complete picture of the genuine spectrum of diarrhoeal diseases in the community, the survey is still significant as it is, based on information available to us, the intensive survey first etiological agents of diarrhoeal disease in Osogbo.

Esherichia coli and Shigella spp were identified in this study as the major causes of diarrhoeal diseases in Osogbo. They are bacteria that have been consistently identified as aetiological agent of diarrhoea. Statistical analysis showed that E. coli was significantly associated with diarrhoea in patients aged 3-5 years (P<0.05). It was found to be more prevalent than the other enteropathogens, which is consistent with the result of other researchers (11, 12). While the isolation of Shigella spp is also

similar to results obtained by other workers, the frequency of isolation of Shigella spp (16.3%) in this area was higher than those reported in other tropical countries (1, 9). This could be indicative of the important role Shigella spp plays in the outbreak of diarrhoea in children aged less than 5 years. The seasonal variation in incidence of the pathogen is similar to results obtained elsewhere (13, 14).

REFERENCES

- Guerrant RL, Hughes JM, Lima NL, Crane J. Diarrhoea in developed and developing countries; magnitude, spread settings and etiologies. Rev. Infect. Dis. 1990; 12 (Suppl.r): 541-550.
- Echeverria PD, Taylor N, Lexsonboon U, et al. Case control study of endemic diarrheal disease in Thai children. J. Infect. Dis. 1989; 159: 548.
- Ogunsanya TI, Rotimi VO, Adenuga A. A study of the aetiological agent of childhood diarrhoea in Lagos, Nigeria. 1994
- Albert MJ, Faruque SM, Faruque ASG, et al.
 Controlled study of Escherichia coli diarrheal

- infections in Bangladeshi children. J. Clin. Microbiol. 1995; **33**: 973-977.
- 5. Echeverria P, Orskov F,
 Orskov I, et al. Attaching
 and effacing
 enteropathogenic
 Escherichia coli as a cause of
 infantile diarrhea in
 Bangkok, J. Infect. Dis. 1991
 ; 164: 550-554.
- 6. Antai SP, Anozie SO. Infantile diarrhoea due to enteropathogenic Escherichia coli in Port Harcourt metropolis. J. Appl. Bacteriol. 1987; 62: 227 229.
- 7. Suyder SD, Merson MH. The magnitude of the global problem of acute diarrhoea disease. A review of active surveillance data. Bull. WHO. 1982; 66: 605-613.
- 8. Bhatnagar S, Bhan MK, Summer felt H, et al. Enteroaggregating

 Escherichia coli may be a new pathogen causing acute and persistence diarrhoea.

 Scand. J. Infect. Dis. 1993;
 25: 579-583.
- Yamashiro T, Noboru N,
 Naomi Higa MA. Etiological
 Study of Diarrheal Patients

- in Vientiane, Lao People's Democratic Republic. J. Clin. Microbiol. 1998; 2195-2199.
- 10. Agbodaze D, Abrahams CA, Arai S. Enteropathopenic and enterotoxigenic Escherichia coli as etiological factors of infantile diarrhoea in urban and rural Ghana. Trans. R. Soc. Trop. Med. Hyg. 1988; 82: 489-491.
- 11. Levine MM; Losonsky G, Herrington D, et al. Pediatric diarrhoea: the challenge of prevention. Pediatr. Infect. Dis. 1986; 5 (suppl): 29-43.
- 12. Adkins HJ, Escamilla J, Santiago LT, et al. Two-year survey of etiologic agents of diarrheal disease at San Lazaro Hospital, Manila, Republic of the Philippines. J. Clin. Microbiol. 1987; 25: 1143-1147;
- 13. Germani Y, Morillon M, Begaud E, et al. Two-year study of endemic enteric pathogens associated with acute diarrhea in New Caledonia. J. Clin. Microbiol. 1994; 32: 1532-1536.

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PHAGE AMPLIFICATION TECHNOLOGY AND ANTI-TUBERCULOUS DRUG SUSCEPTIBILITY TESTING IN NIGERIA

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The emergence of multi-drug resistant tuberculosis (MDR-TB) defined as combined resistance to the two most effective enti-tuberculosis drugs, rifampicin and isoniazid, threatens to create a public health hazard of unprecedented proportion. The fact that MDR-TR is more difficult and expensive to cure creates the need for prompt diagnosis. Conventionally, the proportion method on Lowenstein Jensen (LJ) medium is used in most developing countries as the 'gold standard' in the drug susceptibility testing of Mycobacterium tuberculosis (MTB) and it takes 3-4 weeks to give results from an MTB culture.

The use of phage as a diagnostic is fast gaining ground today. It involves targeting viable MTB cells from culture with a specific mycobacteriophage. After a one hour incubation, it is treated with an antivirus to destroy the phage that are not protected with the bacilli. Upon addition of cells of growing, non-pathogenic Mycobacterium smegmatis (sensor cells), progeny phage from the MTB cells infect the sensor cells, thus amplifying the effect of the phage. When plated in an agar medium overnight, plaques occur in the cell lawn indicating the presence of viable MTB in an original sample. A comparison is made between the number of plaques produced in a drug-free control and a sample incubated in the presence of the drug. While the presence of plaques beyond a cut-of point indicates drug resistance, the absence of plaques indicates that the drug destroyed MTB cells. Overall accuracy from several trials so far conducted is put at 97-98% compared with the 'gold standard'. With the phage amplification method, antituberculosis drug susceptibility results are obtained from MTB culture within 48 hours as opposed to the L J proportion method, which gives resulted in 3 to 4 weeks. Also, phage, as a diagnostic, is much more applicable in Nigeria laboratories than newer, rapid methods which requires specially dedicated instrumentation and are therefore very expensive. Phage amplification technology requires no special equipment and the results can be read visually.

Key words: Tuberculosis, drug susceptibility, phage, treatment, FASTPlaque-TB, rifampicin

INTRODUCTION

The World Health Organization (WHO) 1993 in declared tuberculosis (TB) a global emergency (1). Coupled with it deadly alliance with HIV/AIDS (2), TB has emerged as a public health of hazard unprecedented proportion especially for developing world which 95% of its burden (3). Eight million new cases and 2-3 million deaths from TB are recorded annually (4). Nigeria is one of the 22 countries, which bears 80% of

all incident cases of TB worldwide. She is also one of the 16 countries in which the progress of TB control is of greatest concern (5). Average annual case notification is put at 25,000 in the year 2000 26,641 however. cases recorded (7). In an epidemiological study of TB in Lagos, Nigeria, a significant · increase from incident rate of 21% in 1982 to 42% in 1992 was reported (8). Also, a study by Wokoma in 1999 showed a 7.74% increase in the incidence

rate of TB in Port Harcourt, between 1993 and 1999 (9). These were hospital-based studies. It is important to note that disease surveillance in Nigeria is neither intensive nor extensive and so cannot provide exact numbers of new cases or deaths occurring from TB in a year. Due to the stigma attached to TB in the Nigerian society, several cases go unreported. Of the cases reported, up to half or more are not diagnosed and therefor not treated. Smear microscopy has a sensitivity of 30-70% depending on whether direct or processed (concentrated) smears are examined (10).

TB is a curable disease, which ravaged the globe for over two centuries after its emergence. The discovery of streptomycin in 1946 offered a respite to TB sufferers because prior to its discovery, TB patients subjected to mutilating surgical procedures such as thoracoplasty in which large sections of the chest wall were excised in order to close cavities. TB came to be known as "captain of all men of death" as it continued to kill several famous people because there was no cure (12, 13). Several other anti-TB drugs were later discovered and they included rifampicin, pyrazinamide, ethambutol, ethionamide, thioacetazone, kanamycin,

viomycin, capreomycin and cycloserine. Of these, rifampicin and pyrazinamide are bactericidal in vivo while the others are bacteriostatic (13). For the treatment of TB, the WHO has listed five essential drugs, which include isoniazid (H), rifampicin (R), pyrazinamide (Z), streptomycin (S) and ethambutol (E). These are the most effective drugs against TB and are referred to as first line drugs. second group consists capreomycin, cycloserine, ethionamide, kanamycin, and a host of others. They are referred to as 'second line' drugs and are only used in cases of relapse, treatment failure or chronic cases (14,15). Usually a treatment regimen is made up of two phases namely, the initial phase during which the firstline drugs are used in combination to ensure rapid smear conversion from positive to negative, and a continuation phase (15). In Nigeria, the full regimen is rifampicin (R)/isoniazid (H) (combined tablet), pyrazinamide (Z) and ethambutol (E) daily for two months followed by thiazina (isoniazid plus thiacetazone) or ethambutol daily for 6 months. This is standardized according to the patient's age and body weight as well as the type of TB (16).

DRUG RESISTANCE IN TUBERCULOSIS

The greatest threat to chemotherapy today is the

frequency and rapidity with which bacteria develop resistance drugs (17). Several workers have demonstrated drug resistance. In a study by Fattorini and coworkers, the activity of sixteen antimicrobial agents was tested against drug resistant strains of Mycobacterium tuberculosis (MTB). Of the first line drugs, isoniazid was ineffective against all strains while resistance streptomycin; rifampicin, pyrazinamide ethambutol and were 80.4%, 71.7%, 39.1% and respectively. Amongst 8.7% second line anti-TB drugs, ciprofloxacin. resistance to ofloxacin, sparfloxacin, amikacin and kanamycin was 20%. About 10% of strains were resistant to capreomycin and cycloserine and 4.3% resistant were to ethionamide (18). In another study by Karamat and co-workers, four first line anti-TB drugs were tested 300 isolates form clinical samples at the Armed Forces Institution of Pathology, Rawalpindi. Of these, 52.66% were resistant to one drug at least. Of the resistant isolates, 26.33% were resistant to isoniazid, 24.0% to rifampicin, 28.0% to streptomycin and 23:33% to ethambutol with or without resistance to other drugs.

Besides MTB resistant to single drugs, there have been dramatic outbreaks of multidrug resistant tuberculosis (MDR-TB). It is described as a man-made disease because it is caused by treatment, inadequate improper drug supplies as well as erratic and indiscriminate use of drugs. The availability of drugs over the shelf most developing countries including Nigeria has led uncontrolled self-medication (20). Multi-drug resistance in TB has described combined been as rifampicin resistance to and isoniazid. Patients with MDR-TB face chronic disabilioty and death and they represent an infectious hazard for society (21). In the study by karamat and co-workers (19), multi-drug resistance was found in 41 isolates (13.66%). Studies have shown that rifampicin resistance is a good predictor of MDR-TB. This conclusion was drawn after studies in several parts of the world pointed the fact that MTB strains resistant to rifampicin are often invariably resistant to isoniazid at least and are thus considered MDR-TB (22). Some of such studies include those conducted in Estonia, Ethiopia, India (Delhi) and Latvia in which of the rifampicin resistant strains, 100%, 100%, 95% and 96% respectively were MDR-TB (22,23).

ANTI-TUBERCULOSIS DRUG SUSCEPTIBILITY TESTING

The management of MDR-TB requires a quick diagnosis as well as rapid and accurate susceptibility results to ensure early administration of a new regimen,

usually based on a quinolone, for retreatment of the patient. This is a major problem in developing countries including Nigeria.

In Nigeria, TB culture is limited to the National Reference Laboratory, which so far does not have a good liaison with peripheral laboratories where TB diagnosis by direct smear microscopy (16) is under the National done Control Programme. Even when such samples are sent to the reference laboratory, it takes 3-4 weeks after the availability of culture (which itself takes 6-8 weeks) for drug susceptibility results to be available. This is because under currently available infrastructure, it is only feasible to carry out drug susceptibility testing using the conventional proportion method on Lowenstein Jensen (LJ) medium.

The proportion method. which can also be done on the agar based Middlebrook 7H10 and 7H11 media is regarded as the 'gold standard' in susceptibility testing of TB (24) and is recommended by the WHO the International Union Against Tuberculosis and Lung Disease (IUATLD) (22). Though susceptibility testing can performed directly on sputum samples through this method to give results within 3-4 weeks. there is often the problem of inoculum standardization. Besides

the proportion method, there exist other methods, which include the resistance ratio and absolute concentration method, which are not commonly used.

The for need prompt treatment of MDR-TB has led to a search for more rapid method of susceptibility testing. There exists semi-automated methods like the Bactec 460 system (Becton. Dickinson) which gives results within 7-21 days, the automated MB/Bac T (Organon Teknika) and the Mycobacterial Growth Indicator Tube (MGIT) (Becton Dickinson) which also has manual use. While the Bactec 460 system detects growth by the evolution of radiolabeled 14CO2, the other two use non-radioactive materials. The results obtained by testing sputum samples directly can Ъe quite confusing and misleading because of insufficient growth of the control as well as contamination (22). Special skills and experience are required to read the results correctly. It will also be tantamount to a gross waste of reagents to perform susceptibility tests on organisms, which may turn out to be something other than MTB. It is easier and economical to perform susceptibility tests after culture results available. In most developing countries including Nigeria, the cost of these machines, servicing, cost of reagents and

personnel expertise prohibit their use.

Global The WHO Tuberculosis Programme recognizes the need for new, rapid and affordable diagnostic developing countries to replace the slower conventional methods. The committed GTB is to the promotion of new diagnostic of in TB proven value control programmes (26).

PHAGE AMPLIFICATION TECHNOLOGY

The of use mycobacteriophage in the disagnosis of TB is fast gaining ground today. Research into the use of phage in diagnosis dates back to 1947, when Gardner and Weiser identified phages that are specific for the mycobacteria (27). In 1960, Redmond and Cater isolated phages that specifically infect M. tuberculosis and M. bovis (28). The fact that phages are specific in their infection of hosts is the backbone of this technology. In the procedure, a specific phage that infects members of the MTB complex is used to target viable MTB in a specimen. If it is a positive sample, the phages infect the MTB cells (27). Any phage not incorporated within MTB cells are destroyed on treatment with a specific virucide. To confirm the presence of phage in the MTB cells, rapid growing Mycobacterium smeqmatis, а non-pathogenic mycobacterium, which is susceptible to infection by the phage is added. Using the pour plate method. this mixture is inoculated into molten agar, which is allowed to solidify, and the plate is incubated overnight. The phage particles within the MTB cells replicate and lyse the cells to release progeny phage. These then infect the cells of M. smeamatis. thereby amplifying the presence of the phage. Since M. smeamatis is a rapid grower, growth is seen in the medium overnight and plaques are seen where lysis of the cells occurred. A comparison of the number of plaques observed is made between culture suspensions incubated in both the presence and . absence of the antibiotic. number of plaques visualized from a given sample is related to the number of viable tubercle bacilli in the original sample (22,29,30-33).

FASTPLAQUETB-MDRITM

The FASTPlaque TB-MDRI™ (FPTB-MDRi) is a commercially available kit (Biotec Laboratories, Ipswich, UK) for the rapid detection of MTB resistance to rifampicin, a proven indicator of MDR-TB $\{22,23\}.$ Results of the trials conducted so far have been very impressive with 100% sensitivity and overall accuracy of 97%-98% compared with the LJ proportion method, which is used in most developing countries including Nigeria.

PHAGE AMPLIFICATION TECHNOLOGY VERSUS CONVENTIONAL METHODS

In comparison with the LJ proportion method. amplification technology is rapid; giving conclusive results within 48 hours after culture is available. With the FASTPlaque TB-MDRi, further studies have indicated that it may be possible to achieve a 24hour test by the use of a shorter rifampicin incubation period (31). Availability of susceptibility results in a shorter time will reduce morbidity and mortality from MDR-TB and treatment can be promptly initiated. It will thus check the spread of MDR-TB in community. , the amplification technology does not complicated procedures involve requiring extra-ordinary expertise. It involves simple microbiological skills like centrifugation, pipetting and pour plate culture method. The results are read visually and it requires no specially dedicated instrumentation hence it is less expensive than the new automated methods and it is applicable under currently available infrastructure in most laboratories in developing countries. One of the attractions of the FASTPlaque TB-MDRiTM Kit (Biotec Laboratories, Ipswich, UK) is that it contains all materialsreagents, freeze-dried virus, M. smegmatis, virucide, and the drug, medium and reagent bottles

needed for the test. It is thus very convenient. The FASTPlaque TB-MDRiTM assay has been evaluated and compared with conventional methods in several studies. In a study conducted at the Reference Laboratories of the South African Institute of Medical Research (SAIMR) in Johannesbourg (LAB I) and Cape Town (LAB II), the ability of the FPTB-MDRi kit to correctly identify rifampicin susceptibility on strains Mycobacterium tuberculosis culture on solid media was and evaluated compared with conventional methods. By conventional methods, 81 out of 191 strains were found to rifampicin resistant and 110 strains out of 191 were rifampicin susceptible. Respectively, sensitivity, specificity and overall accuracy of the FASTPlaque TB-MDRi for LAB I were, 100%, 97% and 98% and for LAB II they were 100%, 94% and 97% (22). In another study at the TB Unit of SAIMR, Johannesbourg, the ability of FPTB-MDRi to detect rifampicin resistance within 48 hours was evaluated using samples with Bactec 460 known automated culture results. In this study, the use of FPTB-MDRi was compared to the Bactec 460 radiometric method drug in determining TB susceptibility patterns to rifampicin. Also, the possibility of reducing the time for reporting

results from 48 hours to 24 hours was explored. The study showed that FPTB results are available within 48 hours (sooner than the automated Bactec 460 method). With a reduced rifampicin incubation time, the overall time for reporting results could be reduced to 24 hours. Overall accuracy was 90% (31).

COST-EFFECTIVENESS OF PHAGE AMPLIFICATION TECHNOLOGY

TB is a disease symptomatic of poverty. MDR-TB is particularly common among those who have previous had anti-TB chemotherapy, ethnic minority groups, migrants, refugees, substance abusers, HIV positive patients and the homeless (34). is therefore Cost a major consideration that determines the applicability of diagnostics in the developing world including Nigeria. This is exemplified by the fact that despite the availability of rapid sophisticated methods for the diagnosis of TB and the detection of MDR-TB, none of these are being used in National TB control programmes in most third world countries. Any new test- must therefore compare favourably with the conventional methods otherwise their high cost would prohibit their use.

The cost of full diagnosis of TB (AFBX3, x-ray, culture and sensitivity) is about N2000

(approximately \$15). This is highly variable depending on where mile tests are done and the method used. The FASTPlandeTB MDRi kil costs \$400 per determination of 50 if one buys using the small quantities from Biotec Laboratories, Ipswich, UK (35). It is difficult to work out what the cost wper test would eventually be in Wigeria Will the availability of discount of build purchases but it is unfikerento go beyond WHO recommendations of for avzedflad \$10 than susceptibility test. It balaces very little demand on theib scarce resources of Nigerian laberatories because it is manual, requirities no MARKEY specially instrumentation. MMGL 9 .

It has been suggested that it may be possible to reduce the cost of this technology by propagating the phage and M. smegmatis in a local laboratory. Studies however shown that a particular gene in M. smegmatis when over expressed induces resistance to certain phages (36). Coupled with the problem of standardization, it the may be better to use commercially available kit Whichi passes through a quality contible test to ensure high pefformälfde' standards before it is planged पारिक suscernfility result the market.

The ability of of phage amplification of the chrology and the promptly detect ambred will reduce morbidity and mortality

the disease. from Also. the number of contacts that patient has a chance to infect will be greatly reduced. Thus the number of people requiring anti-TB drug susceptibility testing as well as treatment with the more expensive second line drugs in the future will be reduced. These future savings could be used to pay for the diagnostic (26). With increased budgetary allocation to the National Tuberculosis and Control **Programme** Leprosy (NTLCP) and donor support, the cost of diagnosis and treatment on the individual patient would be greatly reduced.

CONCLUSIONS AND RECOMMENDATION

The emergence of MDR-TB is indeed a major threat to the treatment of TB and an important impediment to National Control Programmes. In order to avoid its fatal effects on individual patients and its spread within communities, there is need for diagnosis. FPTB-MDRi gives susceptibility results from MTB cultures within 48 hours and is therefore recommended as a substitute for the conventional LJ proportion method, which takes about one month to provide susceptibility results. This would ensure that appropriate treatment is initiated in good time.

Cost has emerged as a major consideration and an

important factor determining the applicability of new diagnostics in the National TB Control Programmes of developing counties including Nigeria. There is urgent need for all tiers of government to prioritize TB control its because of chronic debilitating effects on the workforce and the general population as well as its deadly alliance with HIV/AIDS, which has its highest prevalence in developing countries. Prioritization of TB control goes expression of the beyond an political will to do so. It involves actual dedication and disbursement of funds to the National TB Control Programme. This has necessary because majority of TB patients are poor and so cannot afford exorbitant testing and treatment. Ensuring that such patients undergo and receive free or highly subsidized testing treatment would be an indirect investment into the workforce of nations. The right to quality/affordable healthcare entrenched in several international human rights documents as well as the Nigerian constitution.

There is the need for the NTLCP in Nigeria to embark on laboratiories inspection and to identify and equip strategically located centers where TB testing can be done nationally. This would do little to reduce the spread of MDR-TB as the onus of culture and

drug susceptibility testing lies with the National TB Reference Laboratory in Lagos which so far has no strong liaison peripheral laboratories. Ιt is therefore recommended that a network of laboratories be established so that samples from cases of treatment failure can be promptly submitted to. the Reference Laboratory for further diagnosis. Amongst other things, vehicles provision of and biohazard containers/ice chests for the transportation of samples from peripheral centers to the National Reference center is a top priority.

The World Health Organisation as the regulatory body for National TB Control Programmes has a great role to play in promoting the evaluation of new diagnostics as well as recommending their use. It is thus recommended that the WHO assists National Reference Laboratories in developing countries where TB prevalence is highest to evaluate new kits in other to ascertain their adaptability particular to situations. The WHO also has a play role in supporting laboratory personnel in charge of TB diagnosis to attend training courses to improve their skills especially in new technologies.

Though considered rapid compared with the LJ proportion

method, FPTB-MDRi relies on culture. There is the therefore need for the manufacturers to engage in continuing research in order to produce kits that could detect MDR-TB rapidly and directly form sputum and other samples.

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REFERENCES

- World Health Organization (WHO). Tuberculosis— a global emergency.WHO report on the TP epidemic. Geneva-27, 1211, Switzerland. 1994.
- 2. Toosi Z, Nicolacakis K, Xia L, et al. Activation of latent HIV-I by Mycobacterium tuberculosis and its purified protein derivative in alveolar macrophages from HIV infected individuals in vitro. J. Acquir. Immune Defic. Syndr.

- Hum. Retrovirol. 1997 15(5): 325-31.
- 3. Snider DE Jr. Tuberculosis: the world situation. In: Porter JDH, McAdam KPWJ (eds) Tuberculosis: back to the future. John Wiley and sons, Chichester, UK. 1994: 13-31.
- 4. Dye C, Scheel S, Dolin P, et al. Global Burden of Tuberculosis. Estimated Incidence, Prevalence and Mortality by Country. JAMA. 1999; 282 (7): 677-685.
- 5. Netto EM, Lye C, Raviglione MC. Progress in global tuberculosis control 1995-1999, with emphasis on 22-high burden countries. Global Monitoring and Surveillance Project, Int. J. Tuberc.Lung Dis. 1999; 3(4): 310 320.
- 6. National Tuberculosis and Leprosy Control Programme (NTLCP). Works manual. Federal Ministry of Health, Lagos. 1991: 96.
- 7. National Tuberculosis and Leprosy Control Programme (NTLCP). TB situation, 2000. Federal Ministry of Health, Abuja, Nigeria. Unpublished report 2001.

- 8. Idigbe EO, Sofola TO, John EKO, et al. The trend of pulmonary tuberculosis in Lagos, Nigeria.

 Biomedical Letters. 1995; 31: 99-109.
- 9. Wokoma FS. Trends in case occurrence of pulmonary tuberculosis in Port Harcourt Teaching Hospital-A five year analysis of admission. Nig. Med. Practitioner. 1999; 37 (3/4): 41-46.
- Strumpf IJ, Tsang AY, Sayre JW. Re-evaluation of sputum staining for the diagnosis of pulmonary tuberculosis.
 Ann. Rev. Respir. Dis. 1979;
 119: 599 602.
- 11. Behr MA. Transmission of Mycobacterium tuberculosis from patients, smear negative for ácid fast bacilli. Lancet. 1999; **353**: 444-449.
- 12. Chadwick MV. Mycobacteria. Wright P.S.G., London. 1982:
- 13. Collins CH, Grange JM, Yates MD. Organisation and Practice in Tuberculosis Bacteriology. Butterworts, London. 1985: 1-110.
- 14. Villiorina MF, Geiter LJ, Simone PM. The Multidrug Resistant Tuberculosis

- Challenge to Public Health Efforts to Control Tuberculosis. <u>Pub. Health</u> <u>Rep.</u> 1991; **107**: 616-625.
- 15. World Health Organization.
 Treatment of Tuberculosis:
 Guidelines for National
 Programmes. WHO, Geneva27, 1211; Switzerland. 1993
 : 1-29.
- 16. National Tuberculosis and Leprosy Control Programme (NTLCP). Works Manual. Federal Ministry of Health. 1998: 41, 164.
- 17. Olanipekun KA, Montefiore
 O. Bacterial infections,
 patterns and chemotherapy
 among hospital
 patients in the tropics.
 Scand. J. Infect. Dis. 1978,
 10: 906-911.
- World Health Organisation. Anti-TB drug resistance surveillance. WHO, Geneva-271211, Switzerland. 1997.
- Heifets LB. Tuberculosis In: Lynch JP (ed.). Seminars in Respiratory and Critical care Medicine. Thieme Medical Publishers. 18:5 New York. 1997.
- Siddiqi SH, Libonati JP;
 Middlebrook G. Evaluation

- of a rapid radiometric method for drug susceptibility testing of *Mycobacterium tuberculosis*. *J. Clin. Microbiol*. 1981; 13: 908-912.
- 21. Foulds J, O'Brien R. New Tools for the diagnosis of tuberculosis: the perspective of developing countries. Int. Tuberc. Lung Dis. 1998; 2(10): 778-783.
- 22. Gardner GM, Weiser RS. A bacteriophage for Mycobacterium smegmatis.

 Proc. Soc. Exper. Biol. Med. 1947; 66: 205-206.
- 23. Redmond WB, Cater JC. A bacteriophage specific for Mycobacterium tuberculosis vars hominis and bovis. Am. Rev. Respir. Dis. 1960; 82:781-786:
- 24. Wilson SM, Al-Suwaidi Z, McNerney R, et al. Evaluation of a new rapid bacteriophage-based method for drug susceptibility testing of Mycobacterium tuberculosis.

 Nature Medicine. 1997;
 3:465-468.
- 25. Mole RJ, Maskell T. WO'C phage as a diagnostic-the use of Phage in TB Diagnosis. J. Ghem. Technol. 2001; 76:: 683-688.

- 26. Albert H, Heydenrich A.
 Rapid rifampicin
 susceptibility testing of
 Mycobacterium tuberculosis
 using FAST plaque TB-RIF.
 ASM General Meeting, Los
 Angeles, U.S.A. 2000
- Eltringham IJ, Drobniewski 27. Mangan JA. et Evaluation of reverse transcription-PCR and a bacteriophage-based assay for the rapid phenotypic rifampicin detection of resistance in clinical isolates Mycobacterium ofClin. tuberculosis. J. Microbiol. 1999; 37 3524-3527.
- 28. Eltringham, IJ, Wilson SM, Drobniewsk, FA. Evaluation of a bacteriophage-based (phage amplified assav biological assay) as a rapid screen for resistance to isoniazid, ethambutol, streptomycin, pryazinamide and ciprofloxacin among clinical isolates of Mycobacterium tuberculosis. J.Clin. Microbiol. 1999; 37: 3528-3532.

- Zhang Y. Molecular Basis of 29. Drug Resistance in Mycobacterium tuberculosis. In: Hart CA, Becching NJ, ΒĬ (eds). Duerden Tuberculosis into the next century. J. Clin. Microbiol. 1996; **24**: 1-34.
- 30. Personal Communication with Rowland king, Biotec Laboratories, Ipswich, UK
- 31. Barsom EK, Haftull GF. Characterization of a Mycobacterium smegmatis gene that confers resistance to phages L5 and D29 when over-expressed. Mol. Microbiol. 1996; 21: 159-170.

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METHICILLIN RESISTANCE IN STAPHYLOCOCCAL ISOLATES FROM CLINICAL AND ASYMPTOMATIC BACTERIURIA SPECIMENS: IMPLICATIONS FOR INFECTION CONTROL

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The study assessed the importance of Staphylococcus aureus as a urinary pathogen and the incidence of multidrug resistant (MDR), methicillin-resistant Staphylococcus aureus (MRSA). A total of 86 staphylococcal isolates made up of 50 clinical isolates from urine samples submitted to the Medical Microbiology Laboratory of Ahmadu Bello University Teaching Hospital and 36 asymptomatic bacteriuria isolates from urine samples of 'healthy' volunteers within the university community were tested for their susceptibility to various antibiotics and production of -lactamase enzyme. A total of 27 isolates (31.4%) were methicillin resistant, with 12(44.4%) being methicillin resistant coagulase-negative staphylococci (MRCNS). Majority of the isolates tested were resistant to the cheap, readily available brand-spectrum antibiotics; ampicilin, amoxicillin, chloramphenicol, tetracycline and penicillin G. All the isolates were resistant to three or more of the antimicrobial agents tested. A total of 14/50 (28%) of the clinical isolates and 17/36 (47,2%) of the 'community' isolates from healthy volunteers were resistant to 7 or more of the antimicrobial agents tested. Analysis of the multiple antibiotic resistance (MAR) index of isolates and the production of lactamase enzyme showed that 56 isolates representing 65.1% of the total number tested had an MAR index of 0.5 and above indicating that they probably originated from an environment where antibiotics are frequently used. The implication of these findings for instituting effective control measures aimed at reducing the pool of antibiotic-resistant organisms is discussed.

Key words: Methicillin-resistant, staphylococcus aureus, asymptomatic bacteriuria, infection control

INTRODUCTION

Antimicrobial resistance is a well recongnised problem worldwide (1, 2). The resistance organisms have however been associated primarily with hospitals; especially in intensive care units (1). The ubiquitous occurrence of staphylococci ensures that man is constantly exposed to this group microorganisms, thus infection of various parts of the body caused by staphylococci are very common (3,4).

Staphylococcus aureus continues to be a major cause of community acquired and healthcare related infections around the world (5,6). The emergences of high level of penicillin resistance followed by the development and spread of strains resistance to the semi-synthetic penicillins (methicillin, nafcillin oxacillin), macrolides. tetracyclines and aminoglycoside made the theraphy have staphylococcal disease a global challenge (6,7,8).

Methicillin-resistant

Staphylococcus aureus (MRSA) is a virulent organism that causes significant mortality and morbidility, especially to patients in critical care areas (9). MRSA can (and does in some cases) also contribute to an increased length of hospital stay and healthcare costs. Infections with methicillinresistant Staphulococcus aureus methicillin-resistant and coagulase-negative staphylococci (MRCNS), have been widely reported. Those infections were initially confined to hospitals and nursing homes especially intensive care units where combination οf debilitated patients, invasive technology, and high antimicrobial use facilitates infections by multi-drugs resistant staphylococci, enterobacteria resistant to third-generation cephalosporins and imipenem resistant non-fermentative bacteria (10). However, cases of community-acquired MRSA have been reported, primarily persons with history of injection drug use and other high-risk patients (11).

Recently, communityacquired MRSA been have described in both adults and who did children. not have extensive exposure to hospitals or apparent risk factors **Antimicrobial** (12,13,14,15,16). resistance often leads to

failure of empirical therapeutic | therapy; therefore, knowledge of the local prevalence of pathogens and antimicrobial sensitivity their patterns is essential for clinicians in their routine work (17). Effective antibiotic therapy in developing countries is severely limited by the large reservoir of antibiotic resistant bacteria that exist within their population. The healthy members of any community represent its largest reservoir of bacteria resistant to antimicrobial agents (18).

The increase in antimicrobial resistance is creating a lot of problems. These have focused attention measures for upon fighting resistance. foremost which is susceptibility surveillance (19). The rapidity of emergence of antibiotics multiple resistant organisms is not being reflected by the same rate of development of new antimicrobial agents. It is therefore conceivable that patients with serious infections will soon no longer be treatable with currently available antimicrobial agents (20).

Before instituting control measures that will be appreciated all healthcare professionals, there must be scientific data to ascertain the extent of the problem posed by multi-drug resistant organisms like the MRSA to the of antimicrobial outcome chemotherapy in the hospital and immediate community. Unfortunately, surveillance studies on the epidemiology of MRSA and their antimicrobial susceptibility lacking in this are patterns environment.

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assessing the importance of Staphylococcus aureus έt urinary pathogen determine the incidence of multi-drug resistant, methicillin-resistant Staphulococcus aureus (MRSA) in clinical isolates from urine in a University Teaching Hospital and compare these with isolates from 'healthy' individuals within the community. The

implications instituting for effective control measures that can reduce the pool of antibioticresistant organisms within healthy members of the community and in the hospital setting are discussed.

MATERIALS AND METHODS Bacteriology

Staphylococcal isolates from obtained urine sample submitted to the Department of Ahmadu **Med**ical Microbiology, Teaching Bello University Hospital, Zaria, and 'healthy' student volunteers from Ahmadu Bello University were analysed.

The isolates were characterized using established methods, which included colonial morphology, Gram characteristics, ability to produce the enzyme peroxidase, coagulase and the presence of heat stable DNAse activity to separate Staphylococcus aureus strains from coagulase-negative the staphylococci (CNS). A standard Staphylococcus aureus strain ATCC 13709 was obtained from the οſ National Institute and Research Pharmaceutical Development, Abuja, Nigeria.

Chemicals and media

The media used WCTC Nutrient Broth (NB), Nutrient (NA) and Mannitol Salt Agar (MSA) all from Oxoid. The chemicals include hydrogen peroxide (3%).deoxyribonucleic acid, sodium iodine chloride, starch and solution.

Antimicrobial sensitivity testing

The susceptibility pattern of the following theisolates to antibiotics was determined; Ampicillin 25 µg, Chloramphenicol 20 ug, Cloxacillin 10 μg, Erythromycin 10 µg, Gentamcin µg, Penicillin G 1.5 iu, Streptomycin 10 μg, Amoxycillin 25 μg, Ciprofloxacin 5 µg and Methicillin 5 µg, using the Bauer modified Kirby diffusion technique (21). The isolates were grown overnight in nutrient broth and the inocula spread on the surface of the previously prepared plates sterile nutrient agar flooding with 2mls of the standardized suspension. Excess were drained off and allowed to dry in a warm incubator for about 15-20 minutes. Using sterile forceps, multiantibiotic discs were placed on

the dried nutrient agar plate and left at room temperature for about 25minutes to allow the antibiotics to diffuse in the agar medium. Similar treatment was extended to standard the Staphylococcus aureus ATCC 13709. All the plates were incubated at 37° C for 24 hours in inverted position. Thereafter, the diameter of the zones of inhibition of the isolates and the standard Staphylococcus aureus were measured to the nearest millimeter.

Determination of Methicillin sensitivity

Nutrient agar medium containing 5% w/v sodium chloride (22) was prepared, distributed into 20ml aliquots and sterilized at 1210 C for 15 minutes. Overnight cultures of the isolates were used to flood the surfaces of the prepared agar media, drained and allowed to dry. Methicillin discs (containing 5µg of methicillin) were placed on the dried agar plate and treated as previously described above, but incubation was at 350 C. The diameter of the zones of inhibition was similarly determined.

Test for β-lactamase production

Suspensions of the isolates were prepared in triplicates by emulsifying bacterial colonies (from an overnight nutrient agar culture) with sterile loops in 0.5 ml of phosphate buffer solution containing 0.06 mg/ml (10,000)

units/ml) of Penicillin G. As control, cell suspension of the standard typed culture of Staphylococcus aureus (ATCC 13709) was similarly set-up. They were incubated at room temperature for at least 1 hour. Thereafter, 2 drops of freshly prepared 1% aqueous solution were added to each bacterial suspension and shaken. To this was added 1 drop of iodine solution and allowed to stand for 10 minutes at room temperature. βlactamase producing organisms changed the colour of the reaction mixture from blue-black colourless within the 10 minutes.

Determination of multiple antibiotic resistance (MAR) index

The MAR index was determined for each isolate by dividing the number of antibiotics to which the isolates is resistant by the total number of antibiotics tested (23,24).

MAR index = Number to which isolate is resistant

Total number of antibiotics tested

RESULTS

Of the staphylococcal isolates, 50 were clinical isolates from urine samples submitted to the Department of Medical Microbiology, Ahmadu Bello Teaching Hospital, while 36 were "community" isolates form urine samples of 'healthy' volunteers within the university community. A total of 27 isolates (31.4%) were

resistant. 12 methicillin with (44.4%) being MRSA, while 15 (55.6%) were MRCNS. Out of the 59 methicillin-sensitive staphylococcal isolates, 40 (67.8%) were MSSA while 19 (32.2%) were Figure 1 shows MSCNS. proportion of the staphylococcal isolates resistant to various antibiotics.

The MAR index of isolates by the proportion, that are β-lactamase positive and methicillin-resistant is shown on Table 1. A breakdown of the analysis of the MAR index of the 'community' and clinical isolates is shown in Table 2.

All the isolates were resistant to three or more antibiotics, while 17/36 (47.2%) 'community' isolates and 14/50 (28%) clinical isolates showed multi-drug resistance to seven or more of the antibiotics Figure 1 shows the tested. susceptibility Ωſ patterns staphylococcal different to antibiotics. Table 1 shows multiple antibiotic resistance (MAR) index of Staphylococcus aureus isolates. with the proportions that are βlactamase positive and methicillinresistant. Table 2 shows the analysis of MAR index of the clinical and asymptomatic bacteruria (community) isolates.

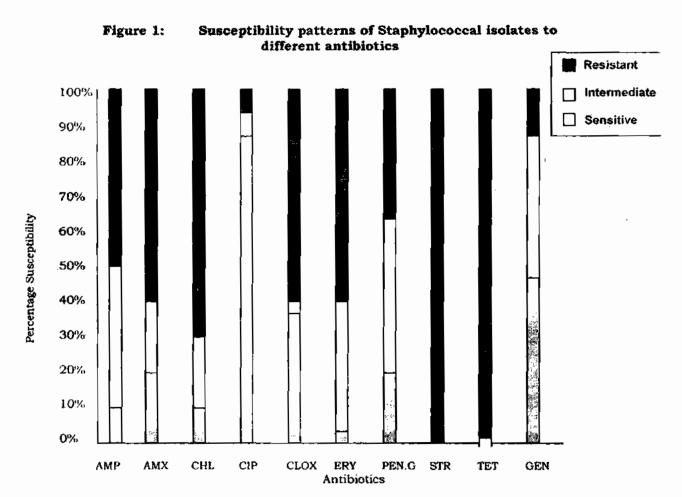


Table 1: Multiple antibiotic resistance (MAR) index of Staphylococcus aureus isolates, with the proportions that are β -lactamase positive and methicillin-resistant.

MAR index	No. of isolates (%)	-	MR (%)	
0.3	19(22.1)	3(15.8)	2(10.5)	
0.4	10(11.6)	0(0.0)	· [(10,0)	
0.5	12(14.0)	3(25.0)	1(8.3)	
0.6	12(14.0)	7(58.3)	3(25.0)	
0.7	11(12.7)	7(43.6)	6(54.5)	
0.8	15(17.4)	10(66.7)	8(53.3)	
0.9	6(7.0)	3(50.0)	6(100.0)	
1.0	1(1.2)	1(100.0)	0(0.0)	

Table 2: Analysis of MAR index of the clinical and asymptomatic bacteruria (community) isolates number of

MAR index	Community isolates	Clinical isolates		
0.3	.3	17		
0.4	3	7		
0.5	4	8		
0.6	8	4		
0.7	6	5		
0.8	4	7		
0.9	4	2		
1.0	1	0		
TOTAL	36	50		

DISCUSSION

Nosocomial infection caused by multi-resistant organisms in developing countries respresent a major public health problem that is not universally recognized (25). Results from various studies in the past did not identify Staphylococcus aureus as**imp**ortant urinary pathogen (3,26,27), rather there has been a

focus on the CNS identified as a mjor cause of infections associated with prosthetic implants and medical devices (28) and urinary tract infection, particularly in young sexually active women (29,30). The 72.4% prevalence of Staphylococcus aureus in this study points to the increasing important of this organism as a urinary

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pathogen and a common isolate in asymptomatic bacteriuia in this environment.

of the isolates Majority tested were resistant to the cheap, readily available broad- spectrum Ampicillin antibiotics; (89.3%), (83.3%),Amoxycillin Chlorampheniol (89.3%),Tetracycline (98.8%) and Penicillin (83.3%). This result consistent with the observation clinical that staphylococcal isolates are resistant to a large number of commonly prescribed antimicrobial agents (30).

The level of multi-drug resistance exhibited by staphylococcal isolates in this study is alarming. All the isolates were resistant to three or more of the antimicrobial agent tested. The MDR strains came from both clinical and 'community' isolates. A total of 14/50 (28%) of the clinical isolates and 17/36 (47.2%) of 'community' isolates healthy volunteers wee resistant to seven or more of the antimicrobial agents tested. The high percentage of isolates form 'healthy' individual showing high MDR goes to confirm the assertion that the healthy members of the community represent its largest reservoir of bacterial resistant to antimicrobial agents (18,31). MAR index higher that 0.2 has been said to be an indication of isolates originating from an environment where antibiotic were often used (23,24).

Since all the isolates were not hospital environment from the where antibiotic are often used but also from urine of asymptomatic volunteers in 'healthy' university community. this observation goes to confirm widespread abuse/misuse of antibiotics this community. in Administration of antibiotics often selection permits the and overgrowth of multiply resistant organi**sms** $\{32\}.$ The selective pressures favouring resistant strains are known to arise form misuse and overuse antimicrobials (notably extended spectrum cephalosporins) increased numbers of immunocompromised hosts, lapses in infection control (where they exist), increased use of invasive procedures and devices, and widespread use of antibiotic in agriculture and animal husbandary (33).

It has been documented that resistance properties are easily transferred between organisms of the same or different through the agency of plasmid. Evidence of transfer of high-level resistance to gentamicin, tobramycin and kanamycin between staphylococi of the same different species by filter mating also exists (34). Restriction endonuclease analysis of plasmids five isolates from of Staphylococcus epidermidis also supported the hypothesis that plasmid transfers tween the two species occur in nature (35). Transfer of such resistance determinants may have been responsible for the high level of MDR encountered in this study.

A breakdown of the number of isolates with a particular MAR index and proportion that are βlactamase-positive showed that 56 isolates representing 65.1% of the total number tested, had an MAR of 0.5 and above, while 29 isolates (33.7%) had MAR index less than 0.5. This is a relative indication of the susceptibility of the isolates to the test antibiotics. One isolate that was resistant to all the 10 antibiotics tested was isolated from the urine of a 'healthy' volunteer in community. The isolate produced β-lactamase. coagulasewas positive but sensitive to methicillin.

Multi-drug resistant Staphylococcus aureus have been known to produce β-lactamase in greater amounts than strains that are fully sensitive to antibiotic or resistant to only to penicillin (22). Contrary to the findings from other studies (4,36,37), bacterial resistance to ciprofloxacin (a fluoroquinolone) as high as 13.1% was encountered in this study. The increasing resistance to such a new and expensive, reserve drug

is probably an indication of the increasing level of availability, misuse and overuse.

The tremendous therapeutic advantage afforded by introduction new antibiotic is threatened by the emergence of increasingly resistance strains of microbes (33). Introduction of new antibiotic are essential, but their useful life will be enhanced only if used wisely and sparingly (38). The ever-present danger of individuals infections contracting in community makes it imperative that measures aimed at reducing the pool of antibiotic resistant organisms existing within healthy members of the community be instituted within delay.

ln concert with improved efforts prescribing habits. and isolate resistant identify organisms that can be introduced into healthcare setting from outside institutions are essential. Evidence from various studies have shown that, surveillance when used to guide polices on antibiotic use and infection control, can be helpful to control the development and spread of antimicrobial resistance within hospital setting and the community at large (19,39,40).

We agree that persistence would be required in influencing the behaviour of healthcare professionals and to maintain optimal infection control policies and procedures within the hospital

and the community at large. In the meantime, it is highly desirable to continuously monitor the antibiotic resistance situation so as to maximize the possibility of administering an effective antimicrobial agent whenever there is need to do so.

REFERENCES

- 1. Flaherty JP, Weinstein RA.
 Nosocomial infection caused
 by antibiotic resistant
 organisms in the intensivecare unit. Infect. Control.
 Hosp. Epidermiol. 1996; 17
 : 236- 48.
- 2. Finch RG. Antibiotic resistance. J. Antimicrob.

 Chemother. 1998; 42: 125-28
- Shanson DC. Staphyloccal infections in hospitals. Brit.
 J. Hosp. Med. 1986;35: 312-320
- Simon C, Stille W, Wilkinson J. Antibiotic Therapy in Clinical Practice.2nd edition. Schattauer Stuttgart-New York. 1993: 3-15, 37-8
- Kauffman CA, Bradley SF.
 Epidermiology of community acquired infection. In:
 Crossley KB, Archer GL (editors). The staphylococci

- in human disease. Churchill Livingstone. NewYork. 1997: 287-308.
- 6. Lowy FD. Staphylococcus aureus infections. New. Engl. J. Med 1998; **339**: 520-32.
- 7. Struclens MJ, Mertens R
 National survey of
 methicillin-resistant
 Staphylococcus aureus. Eur.
 J. Clin. Microbiol. Infect. Dis.
 1994; 13: 56-63.
- 8. Maranan MC, Moreira B, Boyle-Vavra S, Daum RS: Antimicrobial resistance in staphylococci: epidermiology, molecular mechanisms, and clincal relevance. Infect. Dis. Clin. North. Am. 1997; 11: 813-49.
- 9: Baird VL, Hawley R. Methicillin-resistant

 Staphylococcus aureus (MRSA): is a need to change clinical practice? Int. Crit. Care. Nurs. 2000; 16(6): 357-366.
- 10. Archibald L, Philips L,
 Monnet D, et al.
 Antimicrobial resistance in
 isolates from inpatients and
 out-patients in the United
 States: increasing importance
 of the intensive care unit.

Clin. Infect. Dis. 1997; **24**: 2.1-5.

44

- Saravolatz LD, Pohold DJ, 11. Arking LM. Commnuityacquired methicillin resistant Staphylococcus aureus infections: a new for noscomial source Ann. outbreaks. Intern. Med.1982; 97:325-9
- 12. Moreno F, Crisp C, Jorgensen JH, Patterson JE. Methicillin-resistant Staphylococcus aureus as a community organism. Clin. Infect. Dis. 1995; 21(5): 1308-1312.
- 13. Herold BC, Immergluck LC, Marauen MC, et al. Community-acquired methicillin resistant Staphylococcus aureus in children with no identified predisposing risk negative staphylococcus risk JAMA. 1998; 279(8): 593-8.
- 14. Gorak EL. Yamada SM. JD. Community Brown methicillinacquired Staphylococcus resistant hospitalized aureus in adults and children without known risk factors. Clin. Infect. Dis .1999; 29: 797 -800.

- 15. Gosbell IB, Mercer JL, Neville SA, et al. Non-multiresistant and multi-resistant methicillin-resistant

 Staphylococcus aureus in community acquired infections. Med. J. Aust. 2001; 174: 627-30.
- 16. Jones TF, Kellum ME, Porter SS, et al. An outbreak of community acquired foodborne illness causedby methicillin-resistant

 Staphylococcus aureus.

 Emerg. Infect. Dis. 2002; 8(1): 82-84.
- Andrasevic AT, Tambie T. 17. Kalenic S, Jankovic V and tne Working Group of the Committee Croatian for Antibiotic Resistance Surveillance for Antimicrobial Resistance Croatian in Infectious Diseases. Emera. Infect. Dis. 2002; 8(1): 14 -18)
- 18. Lester SC, Pla MP, Wang F, et al. The carriage of Escherichia coli resistant to antimicrobial agents by healthy children in Boston, Caracas, Venezuela and Qui Pu, China. N. Engl. J. Med. 1990; 323: 285 89.
- 19. Masterton RG. Surveillance studies: how can they help the management of infection?

- J. Antimicrob. Chemother.
 2002; 46 Suppl T2: 53-8.
- 20. Elliot TS, Lambert PA.
 Antimicrobial resistance in
 the intensive-care unit:
 mechanisms and
 management. Br. Med. Bull.
 1999; 55 (10): 59-76.
- 21. Cheesbrough M. Medical Manual for Laboratory Tropical Countries. Vol..II Microbiology. Butterworth Heinemann Ltd. Linacre House, Jordan Hill Oxford OX2 8DP; 1993.
- 22. Cookson B, Phillip I.

 Methicillin-resistant
 staphylococci. In:
 Staphylococci. Jones D,
 Board RG, Sussnan M (eds.).
 J Applied Microbiology 1990;
 69 Symposium series No 19:
 55-70.
- 23. Krumperman PH. Multiple antibiotic indexing Escherichia coli to identifying risk sources of faecal contamination of foods. Appl. Environ. Microbiol. 1983; 46: 169-71.
- 24. Paul S, Bezbarauh RL, Roy MK, Ghosh AC Multiple antibiotic resistance (MAR) Index and its reversion in Pseudomonas aeruginosa.

- Appl. Microbiol. (Lett.) 1997; **24**: 169-71.
- 25. Ponc-de-Leon, S. The needs of developing countries and the resources required. J. Hosp. Infect. 1991; 18 Suppl A: 376-81.
- 26. Kosimidis. J, Gargalianos. P, Adamis G, et al. Fleroxacin in single dose oral therapy of uncomplicated urinary tract infection. J. Antimicrob. Chemother. 1988; 22 suppl D: 219-21
- 27. Kosakai N, Kumamoto Y, Hirose T, et al. Comparative studies on activities of antimicrobial agents against causative organisms isolated form urinary tract infections 1: susceptibility, distribution. *Jpn. J.Antibiol.* 1990; 43 (6): 919-53.
- 28. Hogt AH, Dankert J, de Vries JA, Feijen J. Adhesion of coagulase negative staphylococci to biomaterials. J. Gen. Microbiol. 1983; 129: 2959-2968.
- 29. Gerald LM, Dougglas RG, Benette JE. Principles of infectious diseases. 3rd edition. Churchill Livingstone. New York. 1990: 582-590.

- 30. Ol koya DK, Asielue JO, Olasupo NA. Ikea JK. Plasmid profiles and antibiotic resistance patterns of Staphylococcus aureus isolates from Nigeria. Afr. Med. Sci. 1995; 24; 135-39.
- 31. Lamikanra A, Ako-Nai AK,
 Ogunniyi DA. Transferable
 antibiotic resistance in
 Escherichia coli isolated form
 healthy Nigerian school
 children: Intern.
 J.Antimicrob. Agents. 1996;
 7:59-64.
- 32. Fujita K. Hospital infection in paediatric patients. Hokaido. Igaku. Zasshi. 1994; **69 (2)**: 161-65.
- File FM. Overview of resistance in the 1990s.
 Chest. 1999; 115 (3): 35-85.
- 34. Archer GL, Johnston JL. Self-transmissible plasmids in staphylococci that encode resistance to aminoglycosides. Antimicrob. Agents. Chemother. 1983; 24 (1): 70-7
- 35. Jaffe HW, Sweeney HM,
 Nathan C et al. Identity and
 interspecific transfer of
 gentamicin-resistance
 plasmid in Staphylococcus

- aureus and Staphylococcus epidermidis. J. Infect. Dis. 1980 : 141(6): 738-47
- 36. Goldstein E.J. Prevention of bacterial resistance among urinary tract infections. Eur. Urol. 1991; 19 Suppl: 8-32.
- 37. Rubinstein E, Mark Z, Karen G, et al. Comparative activity of ofloxacin with reference to bacterial strains isolated in inpatients and out-patients. Infects. 1986; 14 (Suppl 10): 520-25.
- 38. Wenzel RP, Edmond MB.
 Vancomycin resistant
 Staphylococcus aureus:
 infection control
 considerations. Clin. Infect. Dis.
 1998; 27(2): 245-49.
- 39. De Man P, Verhoeven BAN, Verbrugh HA, et al. An antibiotic policy to prevent emegence of resistant bacilli. The Lancet. 2000; 355: 973-78.
- 40. Leverstein-van Hall MA, Fluit AC, Blok HE, et al. Control of nosocomial multi-resistant enterobacteriaceae using temporary restrictive antibiotic agent policy. Eur. J. Microbiol. Infect. Dis. 2001; 20 (11); 785-91.

BACTERIOLOGY OF CHRONIC SINUSITIS IN ILORIN, NIGERIA

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A prospective study of the bacteriology of 120 patients with chronic sinusitis and 55 control subjects seen between January 1995 and December 1998 in the Ear, Nose and Throat (ENT) Diseases Clinic of University of Ilorin Teaching Hospital, Ilorin, Nigeria was made. Whereas all cultures from the control group yielded only Staphylococcus (63.6% Coagulase positive and 36.4% Coagulase negative), cultures of patients with chronic sinusitis yielded *Staphylococcus aureus* (48.1%), *Escherichia coli* (20.4%), *Klebsiella spp.* (20.4%), Streptococci (7.4%) and *Streptococcus pneumoniae* (3.7%). The isolates were 100% sensitive to Ofloxacin, while penicillin was the least effective antimicrobial agent across board.

It was concluded that because of the difficulty in differentiating pathogenic organisms from commensals, the result of nasal swabs should be interpreted with caution. However, non-otolaryngologists involved in the management of the vast majority of patients with chronic sinusitis should request a carefully obtained posterior nasal mucosal swab.

INTRODUCTION

Chronic sinusitis is a common otolaryngologic disease worldwide. and particularly so in developingcountries (1-7). The aetiology of sinusitis is often multifactorial with considerable overlap clinical manifestations Purulent nasal discharge may not necessarily signify infection. Nasal with discharge very eosinophil count associated with allergic rhinitis may appear yellow or green (8). Also, clear nasal discharge may not always be of allergic origin.

There are multiple problems associated with management of chronic sinusitis. The difficulty in interpreting nasal mucosal swabcultures as opposed to sinus

cavity samples is the main reason for preferring the latter in the management of infective chronic sinusitis as they yield Review pathogens. of several studies show that the normal commensal flora of the nasal vestibule include Staphulococcus epidermidis, Micrococcus, Staphylococcus aureus. Diphtheroids and Gram negative bacteria, while that of the posterior nasal fossa include Streptococcus pneumoniae, **Haemophilus** influenzae, Streptococcus pyogenes, Staphylococcus aureus. meningitidis and Gram negative bacteria (8). These results raise doubts on the usefulness of nasal swab bacteriology in the treatment of infective rhinosinusitis (8-11). Apart from sampling technique, inappropriate transport method

could also interfere with optimal conditions thereby inhibiting the growth of fastidious organisms, while promoting the overgrowth of commensal flora.

As a result of failure to establish causative pathogens before starting treatment, there is tendency to over prescribe antibiotics and hence development of drug resistant (12,13).Inspite organisms awareness of these potential problems, most cases of chronic sinusitis are being managed by general practitioners, with no access to obtaining specimens of sinus cavity. Therefore, this study was undertaken to establish the reliability of results of nasal culture in the management of chronic. sinusitis and to recommend precautions which when taken, will maximize the effectiveness of treatment chronic sinusitis.

MATERIALS AND METHODS

A prospective study of 120 consecutive patients with chronic sinusitis seen between January 1995 and December 1998 in the Ear, Nose and Throat (ENT) diseases clinic of the University of Ilorin Teaching Hospital, Ilorin Nigeria was made.

Fifty-five control subjects, not known to be chronic sinusitis patients, and who had no symptoms of sinusitis over the

preceding month before sampling, were enrolled into the study. A provisional diagnosis of chronic sinusitis was made if there were at least two of the following signs and symptoms for duration of at least two months (14); nasal blockage, discharge (mucoid, nasal mucopurulent or purulent), postnasal drip, excessive sneezing and halitosis. Allergic, vasomotor and infective processes were not differentiated since these may coexist or one may lead to the other.

Nasal swab specimens were carefully obtained from patients by on first clinic ENT surgeons attendance after thorough physical with examination documentation of their relevant biodata, (e.g. age, sex) and drug history. Very slender nasal swabs were used to collect the exudate from the posterior nasal mucosa. In cases where the nasal cavity was filled with pus, this was initially suctioned to allow access to the posterior nasal mucosa. Patients on antimicrobial therapy within 72 presentation of were hours excluded from the study. The same procedure was followed for the control group.

Specimens so collected were immediately sent to the University of Ilorin Teaching Hospital laboratory (within a short distance) where streaking on Chocolate, MacConkey and Saborauds Dextrose Agar plates, were carried

out and incubated aerobically at 37°C for 18-24 hours. Isolates were identified by standard methods (15).

The antimicrobial susceptibility patterns of isolates were determined by standard disc diffusion methods (15).Mueller-Hinton agar employing Habdisc multidiscs with following antibiotics: penicillin 1 mu, gentamycin 10µg, colistin 25µg, cloxacillin 5µg, clotrimazole tetracycline 25µg. 10µg, erythromycin 5µg and chloramphenicol 10µg. Single disc of ofloxacin 10µg and cefuroxime 30µg were added where applicable. Growth inhibition zone diameter was measured in millimeters with a calibrated ruler after 18-24 hours incubation at 37°C. Results were interpreted as susceptible or resistant, while Staphylococcus aureus (NCTC 6571) and Escherichia coli (NCTC 10413), served as control organisms. **Facilities** for retrieving culturing anaerobic bacteria were lacking in the centre during the period of the study.

RESULTS

A total of 120 posterior nasal swabs from 58 males and 62 females were examined. The age range was 5-70 years. Less than 5% of patients were in the paediatric age group.

Fifty-four patients (45%) had pathogenic isolate. Staphylococcus aureus (48.1%), Escherichia coli (20.4%), Klebsiella spp. (20.4%), (7.8%);Streptococci and Streptococcus pneumoniae (3.7%) (Table I). Of the 55 control subjects studied (16 females and 39 males between ages of 18 and 55years), 22 (40%) were culture positive. exclusively Their isolates were Staphylococci (63.6% coagulase positive 36.4% coagulase and negative).

As shown in Table II, all the isolates were susceptible to ofloxacin, while penicillin was the least effective antimicrobial agent across board.

Table 1: Significant isolates from patients with Chronic Sinusitis in Ilorin, Nigeria.

Organism	Frequency (%) (N=54)
Staph. aureus	26(48.1)
E.coli	11(20.4)
Kebsiella spp	11(20.4)
Streptococci	4(7.4)
Strept. pneumoniae	2(3.7)
Total	54(100)

Table II: Antimicrobial Susceptibility Patterns of bacterial isolate

Antimicrobial Agent (Disk Concentration)	at (Disk n=26 n=4 n=2		S.pneumoniae n=2	E.coli n=11	K.pneumoniae n=11
Ampicillin (25ug)	3(11.5)	2(50.0)	1(50.0)	2(18.2)	0(0)
Penicillín Iug	O(O)	3(75.0)	1(50.0)	0(0)	0(0)
Erythromycin (5ug)	21(80.8)	4(100.0)	2(100)	-	-
Tetracycline 10ug	5(19.2)	3(75.0)	2(100)	8(72.3)	6(54.5)
Cloxacillin (10ug)	23(88.5)	4(100)	2(100)	-	-
Gentamycin (10ug)	16(61.5)	2(50.0)	1(50.0)	10(90.1)	8(72.8)
Cotrimoxazole (25ug)	6(23.1)	3(75.0)	1(50.0) 8(72.8		8(72.8)
Chloramphenicol (10ug)	12(46.2)	3(75.0)	1(50.0)	÷	
Colistin (25ug)	-	-		11(100)	11(100)
Ofloxacin (10ug)	26(100)	4(100)	2(100)	11(100)	11(100)
Cefuroxime (30ug)	20(77.0)	4(100)	2(100)	10(90.1)	10(90.1)

Note: - - Not tested.

DISCUSSION

Although a few studies suggest that paranasal sinuses are normally sterile, both aerobic and anaerobic bacteria have been recovered from sinus aspirates of presumably normal human subjects (10,16,17). It is however, generally agreed that the knowledge

of the bacteriology of secretions obtained directly from maxillary sinus by needle aspiration (with avoidance careful contamination from mucosa surfaces), provides a more reliable information essential the planning of anti-microbial therapy (8,10,11). Thus in decreasing order of reliability, sample sites can be arranged in the following sequence; sinus cavity, sinus ostium, posterior nasal mucosa and anterior nasal mucosa (8-11). the majority of For otolaryngologists involved in the management of chronic sinusitis the posterior nasal fossa recommended for obtaining contamination specimens but should avoided and be limitations in interpreting reports of such culture recognized.

As reported else where (18-21), Staphylococcus aureus was the commonest pathogen associated with chronic sinusitis in this study. However, there are increasing recent reports of prevalence of enteric Gramnegative bacilli in this condition (17). Although, due to lack of facilities, anaerooic organisms were not cultured in this study. It has been documented anaėrobic organisms play a major chronic sinusitis role in (8,9,19,22). Fungal sinusitis is rare, and its presence should raise

the suspicion of an immune deficiency disorder (12).

All isolates in this study were susceptible to ofloxacin 100% was while (cefuroxime second) penicillin was the least effective anti microbial board. agent across gram-negative Although, the organisms susceptible were colistin, it is no longer in common use because it is ototoxic. This study suggests that ofloxacin is the drug of choice for the management chronic sinusitis in environment. But where ìŧ is as in children, contraindicated cefuroxime may be employed, as it most second effective antibiotic in this study. Although, some earlier studies recommended gentamicin and colistin, which are ototoxic, most recent studies also recommend newer drugs such as ofloxacin and amoxicillinclavulanate as the drugs of first choice (13,16,22). Due to the high frequency, of anaerobic organism as recorded in literature, an empirical trial of metronidazole is warranted by clinical suspicion, in cases not responding to the recommended regimen (8,22).

This study confirms that one must be careful in interpreting the result of nasal swabs because of the difficulty in differentiating pathogens from commensals. The establishment of Ear, Nose, Throat Disease and Laboratory Centers with full compliment and facilities

for reliable diagnosis and treatment of chronic sinusitis is recommended.

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REFERENCES

- 1. Scadding G. Rhinitis: Common, debilitating and treatable. *The Practitioner*. 1996; **240**: 48-53.
- Burhholter D, Schiffe P. The epidemiology of atopic disease in Europe-A review.
 Allergy Clin. Immunol. News.
 1995; 7: 113-25.
- Al-Samarrae SM. Treatment of "vasomotor rhinitis" by the local application of Silver Nitrate. J. Laryngol. Otol. 1991; 105: 285-287.
- Okafor BC. Otolaryngology in South Eastern Nigeria II: Pattern of Disease of the Nose Nig. Med. Pract. 1983;
 13:21-29.
- Obiakor MN. Cattarrh. Nig. Med. Pract. 1987; 14 (5/6): 87-93.

50,000,00

- Bhatia PC, Varughese R. Pattern of Otolarygological Disease in Jos community.
 Nig. Med. J. 1987; 17: 67-93.
- Ezeanolue BC, Aneke EC, Nwagbo DFE. Correlation of plain radiological diagnostic features with antral lavage results in chronic maxillary sinusitis. WAJM. 2000; 19 (1): 16-18.
- 8. Weir N, Golding-Wood DG.
 Infective rhinitis and sinusitis. In: Kerr AG,
 Mackay IS, Bull TR (eds.)
 Scott Browns Otolaryngology.
 6thed.Butterworth/Heineman:
 Oxford. 1997 : 4/3/1-4/3/22.
- Verschraegen G, Mione S. Difficulties in interpretation of culture results in sinusitis.
 Rhinology. 1998; 36 (2): 55-58.
- 10. Wald ER. Sinusitis in infants and children. Ann. Otol. Rhino. Laryngol. 1992; 101: 37-41.
- 11. Jiang RS, Hsu CY, Jang JW. Bacteriology of the maxillary and ethmoid sinuses in chronic sinusitis. J. Laryngol. Otol. 1998; 112 (9): 845-8.

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- 12. Slack RA Realistic view of sinusitis. *ThePractitioner*. 1996; **240**:54-57.
- 13. Ijaduola G T A, Anyiwo C E. Microbiological study of chronic maxillary sinusitis in Nigerian adults. Nig. J. Microbiol. 1984; 4(1-2): 49 54.
- 14. Richtsmeier W J. Medical and surgical management of sinusitis in adults. Ann. Otol. Rhinol. 1992; 101: 46-50.
- 15. Barrow GL, Feithan RKA. Cowan and Steel's manual for the identification of medical bacteria. 3rd edition, Cambridge University Press 1993: 106-108.
- Klossek JM, Dubreuil L, Richet H, et al. Bacteriology of chronic purulent secretions in chronic rhinosinusitis. J. Larngol. Otol. 1998; 112(12): 1162-66.
- 17. Brook I. Aerobic and anaerobic bacterial flora of normal maxillary sinuses.

 Laryngoscope. 1981; 91:372-76.

.. .

- Rhamadan HH. What is the bacteriology of chronic sinusitis in adults? Am. J. Otolaryngol. 1995; 16 (5): 303-306.
- 19. Hsu J, Lanza DC, Kennedy DW. Anti microbial resistance in bacterial chronic sinusitis. Am. J. Rhinol. 1998; 12 (4): 243-48.
- 20. Busaba NY, Siegel N, Salman SD. Bacteriology of non-traumatic maxillary sinus mucoceles versus chronic sinusitis. *Laryngoscope*. 2000; **110 (6)**: 969-71.
- 21. Erkan M, Asian T, Ozcan M, Koc N. Bacteriology of antrum in adults with chronic maxillary sinusitis.

 Laryngoscope. 1994; 104
 (3pt1): 321-24.
- 22. Gehanno P, Cohen B. Effectiveness and safety of ofloxacin in chronic otitis media and chronic sinusitis in adult out-patients. Eur. Arch. Otorhinolaryngology. 1993; 250 (1): 513-514.

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CONTROL OF REPATITIS B INFECTION

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Hepatitis B infection remains a public health problem word-wide because of its high endemicity in most countries of the world and the presence of the virus in many body fluids/secretions, which is responsible for several alterative modes of transmissions. Also, the fact that most of the hepatitis B virus (HBV) infections occur in infancy or early childhood where over 70% of the infections is asymptomatic and nearly over half of the primary infection results in chronic infection/ carrier is another reason. Most of the HBV related chronic liver diseases are associated with the chronic infection, which manifestations occur in adulthood. This article examined the various measures that can be used in the control of the disease taking into consideration the epidemiological factors responsible for the occurrence and distribution of the disease.

INTRODUCTION

Hepatitis B virus infection is a major public health problem world wide. About 2 billion people serological have evidence of infection with HBV (1), and it is estimated that 350 million of them have chronic HBV infection, while about a million die each year from chronic diseases, including cirrhosis and cancer of the liver (2). Man is the only reservoir of the infection, and infected person can have both short-term and long-term When infected. outcomes. person can have either symptomatic disease (acute hepatitis Βì or **symptomatic** with infection no signs symptoms of the disease (2,3). In either case, the individual may

either recover from the infection and develop life long immunity or develop a chronic infection that usually last throughout life. In persons with acute hepatitis B the incubation period after becoming infected is usually 3-4 months, with a range of 6 weeks to 6 months. About 1-2% of those with acute infection die from fulminating hepatitis (2), while about 5% will become chronic carriers (3). Most of the disease burden associated with HBV infection is related to the chronic condition of the disease (2).

THE EPIDEMIOLOGY OF HEPATITIS B INFECTION

Although most HBV related chronic liver disease manifest themselves in adulthood, the infection usually begins in infancy or early childhood in hyper endemic areas. Several factors including

of infection. initial age immunization status of the host and viral factors are known to affect the natural course of the disease (4). The age at which infection occurs is an important determinant factor of the outcome of the disease (2.4.5). Only about 10% of infections occurring among children under the age of 5 years are symptomatic, while 80-90% of infants infected during the first year of life and 30-50% of children infected between 1-4 years of age develop the chronic condition (5,6). However, 30-50% of adults symptomatic when first are infected but only 2-5% become chronically infected (2).

About 45% of the world's population live in areas where chronic HBV infection is highly endemic (i.e. 8% or more of the population are HBsAg positive), 43% are in areas of intermediate endemicity (2-7% are HBsAg~ postive) and 12% live in areas of low endemicity (<2% are HBsAg positive) (2). The infection transmitted either by skin puncture or mucous membrane contact with infected blood or other body fluids. The virus is found in most body fluids / secretions such as blood, wound secretion, vaginal fluid, semen, and urine in varying saliva concentrations. highest The concentration of the virus occurs in blood and wound secretions.

Moderate concentrations of HBV are found in semen and vaginal fluid and lower concentration in sativa and urine (7). The presence of the virus in several body fluid / secretion justify many alternative modes of spread of the disease like perinatal transmission, spread through close personal contact, unsafe injection or use of sharps, contact and blood sexual transfusion.

Perinatal transmission from mothers infected with HBV (i.e. positive for hepatitis B Surface Antigen HBsAg) to their newborn infants is a major source of HBV infection in many countries (4,6) and this usually occurs at the time of birth. Transmission in utero is very low, less than 2% (2) and there is no evidence to support claims of transmission through breast milk (8). Any mother who is a carrier for HBV (i.e. HBsAg positive) can transmit infection the the newborn; however, the risk of perinatal transmission is higher if such mother has hepatitis B e antigen (HBeAg) in her blood. The risk of transmission is in the range of 70-90% in mothers who are HBeAg positive and about 5-20% among those who are **HBeAg** negative (7).

The spread from child to child account for most HBV infection occurring in household settings (9,10), but may also occur in child day care centers and pre-nursery

school (11). The most probable of child-to-child mechanisms transmission involve contact of skin sores, breaks in skin or mucous membranes with blood or skin sore secretions (7). The disease may also spread because of contact with saliva through bites or other breaks in the skin (12,13). Also, the virus may spread from inanimate objects such as shared towels or tooth brushes and other formites since the virus can survive for at least 7 days out side the body and can be found in high titres on objects, even in the absence of visible blood (14).

Transmission associated with unsafe injection practices is a major mode of transmission of the infection in adults in many countries (15, 16),while transmission through blood transfusion is a major source of HBV infection in countries where blood is not screened for HBsAg especially in developing countries (2). Sexual transmission of HBV accounts for a high proportion of new hepatitis B infection among adolescents and adults in countries with low and intermediate endemicity of chronic HBV infection (17). In countries infection is highly where the endemic. sexual transmission does not account for a significant percentage of cases because most people are already infected during childhood. In these high endemic

areas the lifetime risk of HBV infection is more than 60% and most infection are acquired from and child-to-child perinatal transmission (2), when the risk of the developing chronic infection is greatest. The implication here is prevalence of that the acute hepatitis B infection in these areas would be very low since most early childhood perinatal and infections asymptomatic. are However, rate of liver cirrhosis and cancer in adults would be very areas of intermediate high. In endemicity, the lifetime risk of infection is 20-60%, and infections occur in all age groups. Whereas, in low endemicity, the lifetime risk of infection is less than 20%, but most of the HBV infections occur in adults in relatively defined risk groups (2).

CONTROL OF HEPATITIS B INFECTION

of the burden Most associated with HBV infection is related to the chronic condition of the disease resulting from carrier state, and the highest carrier rate occurs in children. These children are not only at risk themselves for the long term complications of HBV but also constitute an important reservoir and source of hepatitis B infection within the community at large. Therefore they constitute important target group that must be focused in any HBV control

programme. The following levels of disease prevention and control useful will be in measures public health reducing the associated with the problem disease.

1. Health promotion through health education is an important strategy in the control of HBV infection (particularly in children). The general public, especially the parents, and the health workers should be targeted in order to create awareness on the disease and solicit necessary support and required cooperation intervention. The key message of education health should the the mode highlight transmission, people at risk of infection, the burden of disease on individual and the nation and the importance of safe injection and blood transfusion practices and use of sterile Sharps. This would go a long way to help parents understand the disease and take measure to protect their children against HBV infection (2).

The role of health education in the control of this disease is evident by the result of a study conducted in Poland where hepatitis B prevention through health education programme was evaluated. The study showed that health education has impact in the reduction of the incidence of HBV infection (18).

of 2. Specific protection children and other at risk groups through vaccination is the most useful strategy. Vaccination against HBV has been well documented to be cost-effective in the prevention and control of the disease (2,19,20). infant hepatitis Routine should be a high vaccination priority for all countries regardless of the degree of endemicity of the disease, while vaccination for other groups at risk of infection will depend on the epidemiological patterns, economic factors, cultural sexual practices of each country (2). Infant immunization chronic infections prevents acquired during early childhood, preventing the consequences of HBV infection. Strategies targeting adolescents and adults risk groups have failed to control HBV adequately because of the difficulty of immunizing people in many risk groups before they initiate high risk behaviours and also due to the fact that infections can occur among people with no identified high risk factor (2). Therefore, routine immunization of infants is most desirable in the control of the disease.

The World Health Organization (WHO) has recommended that hepatitis B vaccine be included in routine immunization schedules for all children in all countries of the world (21), and many countries

have complied by incorporating, through policy and or practice, the vaccine in to heir routine immunization programmes (2, 22-However, an acceptable universal coverage is yet to be achieved (2,24,25). Hepatitis B vaccines are available in monovalent formulation that protect only against hepatitis B and in combination formulations that protect against hepatitis B and other diseases (e.g. DPT-Hep B, DPT-Hep B+Hib, Hib-Hep B). movovalent hepatitis B vaccine has an advantage over the combination vaccines because it can be used for birth dose unlike the combination vaccines have other components that cannot be administered at birth.

pre-exposure In immunization, a course of 3 doses of hepatitis B vaccine induces protective levels of antibody to HBsAg (Anti-HBs) in over 95% of healthy infants and children when given in a variety of schedules (26,27). Children who respond to the vaccine are protected against acute hepatitis B and chronic infection. Studies have shown that those who respond to a 3-dose hepatitis B immunization series are protected for as long as 15 years (28). Long-time protection relies on immunological memory that allows a protective anamnestic antibody response (29).after exposure to HBV

Therefore, booster doses are not recommended (2, 30). Postexposure immunization, beginning at birth with either hepatitis B vaccine alone or combine with hepatitis В immune globuline (HBIG) can prevent the spread of more than 90% of HBV infections from mother to baby. However, the efficacy of giving recombinant hepatitis B vaccine alone is similar to that of giving hepatitis B vaccine with HBIG. (28). Hence, the use of may not be necessary. Optimum efficacy in preventing perinatal HBV infections newborn can be achieved when the vaccine is given within 24hours after birth /exposure. There is no evidence of protection against perinatal transmission if the first dose of vaccine is given more than 7 days after birth (2) - Administration HBIG confers protection individual accidentally exposed following needle stick injuries or sexual partners of acute cases.

3. Early diagnosis/case detection: Prenatal screening of all pregnant women for HBsAg and HBeAg is useful in identifying mothers at risk of transmitting the infection to the newborn. This requires a lot of resources to screen these women and track infants of

cted mothers in order to ensure completion of the vaccine series, especially in developing countries where some pregnant women do not go for antenatal care or where many

are conducted bv deliveries unskilled person or take place out side the health facility. Prenatal screening should therefore to supplement routine childhood immunization (2). Every country should also adopt policy of safe blood transfusion ensuring that all blood are screened for before HBsAg blood transfusion. All bank services should be strengthened in this area.

4. Treatment of hepatitis B infection is mainly supportive. Since chronic infection for the burden accounts associated with the disease, treatment of chronic hepatitis B infection using interferon antiviral analogues would be an important control measure. Many studies have reported some degree of safety of the interferon when use in the treatment of chronic condition. The effectiveness. however, is in the range of 33-43% when use in the treatment of chronic hepatitis infection in children (31-33). However, when you consider the large number of worldwide. infected people vaccination remained the most cost-effective option in the disease (2,31).

5. Surveillance system in HBV infection involves an exercise of continuous scrutiny over the occurrence and spread of the disease and related consequences

such accuracy with completeness to provide basis for effective control. It. involves systematic data collection analysis and consolidation. evaluation in order to discern long term trends and epidemiological patterns resulting from the control measures put in place. Reporting of acute hepatitis B infection can be integrated into the existing notification and surveillance system for easy data generation since children do not usually acquiring symptoms on infection, but tend to develop infection. data chronic occurrence of chronic liver diseases (cirrhosis and liver cancer) must also be collected to be able to estimate the magnitude and burden of the disease over time. However, the interpretation of the take generated must into consideration other aetiological factors that can lead to chronic liver diseases (2).

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REFERENCES

- Kane MA. Global status of hepatitis B immunization. Lancet. 1996; 348: 696-700.
- 2. World Health Organization (WHO). Introduction of hepatitis B vaccine into childhood immunization services: WHO document, Geneva (WHO / V& B / 01.31): 1-30.
- Macleod J. Diseases of liver and biliary system. In: Davidson's Principles and Practice of Medicine. 14th edition. Longman, London; 1984.
- 4. Chang MH. Hepatitis B virus infection in children: epidemiology, natural course and prevention in Taiwan. Formos. Med.Association. 1996; 95(8): 593-98.
- 5. Juszczyk J. Clinical course and consequences of hepatitis B infection. *Vaccine*. 2000; **18(18)**: 523-25.
- 6. Bortolotti F. Chronic viral hepatitis in childhood. Bailieres Clin. Gastroentrol. 1996; 10(2): 1185-1206.

- Margolis HS, Alter MJ, Hadler SC. Viral hepatitis. In: Evans AS, Kalow RA. Viral infection of humans, epidemmiology and control. 4th edition. Plenum Polishing Co. New York. 1997: 376-403.
- 8. Beasley RP. Evidence against breast-feeding as mechanism for vertical transmission of hepatitis B. Lancet. 1975; 2: 740-741.
- 9. Davis LG, Weber DJ, Lemon SM. Horizontal transmission of hepatitis B virus. *Lancet*. 1989; 1:889-893.
- 10. Hurie MB, Mast EE, Davis JP. Horizontal transmission of hepatitis B virus infection to United States-born children of Hmong refugees. *Pediatrics*. 1992; **89**: 269-273.
- 11. Nigro G, Taliane G. Nursery-acquired asymptomatic B hepatitis. *Lancet.* 1989; 1: 1451-1452.
- 12. Beasley RP, Hwang LY. Postnatal infectivity of hepatitis B surface antigencarrier mothers. J. Infect. Dis. 1983; 147: 185-190.
- 13. Macquatrie MB, Forghani B, Wolochow DA. Hepatitis B transmitted by a human bite.

 JAMA . 1974; 230: 723-724.

- 14. Martinson FE. Risk factor for horizontal transmission of hepatitis B virus in a rural district in Ghana. Am.J. Epidemiol. 1998; 147: 478-487.
- Kane A, Lloyd J, Zaffaran M, 15. Transmission al.hepatitis B, hepatitis C and immunodeficiency human viruses through unsafe injections in developing world: modelbased regional estimates. Bulletin of World Health Organization, 1999; 77: 801-807.
- 16. Simonsen L, kane A, Lloyd J, Zaffran M, Kane M. Unsafe injections in the developing world and transmission of blood borne pathogens: a review. Bulletin of the World Health Organization. 1999; 77:. 789-800
- 17. Alter MJ, Margolis HS. The emergence of hepatitis B as a sexually transmitted disease.

 Med. Clin. North America.
 1990; 74: 1529-1541.
- Przewłocka T. Evaluation of the hepatitis B prevention education programme in Poland. Vaccine. 2000;
 18(18): S46-8.

- 19. Wilson N, Ruff TA, Rana BJ, et al. The effectiveness of the infant hepatitis B immunization programme in Fiji, Kiribiti, Tonga and Vanuatu. Vaccine. 2000; 18(26): 3059-66.
- 20. Ahn YO. Strategy for vaccination against hepatitis B in areas with high endemicity: focus on Korea. Gut. 1996; 38 suppl 2: S63-66.
- 21. World Health Organization (WHO). Expanded Programme on Immunizaton; Global advisory group part 1. Weekly Epidemiological Report; 1992; 67: 11-15.
- Federal Ministy of Health,
 Nigeria. National Immunization
 Policy and Standard of
 Practice. 1995: 2-25.
- 23. World Health organization (WHO). Expanded programme on Immuniation (EPI): Immunization schedule in the WHO eastern meditrranian region. Weekly Epidemiological Record, 1996; 71(23): 173-76.
- 24. Woodruff BA, Stevenson J,
 Yusuff H, et al. Progress
 toward integrating hepatitis B
 vaccine into routine infant
 immunization schedules in the
 United States, 1991 through

- 1994. Connecticut hepatitis B project Group. *Paediatrics*. 1996; **97(6)**: 798-803
- 25. Chang MH, Chen DS.
 Prospect for hepatitis B virus
 eradication and control of
 hepatocellular
 carcinoma. Baillieres-Best
 Prac. Res. Clin. Gastroentrol.
 1999; 13(4): 511-517.
- 26. Hadler SC, Margolis HS. Hepatitis B immunization: vaccine types, efficacy and indications for immunization. In: Remington JS, Swartz MN (eds). Current topics infectious diseases, Vol.12. Scientific Blackwell Publications, Boston, 1992: 282-308.
- 27. **Kocks** Apinall S. DJ. Immunogenicity of a low-cost hepatitis B vaccine in the South African Expanded Programme on Immunization. South Afr. Med. J. 1998; 88: 36-38
- 28. Andre FE, Zuckerman AJ.
 Review: protective efficacy of
 hepatitis B vaccine in
 neonates. J. Med. Virol.
 1994; 44: 144-151

- 29. West DJ. Calandra GB. Vaccine induced immunological memory for hepatitis B surface antigen implications for policy booster vaccination. Vaccine. 1996; 14: 1019-1027
- European Consensus Group on Hepatitis B Immunity. Are booster immunization needed for lifelong immunity. Lancet. 2000; 355: 561-565
- 31. Neudort-Grauss R, Bujanover Y, Dinari G, et al. Chronic hepatitis B virus in Isreal, clinical and epidemiological characteristics and response to interferon therapy. Isr. Med. Ass. J. 2000; 2(2): 164-168
- 32. Gurakan F, kocak N, Ozen H, Yuce A. Comparison of standard and high dosage recombinant interferon alpha 2b for treatment of children chronic hepatitis B infection. Pediatr. Infect. Dis. J. 2000; 19(1): 52-6
- 33. Ozen H, Kocak N, Yuce A, Gurakan F. Retreatment with higher dose of interferon alpha in children with chronic hepatitis B infection. *Pediatr. Infect. Dis. J.* 1999; **18(18)**: 694-697.

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PREVALENCE OF TRICHOMONAS VAGINALIS AMONG THE SEXUAL PARTNERS OF WOMEN WITH TRICHOMONIASIS IN IBADAN, NIGERIA.

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This study was conducted to determine the prevalence of *Trichomonas vaginalis* among the sexual partners of women.with Trichomoniasis. While 103 female patients were found to have *T. vaginalis* using both direct wet mount microscopy and culture, only 44(42.7%) male contacts reported for screening. *Trichomonas vaginalis* was isolated from 20(45.5%) of the 44 male contact investigated, whilst there was no laboratory evidence of the agent in 13(29.6%) of the study group. Amongst the 60 control subjects, *T. vaginalis* was recovered from 7(11.7%) of them. The difference in the occurrence of *T. vaginalis* between the patients and the control group was highly statistically significant (p<0.001). This implies that in most cases, infestation with *T. vaginalis* in the male is sitent and might represent a carrier state. Eight (40%) of the 20 men with *Trichomonas vaginalis* and 43.2% of the 44 men investigated were between 20 and 29 years and this parallels that of other sexually transmitted diseases. While only 6(30%) of the men with *T. vaginalis* were diagnosed by direct wet mount preparation, all the 20 (100%) positive cases were diagnosed by culture. This however, indicated the general superiority of culture over fresh smears but it is desirable to use both methods in the diagnosis of Trichomoniasis since they complement each other.

Key Words: Trichomonas vaginalis, Prevalence, Sexual Partners, Trichomoniasis, Ibadan.

INTRODUCTION

Trichomonas vaginalis has been incriminated as one of the common sexually transmitted genital pathogens and is found to be associated with approximately 20% of all cases of non-gonococal urenthritis (NGU) (1).Trichomoniasis is widely distributed all over the world and remains common infection among female patients attending sexually transmitted disease clinics. However, in spite of the fact that the presence Trichomonas vaginalis in the male urinary tract was discovered as early as 1883 by Kuntler, there is scanty information on it's prevalence in the male patients

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asymptomatic cases and limited nature of the infection (2). While majority of female patients harbouring this organism present with vaginal discharge, which is usually frothy, greenish-yellow and offensive, the infection in the male may give rise to no symptom or present low-grade as nongonococcal urethritis (2). Unlike pre-pubertal gonococcal vaginitis cases occasionally Ibadan, encounterd in Trichomoniasis among this age group is very rare (3). Dunlop and Wisdom (4) found no T. vaginalis even in post pubertal virgins from the study done in 1965, which confirms sexual contact as the main mode of transmission. The fact that vaginitis due to T.vaginalis continues to cause considerable and prolonged distress to many women and a high incidence of relapse even after a considerable period of freedom from symptoms because of re-infection by coitus, the study of the infected male becomes a matter of considerable importance (5).**Trichomoniasis** has been frequently found among the sexual partners of patients with proven infection and infact, the parasite has demonstrated in 30-40% of male sexual partners of infected women (6-11). A prevalence of 70% was observed among men who had with sexual contact infected women within the previous 48hr. and only 33% of men were infected if their last contact was 2 weeks previously (12). Several studies have suggested an increased cure rate in women following treatment of their regular sexual partner (13-16). Apart from the retrospective studies done in Ibadan by Osoba (17) and Alausa (18) in 1972 and 1974 respectively, no detailed study has been conducted in our environment to determine of **Trichomonas** prevalence vaginalis among the male sexual partners of infected women even though these sexual contacts are being treated epidemiologically. The aim of the present study was to determine the prevalence of Trichomonas vaginalis among the sexual partners of women with Trichomoniasis

MATERAL AND METHODS Study population

The study population comprised all sexual contacts of women with confirmed diagnosis of Trichomonas vaginalis infection who were invited for screening. These represented a highly selected group of men who had undoubtedly been exposed frequently to Trichomonas vaginalis. Each patient was questioned as regards his marital previous genito-urinary symptoms, data of last coitus and other sexual consorts (whether casual or regular). The previous antimicrobial drugs used before attending the clinic were noted. A clinical examination of the lower genito-urinary tract for signs of infection such as urethral discharge and the nature of the discharge was carried out.

Control group

Sixty male patients randomly selected among the male patients that attended Special Treatment Clinic, University Collage Hospital, Ibadan, but without symptoms during the period of study were entered into the study. This group represented a cross-section of presumable sexually active male, some of which reported because of fear of venereal disease but had no infection.

Laboratory procedures

The urethral specimens were examined for the presence of Trichomonas vaginalis by agitating the cotton swab in 1ml of saline in a tube and a drop of resulting transferred suspension microscope slide. The preparation was covered with a cover slip and then examined at x100 and x400. A Gram-stained smear was also examined for the presence of Gram-negative intracellular diplococci, other bacteria and cells. veast The urethral specimens were then plated on modified Thayer-Martin medium and incubated using standard (19).Culture for method vaqinalis was performed using Nutrient Broth Glucose Serum medium (20). Examination for presence of T. vaginalis was made at 48 hours and 5 days of by incubation making a sediment from mount of the bottom of the Bijou bottles containing the medium and motile T. vaginalis were searched for.

Data analysis

Statistical analysis was performed using the student t and the chi squared tests.

RESULTS

During the period of study, 103 female patients were found to have *Trichomonas vaginalis* using both direct wet mount microscopy and culture whilst only 44 (42.7%) male contacts of the women reported for screening. While *Trichomonas vaginalis* was isolated from 20 (45.45%) of the 44 male

patients investigated, there was no laboratory evidence of the agent found in 13 (29.55%) of the study group (Table 1). There was evidence of urethritis in all the T. vaginalis positive cases as shown by the presence of increase in the number of polymorpho-nuclear leucocytes in voided urine and urethral swabs (>4-5/HPF). examined (15.9%) of the 44 male patients had non-specific urethritis (NSU) with no evidence of incriminating agent. Amongst the 60 control subjects, T.vaginalis was recovered from 7(11.7%) of them. The difference in occurrence of T.vaqinalis the the patients and the between highly control group was statistically significant (p<0.001).

Table 2 shows the distribution of the 44 male patients investigated, 43.2% were within the age of 20 to 29 years while 11.3% were over 50 years of age. Of the 20 men with Trichomonas vaginalis, 8(40%) were between the age of 20 and 29 years. While 39 (37.9%) of the female partners investigated single, 26 (25.2%)married, 11(10.7%) separated, 18 (17.5%)divorced and 9(8.7%) widowed (Table 3). Twenty six (59.1%) of the men investigated were single, 26.7% of whom had contact with casual consorts and 73.3% had contact with regular consorts. Of the 18 (40.9%) that were married; only 16.7% had contact with their wives. Table 4 compares the positive diagnostic rates by the different method used. While 6 (30%) of the 20 *T. vaginalis* isolated were diagnosed

by direct wet mount microscopy, all the 20 (100%) were isolated on culture. All the men with Trichomonas vaginalis were treated and requested to attend for a repeat examination after 2 weeks but seven failed to report.

Table 1: INCIDENCE OF Trichomonas vaginalis IN MALE PARTNERS OF INFECTED WOMEN. N=44.

Diagnosis	Number	Percentage
Trichomonas vaginalis	20	45.5
Candida albicans	3	6.8
T.V & Candida	1	2.3
NSU	7	15.9

Table 2: AGE DISTRIBUTION OF MEN INVESTIGATED n=44.

AGE (years)	Number	Percentage	No. of T.V.Positive Pts.	Percentage
10-19	4	9.1	1	5
20-29	19	43.2	8	40
30-39	8	18.2	3	15
40-49	8	18.2	6	30
> 50	5	11.3	2	10
TOTAL	44	100	20	100

Table 3: MARITAL STATUS OF FEMALE PARTNERS
OF MEN INVESTIGATED

Marital Status	Number	Percentage
Single	39	37.9
Married	26	25.2
Separated	11	10.7
Divorced	18	17.5
Widowed	9	8.7
TOTAL	103	100

Table 4: COMPARISON OF POSITIVE DIAGNOSTIC RATES BY DIFFERENT METHODS.

Wet mounts Prep	Culture	
No.of Positive n=20	6	20
Percentage Positive	30	100

P<0.001

DISCUSSION

Trichomonas vaginalis has been incriminated as one of the common sexually transmitted pathogens and although appears to be highly prevalent with a widespread geographical distribution, it has not been the focus of intensive study nor of active control programs (3, 21). Most patients with Trichomonas vaginalis infection are asymptomatic mildly or symptomatic and hence they are likely to continue to sexually active in spite of infection (21,22).While it has reasonably confirmed centre that T. vaginalis is a cause of non-gonococcal urethritis (NGU) (3), many of these patients still come for treatment because they are the sexual partners of women with symptomatic infection.

In this study, a significantly high proportion of male that are repeatedly exposed to T.vaginalis during sexual contact themselves harbour the agent. While 20 (45.5 %) sexual partners of women with Trichomoniasis had T. vaginalis isolated from their arethral swabs, 7(11.7%) of the 60 control subjects were positive. This implies that in most cases. infestation T.vaqinalis in the male is silent and might represent a carrier state.

Age distribution of male patients with *Trichomonas*

vaginalis parallels that of other sexually transmitted diseases (3, 23-25). A large percentage (43.2 %) of the men investigated as well as 40 % of those with T. vaginalis were within the age range of 20-29 years. This is the period of greatest sexual activity and those in this group tend to be promiscuous and are therefore prone to sexually transmitted diseases. Trichomoniasis has been found to be prevalent among patients with higher levels of sexual activity and larger number of different sexual partners (25). Twenty-six (59.1%) of the men investigated were single, 26.7 % of whom had contact with casual consorts and 73.3 percent regular consorts. with confirms the repeated exposure these men have to T.vaginalis during sexual contact. This is in contrast with the findings of the previous study where a percentage of the men investigated had contact with casual consorts (23). Extramarital exposure to risk of infection by casual contacts among married men also plays an important part in the transmission of infection (17) which may be due the fact that married men abstain from sexual intercourse with their wives during pregnancy and few years after delivery. In this study, 15(83.3%) of the 18 married men had contact with casual consorts whilst only 16.7% had contact with their wives. The data

the prevalence of on Trichomoniasis vary around the world, depending on the reliability of the diagnostic tests used on them and their response medical examination. During the period of study, 103 patients showed cultural evidence of Trichomoniasis and were given contact tracing paper for their partners but only 44 (42.7%) of the male contacts of the women for screening. came confirmed the attitude of the general population investigations of sexually transmitted diseases. In the less educated people, it is difficult to separate venereal diseases from genital infection of other kinds (3). To this set of people, any case of urethral discharge is gonorrhoea and genital ulcer is syphilis. In the case of Trichomoniasis that often lacks symptoms and signs, it is almost impossible to convince these people to come for screening. Some of them declined to be examined or investigated and in spite of the strict confidentiality in the clinic, many of those men invited queried the possibility of STI and repeated reassurance and explanation could not remove the lingering doubts. There conflicting reports on the role of T.vaginalis causing in complications in infected male patients. Compared other transmitted diseases, sexually

Trichomoniasis is relatively devoid of late complications. However, a few cases of acute, non-gonococcal epididymitis have been attributed to Trichomonal infection (1, 26-28). In vaginalis was some cases, T. observed in epididymal aspirates and it has been isolated from about 10 % of some series of infertile men (1.7), but such association does not imply causality. Some workers have observed phagocytosis spermatozoa by T. vaginalis (7), and others have noted that in the presence of large numbers of the agent in vitro, sperm mobility is decreased (1,11). Ogunbanjo et al (29), by retrospective study on infective factors of male infertility among Nigerians, found T. vaginalis to be one of the causative agents of conjugal infertility.

Many workers have found cultural method to be superior to other methods such as fresh wet mount examinations and staining (3,23,30). This has been confirmed by this study. While only six (30%) of men with T. vaginalis were diagnosed by direct wet mount preparation, all the twenty (100%) positive cases were diagnosed by culture. There is a high significant the difference between laboratory methods. This however, indicates the general superiority of culture over fresh smears but it is desirable to use both methods in the diagnosis of Trichomoniasis since they complement each other.

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REFERENCES

- Krieger JN. Urologoical aspects of Trichomoniasis. Invest. Urol. 1981; 18: 411.
- 2. Michael F, Rein MM.
 Protozoan infectionsTrichomonas vaginalis:
 sexually transmitted
 disease. J. Am. Ven. Dis.
 Ass.: 525.
- 3. Bakare RA, Ashiru JO, Adeyemi-Doro FAB, et al. Non-gonococcal urethritis (NGU) due to Trichomonas vaginalis in Ibadan. West Afr. J. Med. 1999; 18(1): 64-68.
- 4. Dunlop EMG, Wisdow AR. Diagnosis and management of Trichomoniasis in men and women. *Brit. Vener. Dis.* 1995; **14**:85.

- 5. Lancely F. Trichomonas vaginalis infection in the male. Brit. Vener. Dis. 1953; 29:213.
- 6. Honigberg B. Trichomonads of importance in human medicine. In: Kreier JP (ed). Parasitic protozoa. 1978; 2: 275.
- 7. Brown MT. Trichomniasis. Practitioner. 1972; **209**: 639.
- 8. Whittington MJ. Epidemiology of infection with Trichomonas vaginalis in the light of improved diagnostic methods. Br.J. Vener. Dis. 1957; 3:80.
- Gallai Z, Sylvestre L. The present status of urogenital Trichomoniasis: A general review of Literature.
 Appl. Therapeutics. 1966; 8: 773.
- 10. Block E. Occurrence of Trichomonas in sexual with partners of women Trichomoiasis. Acta Obstet. Gynaecol. Scand. 1959; **38**: 398.
- 11. Perl G, et al. Male urogenital Trichomoniasis. J. Mt. Sinai (NewYork). 1965; **32**: 495.

- 12. Weston TET, Nicol CS.

 Natural history of
 Trichomonal infection in
 males. Br. J. Vener.
 Dis. 1963; 39: 251.
- 13. Burch TA, et al. Epidemiological studies on human Trichomoniasis. Am. J. Trop. Med. Hyg. 1959; 8: 312.
- 14. Peterson WF, et al.

 Metronoidazole in pregnancy. Am. J. Obstet.

 Gynaecol. 1966; 94: 343.
- 15. Underlhill RA, Peck JE.
 Causes of therapeutic
 failure after treatment of
 Trichomonal vaginitis
 with Metronidazole:
 Comparison of single dose
 treatment with a standard
 regimen. Brit. J. Clin. Pract.
 1974; 28: 134.
- 16. Hager WD, et al. Metronidazole for vaginal Trichomoniasis: Seven day vs. single dose regimens. JAMA. 1980; 244: 1219.
- 17. Osoba AO. Epidemiology of urethritis in Ibadan. *Brit. J. Vener. Dis.* 1972; **48**: 116-120.

- 18. Alausa O, Osoba AO. The role of STD in male infertility in Tropical Africa. Nig. Med. J. 1978; 3:225-29.
- Cowan ST, Steel KJ. Manual for the identification of medical bacteria. 2nd edition. Cowan. Cambridge Univ. Press. 1974.
- 20. Adebayo JA. Isolation of Trichomonas vaginalis: a simple diagnostic medium for use in developing countries. Med. Lab. Sc. 1986; 43: 91-92.
- 21. Sorvoillo F, Smith L, Kerndt P, Ash L. Trichomonas vaginalis, HIV and African-Americans: Synopsis. Emerg. Infect. Dis. 2001; 7(6): 927.
- Wilkinson D, Abdool Karim 22. Harrison A. SS. et al. Unrecognized sexually transmitted infections South African rural women: a hidden epidemic. World Health Bull Organization. 1999; 77: 22-8.

- 23. Bakare RA, Oni AA, Umar US, et al. Prevalence of Trichomonas vaginalis among Commercial Sex Workers (CSWs) in Ibadan, Nigeria. Afr. J. Clin. Exp. Microbiol. 2002; 3(2): 72-77.
- 24. Bakare RA, Oni AA, Umar US, et al. Penicillinase producing Neisseria gonorrhoeae-the review of the present situation in Ibadan, Nigeria. Nig. Postgrad. Med. J. 2002; 9(2): 59-62.
- 25. Tanyuksel M, Gun H,
 Doganci L. Prevalence of
 Trichomonas vaginalis in
 prostitutes in Turkey.
 Centr. Eur. J. Publ. Health.
 1996; 4(2): 96-7

- 26. Fisher I, Morton RS. Epididymitis due to Trichomonas vaginalis. Br. J. Vener. Dis. 1969; 45: 252.
- 27. Catteral RD. Dignosis and treatment of Trichomonal urethritis in men. *Brit. Med.*J. 1960; 2:113.
- 28. Amar AD. Probable Trichomonas vaginalis epididymitis. JAMA. 1967; 200: 417.
- Ogunbanjo BO, Osoba AO,
 Oshei J. Infective factor of male infertility among
 Nigerians. Afr. J. Med. Sci. 1989; 18: 35-38.
- 30. Omigbodun OA, Bakare RA, Thomas JO. Detection of Trichomonas vaginalis by the papaniculaou smear. Trop. J. Obstetr. Gynaecol. 1993; 10(1): 37-39.

BACTERIOLOGICAL AND PARASITOLOGICAL ASSESSMENT OF VAGINITIS IN PREGNANT WOMEN IN ISEYIN, OYO STATE, NIGERIA.

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Specimens of High Vaginal Swabs (HVS) of 135 pregnant women were examined to determine the cause of vaginitis in pregnant women in Iseyin, Oyo State, Nigeria between August and October 1999. Study subjects were selected from patient attending selected antenatal clinics in public, private and mission hospitals/clinics in Iseyin. Samples were collected from subject in lithotomy position using sterile cuscos bivalve speculum. Sample were analysed by using standard technique as described. A structured questionnaire was also administered in order to obtain vital epidemiological information necessary for the study as described. The data analysis was done using chi square test. Results shows that 45 (33.3%) were positive for Candida spp, 15 (11.1%) for Gardnerella vaginalis and 5 (3.7%) for Trichomonas vaginalis. Sexual activities of individual have no significant effect on prevalence of vaginitis while symptomatology was a major indicator of infection. The effect of educational attainment and religion on infection rate was discussed. Infection decreased with age of patients while infection distribution by age of pregnancy gave a confusing pattern and the factors responsible for this were discussed. Since vaginitis could be asymptomatic most time, the screening of all pregnant women with risk factors for preterm labour and premature rupture of membranes must be undertaken. Prompt treatment of cases is also recommended.

Key Words: Pregnant women, vaginitis, aetiologic agents, prevalence, Iseyin.

INTRODUCTION

Vaginitis is inflammation of the vaginal, a disease entity first described by Willinson Abnormal vaginal discharge and related symptoms are frequent complaints in patients attending obsterics and gynaecology clinics some form of and vaginitis inducing organism, such Candida albicans or Bacterial Vaginosis (BV) are commonly diagnosed. In acute vaginitis, the squamous epithelial lining of the is invaded and vaginal wall inflammed causing discomfort, pruritus, or pain in addition to discharge. There is no doubt that

the microorganisms associated with cases of vaginitis frequently are isolated from asymptomatic women (2).

During pregnancy, the vaginal is more susceptible to infection, resulting in a higher incidence of colonization sypmptomatic vaginitis. The clinical attack rate is increased maximally during the third trimester and symptomatic recurrence is more common. Vaginitis has been observed in 61-90% of pregnant vaginal carriers as shown by Carrel et al (3).

Bacterial vaginosis (BV) is an entity receiving extensive attention

in pregnancy and needs greater focus on the part of the clinicians. It is found in 15 to 23% of pregnant women with up to 50% of patients being asymptomatic (4,5,6.). The vulnerability women to these pregnant result of infections is as \mathbf{a} of increased levels oestrogen during pregnancy, which creates a climate for the growth of these agents (7).

Whereas so much work has been done on vaginitis and its aetiologic agents in other parts of world, because \mathbf{of} importance of the subject, little or no work has been reported in Nigeria. Therefore this study is aimed at determining prevalence of the aetiologic agents among of vaginitis pregnant women in Iseyin, Nigeria with a view to having a data base for effective planning case management and control of the problem.

MATERIALS AND METHODS Study area

The study was conducted in Iseyin in Iseyin Local Government Area of Oyo State between August and October 1999. Iseyin, with a population of about 79,838 (national census 1991) predominantly Yoruba, is located on the longitude 4°15E and latitude 8°N, about 98 kilometers North of Ibadan, the Oyo State

1.1

capital. Their main stay of economy is farming and trading with a reputation for the production of native fabrics called 'Aso Oke'. The town is endowed with many educational and health institutions both at primary and secondary levels. It is a community of mixed religions.

Study subjects

For the purpose of the study and sample collection, the hospitals and health institutions in the study area were grouped into mission, public and private. The hospitals from each group were then randomly selected by balloting using lottery method. One hospital from each category was selected. The chosen hospitals for the study were: Our lady catholic hospital (mission). central maternity centre (public) and faith foundation hospital (private) respectively. The target population was pregnant women in Iseyin and those included in the study were those with symptom of vaginitis as recommended by Thomason et al Most of the (8).women presented themselves in the hospital were in the third trimester.

Sample collection

One hundred and thirty five high vaginal swab samples were collected from pregnant women attending the antenatal clinic in the three hospitals enlisted in the study. Non- pregnant women on postnatal visit were included in the

study as controls. Specimens were collect d with the assistance of clinical staff, with patents in lithotomy position using sterile bivalve speculum. coscos questionnaire structured was used to obtain information such as age, educational level, trimester, religion, occupation, marital status, number of sexual partners, symptoms and other important questions about their social life.

Sample analysis

Cultures were done modified Brain Heart Haeme agar, a multipurpose medium by using the conventional rolling method and incubated anaerobically at 37°C for 48 hours. All isolates were identified using the method described by Cruickshank et al (9) characterized using and criteria of Cowan and Steel (10). Wet preparation mount isolation of protozoa was done. slide was prepared suspending the discharge on the swab in a drop or two saline previously placed on the glass and examined microscopically using x10 and x40 objectives to scan at least ten fields for protozoa, Trichomonas vaginalis and yeast cells.

Data analysis

Data were analysed statistically using the Chi-square test to determine the significance of some variable where applicable; a p value of <0.05 was considered significant.

RESULTS

Table 1 shows the distribution of isolates from pregnant women in Iseyin. Of 135 subjects examined, 45(33.3%) were positive for Candida albicans, 15 (11.1%) for Gardnerella vaqinalis and 5(3.7%) Trichomonas vaginalis. Data revealed that infection rate with these agents differ significantly with age of the patients (p<0.05). Generally. there were mixed infections with some other bacterial Table 2 shows agents. distribution of infective agents by number of pregnancies. Result reveals that infection decreases with number of pregnancies. There was a significant difference when rates were analysed using the chisquare test (p<0.05). Fig 1 shows infection rate by sexual activities. There was no significant difference in infection rate between subjects with single and those with multiple sexual partners (p>0.05). Also women who had intercourse during pregnancy were more infected than those who did not. However, the difference was not statistically significant (p>0.05). Table 3 shows infection rate by trimester. Data revealed that the age of pregnancy infection did not affect rate distribution (p>0.05). Table 4 shows distribution by symptomatology and actiological agents. There were higher infection rates among

of with symptoms women discharge and itching than those without any symptoms. The chisquare test revealed significant infection rate difference in between patients that presented with symptoms and the control without group any symptom (p<0.05).Table 5 shows distribution by marital status. Infection rates were higher among the married and polygamous than other groups. Statistically however, the difference was not significant (p>0.05).distribution of agents of vaginitis by educational level is shown in Table 6. Marginally there is a decrease in infection rate with educational level, and there was a zero percent infectivity of all the agents of vaginitis among women

with tertiary education. However, the difference was not statistically significant (p>0.05). The result of the distribution of infective agents by occupation is presented in Table 7. There is a variation in prevalence no definite pattern. The statistical analysis revealed that there was no significant difference in the infection occurrence rate between the occupational groups (p>0.05). Table 8 shows distribution of infection by religion. There was 48% infectivity among muslims and 34% infectivity among christians. The Chi-square test revealed no significant difference statistical between the two (p>0.05).

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Table 1: Distribution of agents of vaginitis and other bacterial isolates by age.

		Isolate distribution						
	Number Examined	Candida spp %	G. vaginalis	T.vaginalis	Sirept. \$pp.	Kieb spp	E.coli	Staph app
≤20	30	11(36.7%)	2 (6.7%)	0 (0)	0 (0)	9(30%)	5(16.7%)	25(83.3%)
21-25	55	17(30.9%)	8(14.6)	2(3.63%)	5(9.1%)	13(23,6%)	11(20%)	43(78.2)
26-30	38	13(34.2)	13(10.5)	3(7.89%)	2(5.3%)	7(18.4%)	10(26.3%)	30(78.9%)
31-35	10	3(30.%)	1(10%)	0 (0)	0 (0)	2(20%)	3(30%)	9(90%)
36 & above	2	1(80%)	0 (0)	0 (0)	0 (0)	2(100%)	0(0%)	2(100%)
Total	135	45(33.3%)	15(11.1%)	5(3.7%)	7(5.2%)	33(24.4%)	29(21.5%)	109(60.7%)

P<0.05

Table 2: Distribution of infection by number of pregnancy

No of	Total No examined	Candida spp	G.vaginalis	T. vaginalis	Total No infected
pregnancy	(%)	1			
1	45(33.3%)	15(33.3%)	4(8.9%)	1(2.2%)	20(44.4%)
2.	41(30.4%)	16(39.9%)	4(9.8%)	1(2.4%)	21(51.2%)
3.	16(11,9%)	5(31.3%)	1(6.3%)	0 (0)	6(37.5%)
≥4	15(11.1%)	3(20%)	4(26.7%)	0 (0)	7(48.7%)

P>0.05

Table 2: Distribution by Trimester

¥5) •2·	Trimester	Total No varnined (%)	Candida spp	G.vaginalis	T. vaginalis	Total No infected
	First	4(2.9%)	3(75%)	0 (0)	0 (0)	3(75%)
1 34	Second	29(21.5%)	6(20.7%)	3(10.3%)	1(3.4%)	10(34.5%)
	Third	102(75.5%)	36(35.3%)	12(11.8%)	4(3.9%)	52(50.9%)
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P>0.05

Table 4: Distribution by symptomatology and aetiologic agents

Symptoms	Total No examined = 135(%)	Candida (%)	G.vaginalis (%)	T. vaginalis (%)	Total No infected (%)
Discharge	127(94.1%)	44(34.6%)	12(9.4%)	5(3.9%)	61(48.0%)
No discharge	8(5.9%)	1(12.5%)	3(32.5%)	0(0)	4(50%)
Itching	57(42.2%)	25(43.9)	3(5.3%)	5(8.8%)	33(57.9%)
No itching	78(57.8)	20(25.6%)	12(15.4%)	0(0)	32(41.1%)
Itching+ discharge	57(42.2%)	27(47.4%)	3(5.3%)	5(88%)	35(61.4%)
Discharge + no itching	70(51.9)	18(25.7%)	10(14.3%)	0 (0)	28(40%)
Control (Neg)	8(5.9)	1(12.5%)	3(37.5%)	0 (0)	4(50%)

P>0.05

Table 5: Distribution of agents of vaginitis by marital status

Marital Status	Total No examined (%)	Candida spp (%)	G.vaginalis (%)	T. vaginalis (%)	Total No infected
Married & polygamous	49(36.3%)	20(40.8%)	7(14.3%)	0(0)	27(55%)
Married & monogamous	58(42.9%)	17(29.3%)	4(6.9%)	1(3.4%)	22(37.9%)
. Single	28(20.7.5%)	8(28.6%)	4(14.3%)	0(0)	12(42.9%)

P>0.05

Table 6: Distribution of agents of vaginitis by educational level

Educational	Total No	Candida	G.vaginalis	T. vaginalis	Total No infected
level	examined (%)	(%)	(%)	(%)	(%)
Non-formal	37(27.4%)	12(32.4%)	6(16:2%)	2(5.4%)	20(54.1%)
Primary	54(40%)	12(38.9%)	2(3.7%)	2(3.7%)	25(46.3%)
Secondary	41(30.4%)	12(29.3%)	7(17.1%)	1(2.4%)	20(48.8%)
Tertiary	3(2.2%)	0 (0)	0 (0)	0 (0)	0 (0)

P>0.05

Table 7: Distribution of agents of vaginitis by occupation

Occupation	Total No examined (%)	Candida (%)	G.vaginali s (%)	T. vaginalis (%)	Total No infected (%)
Trader	97(71.9%)	35(36.1%)	12(12.4%)	5(5.15%)	52(53.6%)
Civil servant	5(3.7%)	1(20%)	1(20%)	0(0)	2(40%)
Farmer	5(3.7%)	1(20%)	0(0)	0(0)	1(20%)
Artisan	24(17.8%)	7(29.2%)	1(4.2%)	0 (0)	8(33.3%)
Housewife	4(2.9%)	1(25%)	2(25%)	0 (0)	2(50%)

P>0.05

Table 8: Distribution of agents of vaginitis by Religion

Religion	Total No	Candida	G.vaginalis	T. vaginalis	Total No
	examined	spp			infected
Christian	23(17.0%)	6(26.1%)	2(8.7%	0 (0)	8(34.8%)
Muslim	112(82.9%)	39(11.6%)	13(11.6%)	5(4.5%)	57(48.2%)

P>0.05

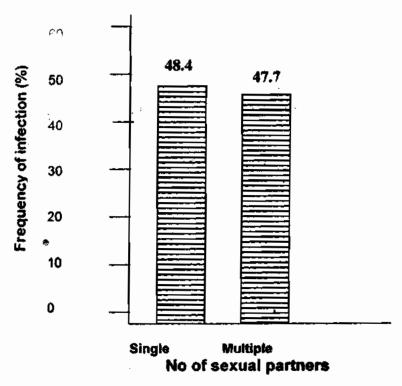


Figure 2a: Infection rate by sexual activities (Number of sexual partners)

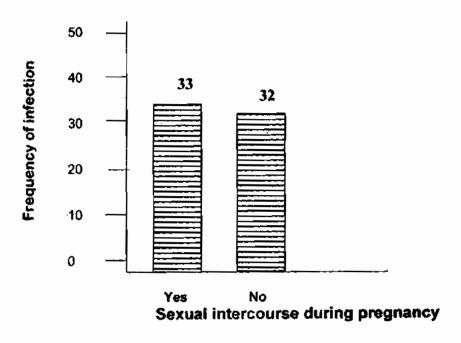


Figure 2b: Infection rate by sexual activities (intercourse during pregnancy)

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DISCUSSION

The study of microorganism incriminated as aetiologic agents of vaginitis serves to emphasis this complicated interaction that may these exist between the host. microorganisms and have that Studies shown organisms associated with vaginitis are part of the host's own microflora or exogenous microorganisms that must interact with species present as part of the host's indigenous flora (2). It has been shown in this study that the main organisms incriminated with vaginitis in pregnancy Trichomonas vaginalis, Candida Gardnerella albicans and This confirms vaginalis. the observation of some authors (10). Candida albicans with occurrence rate of 33.3% in Isevin shown to be the been commonest cause of vaginitis among the pregnant women in the report, which is in area. а consonance with the 30-40% prevalence reported by Hurley (12). This study has shown that G. vaginalis with 11.1% is second to Candida albicans in prevalence. Although this figure does not fall into the range (15.2%) reported by Amsel et al (4), there is agreement position the parasitic in the infection occupies as the second. most prevalent cause of vaginitis in pregnant women. Our report that Trichomonas vaginalis occurs

less frequently is not controverted by other workers (13,14). Mixed infections have remained a common occurrence in the study area. This is in conformity with the report of Lossick (14) who also agree that infection with these agents differ significantly with age of the patients (p<0.05). Garber et al (15) also agree that there is an increased incidence of Trichomenas vaginalis in pregnancy. This study has shown that the occurrence rate of infective agents shows no definite with the pattern number pregnancy (p>0.05). This may not be unconnected with the level of health education among populace. The subjects may have learnt from experience the need for prompt treatment as symptoms of infection manifest.

Result in this study has shown that age of pregnancy has no significant relevance in the distribution of infection (p>0.05).This at variance with the reports of Monton et al (16) and Hopsu-Havu (7), who showed that symptomatic disease is developed in about 10% of women during the first trimester and 36-55% during the third trimester. This disagreement in the report may be as a result of the attitude of the patients in our study area. Most of them usually attend clinic only during the third trimester. This has brought about the imbalance in the numbers in the respective stages of pregna ey, which may have accounted for the inference.

It has been shown in this study that sexual activities of the subjects have no significant impact on the level of infection. There was no significant difference in the level of infection between women who had single sexual partner and those with multiple partners. This is an aberration from the belief that promiscuity and / or unfaithfulness premised infidelity are а major indemiological factor of disease diffusion. The fact of the case may be due in part to regular case treatment, or those partners do not operate outside the regulars who may be bacteriologically clean. It was also observed in this study that sexual intercourse pregnancy does during necessarily promote infection, as there is no statistical difference in the infection rate between those pregnant women who claimed to sexual in intercourse engage during gestation, and those who denied. Therefore, it could be safe to assume that when sexual activity is restricted faithfully to one partner (that is zero grazing) the health of the individual is not thrown into jeopardy. However, the incidence of infection was significantly higher in women married into polygamous setting monogamous those in than homes. On the contrary the data in Table 5 revealed that there is no difference significant between married women and the single pregnant women. This revelation presents a paradoxical picture as opposed to the general belief that sexual promiscuity among partners do expose them more to risk of infection (14).Besides. patients are at greater risk of contacting HIV. This is a subject of future investigation.

study has shown that symptomatology and infection rates are linearly related. There were higher infection rates among subjects with vaginal itching and discharge than those without these symptoms. The factors responsible had been enunciated by many workers (15, 17) who incriminated surface protease action on mucous membrane of the vaginal in cases of Trichonomoniasis. It has been shown that the level of education of women has effect on the incidence of vaginitis in the study area. The uneducated and illiterate were mostly infected. Although the infection rate decreases with the level of education, the difference was not statistically significant (p>0.05). This study also showed that religious belief and occupation had no effect on the distribution of . study infection among the population (p>0.05).

Whereas the infection could be either symptomatic or asymptomatic, many workers

(11,12,18,19) had documented the pathogenesis and sequelae of the infection. The disease could lead rupture premature membranes in pregnancy, preterm labour and procure abortion. The infection may predispose the patients to HIV infection and other opportunistic infections with grave consequences. Therefore, it is recommended that pregnant women must register in the antenatal clinic within the first trimester for adequate and timely management. It is of paramount importance for the pregnant undergo periodic women to screening in order to forestall prelabour and premature rupture of membranes.

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REFERENCES

- 1. Olner-Hanssen O. Clinical manifestation of vaginal Ttrichomoniasis. *JAMA*. 1989; **264**: 471-576.
- Krieger JN, Tam MR, Steven CE. Diagnosis of Trichomoniasis. Comparision of conventional wet mount

- examination with cytologic studies, cultures and monoclonal antibody staining of direct specimens. *JAMA*. 1988; **259**: 1223-1227.
- Carrel CJ. Criteria for diagnosis of vulvovaginitis in pregnant women. J.Obstet. Gynaecol. Br. Com. 1973; 80: 258.
- Amsel R, Totten PA, Spiegel CA. Non-specific vaginitis: diagnostic criteria and microbial and epidemiologic associations. Am. J. Med. 1983. 74: 14-22.
- Gravelt M C, Nelson HP, De-5. Roven Т. Independent BVassociation and of Chlamydia trachomatis infection with adverse pregnancy outcome. JAMA. 1986; **226**: 1899-1903.
- 6. Hill LV, Luther ER, Yung JD. Prevalence of lower genital tract infections in pregnancy. Sex. Transm. Dis. 1988; 15: 5-10.
- Hopsu-Havu KK. Vaginal in parturition and infestation of newborns. Acta. Obstet. Gynaecol. Scand. 1980; 59: 73.
- 8. Thomason JL, Gelbart SM, Anderson RJ. Statistical evaluation of diagnostic criteria for BV. Am. J. Obstet. Gynaecol. 1990; 162: 155-160.

- Cruickshank R, Duguid JP, Marmion BP, Swain RHA. In: Medical Microbiology. The Practice of Medical Microbiology 12th edition. Churchill Livingstone. 1975.
- Cowan ST, Steel J. Manual for the identification of the medical bacteria 2nd edition Cambridge University Press. 1975: 45-114
- 11. Spiegel CA, Amsel RKK.
 Diagnosis of BV by direct
 Gram stain of vaginal fluid. J.
 Clin Gynecol. 1981; 8: 209.
- Hurley R. Recurrent candida infection. Clin. Obstet. Gynecol. 1981; 8: 209.
- 13. Rein MF, Muller M. Trichomonas vaginalis and Trichomoniasis. In: Holmes KK, Mardt PA, Sparling AF et al (eds). Sexually Transmitted Diseases. McGraw Hill, New York. 1989: 481-492.
- 14. Lossick JG. Epidermiology of urogenital Trichomonias. In: Honinbery BM (ed). Trichomonads parasitic in humans. Springen Verlag, New York. 1989: 311-323.
- 15. Garber G E, Lemchuk-Fawer LT, Bowie WR. Isolation of a cell-detaching factor of Trichomonas vaginalis J. Clin. Microbiol. 1989; 27: 1548.

400 800

- 16. Morton RS, Rashid M. Candidal vaginitis; natural history, predisposing factors and prevention. Proceeding of Royal Society of Medicine 1977: 70-73.
- 17. Arroyo R, Alderete JF. Trichonmonas vaginalis surface protein activity is necessary for parasitic adherence to epithelial cells. Infect. Immum. 1989; 57: 2991-2997.
- 18. Rain MF. Clinical manifestations of urogenital Trichomoniasis in women. In: Honinbery (ed). BM Trichomonads parasitic in humans. Spring Hill, New York. 1990; 225-234.
- 19. Douglas IJ. Mannoprotein adhesive of Candida albicans. Bennet JE, Hsv RJ. In: Retuson PΚ (eds). New Strategies in fungal diseases: Livingstone. Churchill Edinburgh.

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